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# A Chiral LC-MS Strategy for Stereochemical Assignment of Natural Products Sharing a 3-Methylpent-4-en-2-ol Moiety in Their Terminal Structures

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#### Abstract

A terminal 3-methylpent-4-en-2-ol (MPO) moiety is a common structural feature in various polyketide natural products. Stereochemical assignments of this moiety have mainly relied on computational analyses of NMR, ECD, and specific rotation data. However, neither of these approaches can be applied to all compounds. In this study, we developed a method to determine the absolute configuration of the terminal MPO moiety with high accuracy and sensitivity by a combination of chemical degradation, chemical synthesis, and chiral LC-MS analysis. The applicability of this method was demonstrated through the stereochemical assignment of (+)-capsulactone (**1**).

### Keywords

3-methylpent-4-en-2-ol moiety; *p*-nitrobenzoyl ester; methyl-3-hydroxy-2methylbutanoate; chemical degradation; chiral LC-MS analysis

## Introduction

Configurational elucidation of natural products is essential for progress in diverse research areas, including the development of synthetic methodologies, the investigation of modes of action, and the evaluation of pharmacological potential, among others. Advancements in spectroscopic techniques, such as NMR and MS, have enabled the structural elucidation of complex natural products at microgram quantities, facilitating the discovery of novel bioactive compounds [1,2]. However, a definitive approach for stereochemical determination that can be applied across all compound classes has yet to be established. Relative configurations are typically

determined using NMR-based techniques, such as coupling constant analysis and NOE experiments, sometimes with the aid of computational chemistry [3-7]. In contrast, absolute configuration remains more challenging to determine, as it frequently requires chemical degradation or derivatization. Several derivatization anisotropic methods using chiral reagents, including α-methoxy-αtrifluoromethylphenylacetic acid (MTPA), phenylglycine methyl ester (PGME), and Marfey's reagents, are widely used [8-10], although their applicability is restricted by the presence of specific functional groups. Total synthesis is a powerful approach for determining absolute configuration through the comparison of specific rotation or chromatographic behavior; however, it requires considerable time and effort. In this context, we have been working on developing effective approaches to determine the absolute configuration of scarce natural products, including heptavalinamide A [11] and poecillastrin C [12,13] by a combination of chemical degradation, chemical synthesis, and LC-MS analysis.

The 3-methylpent-4-en-2-ol (MPO) moiety is commonly found at the terminal position of various polyketide natural products such as a series of azaphilones including chaetomugilins [14], chaetoviridins [14,15], and some other  $\alpha$ -pyrone polyketides [16-18] (Figure 1). Among the available strategies for elucidating the stereochemistry of MPO, X-ray crystallographic analysis, and computational methods have been widely used. For example, the absolute configuration of chaetomugilin B [19] was determined by X-ray crystallography, while that of capsulactone (1) was established using computational techniques, including predictions of NMR chemical shifts and ECD spectra [17]. However, the absolute configurations of compounds such as linearolides [20], juniperolide A [21], certonardosterol A4 [22], and sclerketide D [23] remain unresolved due to the limited sample quantity or ambiguous results, even when the modified Mosher's method is employed; a medium vicinal coupling constant (4–6

3

Hz) prevents reliable differentiation between *threo* and *erythro* configurations of adjacent hydroxy group and methyl group (Figure 1) [24]. Therefore, a general and reliable chemical approach for stereochemical determination of the terminal MPO-containing compounds is required. Herein, we report the development of a method for determining the absolute configuration of the MPO moiety by LC-MS and demonstrate its application to the stereochemical assignment of capsulactone (1) at the microgram scale. The strategy involves the optical resolution of MPO derivatives, chemical degradation of 1, and the stereoselective synthesis of four diastereomers.



**Figure 1.** Representative natural products sharing MPO moiety in their terminal structures, with the moiety highlighted in blue (assigned configuration) or yellow (undetermined configuration).

## **Results and Discussion**

Our degradation strategy of natural products bearing MPO moiety includes (1) acylation of hydroxyl group, (2) oxidative cleavage of olefin to generate 3-hydroxy-2methylbutanoic acid, and (3) its methyl esterification (Scheme 1A). We initially investigated derivatization strategies to enable LC-MS detection of the MPO-derived fragment and to achieve a separation of four candidate stereoisomers. To this end, methylation of commercially available methyl acetoacetate (2) and subsequent reduction of the ketone carbonyl group was carried out to prepare a mixture of four stereoisomers of methyl 3-hydroxy-2-methylbutanoate (3) [25,26]. We then explored LC-MS conditions for their separation. In our initial trial, the stereoisomeric mixture of 3 was successfully separated by preparative HPLC using a chiral column (data not shown). However, each stereoisomer obtained suffered from co-evaporation during solvent removal under reduced pressure, which led us to consider that degradation of the natural product to obtain the corresponding fragment would be challenging. Accordingly, esterification of the hydroxyl group at C3 in 3, and suitable acyl groups were then investigated. First, alcohol 3 was converted to (R)-MTPA ester 4 (Scheme 1B). Despite numerous attempts to optimize the chromatographic conditions, the resulting diastereomers could not be separated (Figure S1). Next, *p*-bromobenzoyl ester 5 was synthesized (Scheme 1B). Although complete separation of the stereoisomers was not achieved, the corresponding peaks exhibited improved resolution compared to those of 4 (Figure S2). To further enhance separation, pnitrobenzoyl (PNB) ester 6 was prepared (Scheme 1B). After repeated trials with several chiral HPLC columns and with various elution profiles, the four stereoisomers of 6 were successfully separated on a CHIRALPAK ® ID-3 column at 40 °C with aqueous MeOH gradient elution (Figure S3).



**Scheme 1.** (A) General strategy for the preparation of the fragment from an MPOcontaining natural product. (B) Synthesis of esters **4–6**, each as a mixture of four stereoisomers.

The developed method for absolute configuration assignment of MPO was applied to the natural product capsulactone (**1**), an  $\alpha$ -pyrone polyketide first isolated in 2021 from the endophytic fungus *Penicillium capsulatum* [17], and re-isolated in 2023 from the endophytic fungus *Neurospora dictyophora* WZ-497 [18], whose absolute configuration had been determined by computational methods. We had also independently re-isolated **1** (1.2 mg) from the fungus *Fusarium* sp. (Supporting Information) and subsequently subjected 100 µg of **1** to the sequential degradation steps; (1) *p*-nitrobenzoylation of the two hydroxyl groups, (2) RuO<sub>4</sub> oxidation to cleave olefins including that at C8–C9, and (3) methyl esterification (Scheme 2). The resulting C9–C12 fragment **7** was successfully detected at *m/z* 304.1 [M+Na]<sup>+</sup> on LC-MS, as expected (see Figure 2b for the chromatogram).



Scheme 2. Preparation of the C9–C12 fragment (7) from capsulactone (1).

We then proceeded with the stereoselective synthesis of four diastereomers: 4-methoxy-3-methyl-4-oxobutan-2-yl 4-nitrobenzoates (2S,3S)-8, (2S,3R)-9, (2R,3R)-10, and (2R,3S)-11, as outlined in Scheme 3. The synthesis of (2S,3S)-8, and (2S,3R)-9 started from commercially available (3S)-12, and stereogenic center at C2 was constructed via stereoselective methylation [27] to afford (2S,3S)-13. Due to the volatility of **13**, the crude solution obtained after the workup (quenching and phase separation) was directly used in the subsequent reaction without purification. Esterification of (2S,3S)-13 with p-nitrobenzoyl chloride (PNBCI) afforded PNB ester (2S,3S)-8 in 45% yield for two steps. Alternatively, PNB ester (2S,3R)-9, the C3-epimer of **8**, was synthesized from (2S,3S)-**13** via a Mitsunobu reaction using *p*-nitrobenzoic acid (PNBOH) and DEAD in 39% yield for two steps [28]. The stereoisomers (2R,3R)-**10**, and (2*R*,3*S*)-**11** were prepared in the same manner starting from commercially available (3*R*)-14, in acceptable yields of 41% and 38% for two steps, respectively. The four stereoisomers 8–11 were separated by LC-MS with the following retention times: (2S,3S)-8 (33.4 min), (2S,3R)-9 (33.5 min), (2R,3R)-10 (33.9 min), (2R,3S)-11 (34.7 min) (Figure 2a). The LC-MS analysis of fragment 7, derived from natural 1, showed a retention time identical to that of (2R,3S)-11 (Figure 2b), thus assigning the absolute configuration of the MPO moiety in **1** as (10*R*,11*R*) [29]. This assignment was consistent with prior reports, and the observed specific rotation of 1, measured as  $[\alpha]_D^{26}$ +12 (c 0.1, MeOH) closely matched the reported values;  $[\alpha]_D^{21}$  +12 (c 0.1, MeOH) [17], and [α]<sub>D</sub><sup>20</sup> +12.8 (*c* 0.24, MeOH) [18].

Recently, the absolute configuration of MPO moiety in ouhanamide [30] (Figure 1), isolated from cyanobacteria, was determined by a combination of chemical degradation and chemical synthesis. In that study, separation of four synthetic stereoisomers of 3-((4-bromobenzoyl)oxy)-2-methylbutanoic acid on chiral HPLC (CHIRALPAK ® IA column with hexane/EtOH/TFA isocratic elution) was achieved. A key advantage of our method is the use of a PNB group as the acyl moiety, which enables direct conversion to a C3-epimer via Mitsunobu reaction, thereby streamlining the synthesis of the four stereoisomers. Additionally, the reversed-phase HPLC conditions without the addition of TFA in the mobile phase can be applied to LC-MS, enabling highly sensitive detection, thereby facilitating reliable stereochemical assignment using only a minimal amount (less than a microgram) of a natural product derivative.



**Scheme 3.** Synthesis of 4-methoxy-3-methyl-4-oxobutan-2-yl 4-nitrobenzoates (2*S*,3*S*)-8, (2*S*,3*R*)-9, (2*R*,3*R*)-10, and (2*R*,3*S*)-11.



**Figure 2.** LC-MS chromatograms of 4-methoxy-3-methyl-4-oxobutan-2-yl 4nitrobenzoates. Conditions: CHIRALPAK<sup>®</sup> ID-3 (4.6 × 250 mm, 3  $\mu$ m) at a flow late of 0.6 mL/min at 40 °C, with gradient elution from 50% MeOH to 100% MeOH. (a) Synthetic (2*S*,3*S*)-**8**, (2*S*,3*R*)-**9**, (2*R*,3*R*)-**10**, and (2*R*,3*S*)-**11**, (b) C9–C12 fragment **7** derived from **1**.

## Conclusion

In conclusion, we have developed a method for the stereochemical determination of the terminal MPO moiety with high accuracy and sensitivity, based on a combination of chemical degradation, efficient synthesis of all stereoisomers of the fragment, and LC-MS analysis using a chiral stationary phase. The applicability of this method was validated through the successful determination of the absolute configuration of capsulactone (1) using only 100  $\mu$ g of sample. This is the first report to establish the elution pattern of the four diastereomers of 3-hydroxy-2-methylbutanoic acid derivatives by LC-MS. Notably, the method developed herein enables the assignment of absolute configuration without the need for the individual synthesis of all four stereoisomers, as the use of stereomixture **6** is sufficient for the stereochemical assignment. This represents a significant advantage over previously reported chemical

methods [24, 30]. Our method would be applicable to a variety of natural products, such as linearomides, juniperolide A, certonardosterol A<sub>4</sub>, and sclerketide D, all of which share an undetermined MPO moiety in their terminal structures. Given the utility of this strategy, it holds promise as a valuable tool for the structural determination of diverse MPO-containing natural products in future studies.

# **Supporting Information**

Supporting Information File 1: Experimental data and NMR spectra.

## **Data Availability Statement**

All data that supports the findings of this study is available in the published article and/or the supporting information to this article.

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# **Conflict of Interest**

There are no conflicts to declare.

### **Author Contribution**

Rei Suo: Conceptualization, Funding acquisition, Investigation, Validation, Visualization, Writing- Original draft preparation; Raku Irie: Conceptualization, Investigation, Validation, Visualization, Writing- Reviewing and Editing.; Hinako Nakayama: Investigation; Yuta Ishimaru: Investigation; Yuya Akama: Investigation; Masato Oikawa: Writing- Reviewing and Editing; Shiro Itoi: Writing- Reviewing and Editing. All authors reviewed the manuscript and approved the final version to be published.

### References

- 1. Molinski, T. F.; Morinaka, B. I.; Tetrahedron 2012, 68, 9307–9343.
- 2. Marialuisa, M.; Concetta, I.; Alfonso, M.; Gerardo, D. S.; Orazio, T. S. *Nat. Prod. Rep.* **2019**, *36*, 476–489.
- 3. Matsumori, N.; Kaneno, D.; Murata, M.; Nakamura, H.; Tachibana, K. *J. Org. Chem.* **1999**, *64*, 866–876.
- 4. Higashibayashi, S.; Czechtizky, W.; Kobayashi, Y.; Kishi, Y. *J. Am. Chem. Soc.* **2003**, *125*, 14379–14393.
- 5. Petrovic A.G.; Navarro-Vázquez A., Alonso-Gómez J.L. *Curr. Org. Chem.* **2010**, *14*, 1612–1628.
- 6. Bertucci, C.; Tedesco, D. J. Chromatogr. A 2012, 1269, 69-81.
- 7. Harada, N.; Nakanishi, K. Acc. Chem. Res. 1972, 5, 257–263.
- 8. Dale, J. A.; Mosher, H. S. J. Am. Chem. Soc. 1973, 95, 512–519.
- 9. Kusumi, T.; Takahashi, H.; Xu, P.; Fukushima, T.; Asakawa, Y.; Hashimoto, T.; Kan,
- Y.; Inouye, Y. Tetrahedron Lett. 1994, 35, 4397–4400.
- 10. Marfey, P. Carlsberg Res. Commun. 1984, 49, 591-596.

11. Suo, R.; Watanabe, R.; Takada, K.; Suzuki, T.; Oikawa H.; Itoi, S.; Sugita, H.; Matsunaga, S. *Org. Lett.* **2020**, *22*, 1254–1258.

Irie, R.; Takada, K.; Ise, Y.; Ohtsuka, S.; Okada, S.; Gustafson, K. R.; Matsunaga,
 S. Org. Lett. 2017, 19, 5395–5397.

13. Irie, R.; Hitora, Y.; Watanabe, R.; Clark, H.; Suyama, Y.; Sekiya, S.; Suzuki, T.;

Takada, K.; Matsunaga, S. Hosokawa, S.; Oikawa, M. Org. Lett. 2024, 26, 5290–5294.

14. Gao, J.-M.; Yang, S.-X.; Qin, J.-C. A. Chem. Rev. 2013, 113, 4755–4811.

15. Chen, C.; Tao, H.; Chen, W.; Yang, B.; Zhou, X.; Luo, X.; Liu, Y. *RSC Adv.* **2020**, *10*, 10197–10220.

Ma, Y.; Wen, Y.; Cheng, H.; Deng, J.; Peng, Y.; Bahetejiang, Y.; Huang, H.; Wu,
 C.; Yang, X.; Pang, K. *Bioorg. Med. Chem. Lett.* **2021**, *40*, 127921.

17. Xian, P. J.; Chen, H. Y.; Feng, A.; Zhao, W.; Yang, X. L.C. *J. Asian. Nat. Prod. Res.* **2021**, 23, 1100–1106.

18. Li, X.; Gong, Y. -X.; Feng, L.; Wang, X. -J.; Wang, J. -W.; Zhang, A. -X.; Tan, N. H.; Wang, A. *Phytochemistry* **2023**, *207*, 113579.

19. Yamada, T.; Doi, M.; Shigeta, H.; Muroga, Y.; Hosoe, S.; Numata, A.; Tanaka, R. *Tetrahedron Lett.* **2008**, *49*, 4192–4195.

20. Ueki, M; Koshiro, N.; Aono, H.; Kawatani, M.; Uramoto, M.; Kawasaki, H.; Osada,
H. *J. Antibiot.* **2013**, 66, 333–337.

21. Raju, R.; Gromyko, O.; Fedorenko, V.; Luzhetskyy, A.; Plaza, A.; Müller, R. *Org. Lett.* **2012**, *14*, 5860–5863

22. Wang, W.; Jang, H.; Hong, J.; Lee, C.; Im, K. S.; Bae, S.; Jung, J. H. *J. Nat. Prod.* **2004**, *67*, 1654–1660.

23. Liu, Z.; Qiu, P.; Liu, H.; Li, J.; Shao, C.; Yan, T.; Cao, W.; She, Z. *Bioorg. Chem.* **2019**, *88*, 102973.

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24. The following reference describes a rare case of stereochemical determination of a terminal MPO moiety using *J*-based configurational analysis (JBCA): Neuhaus, G. F.; Adpressa, D. A.; Bruhn, T.; Loesgen, S. *J. Nat. Prod.* **2019**, *82*, 2780–2789.

25.Trotta, A. H. Org. Lett. 2015, 17, 3358-3361.

- 26. Kalaitzakis, D. Kambourakis, S. Rozzell, D.J Smonou, J. *Tetrahedron: Asymmetry* **2007**, *18*, 2418–2426.
- 27. Jacolot, M.; Jean, M.; Levoin, N.; van de Weghe, P. Org. Lett. 2012, 14, 58-61.
- 28. Mitsunobu, O. Synthesis 1981, 1981, 1–28.
- 29. Note that Cahn–Ingold–Prelog priorities at C2 in **8–11** are reversed from that at C10 in **1**; i.e., if the configuration in the degradation fragment **7** is found to be (10*S*), then the absolute configuration at C10 in **1** is (10*R*).
- 30. Niiyama, M.; Kurisawa, N.; Umeda, K.; Jeelani, G.; Agusta, A. L.; Nozaki, T.; Suenaga, K. *J. Nat. Prod.* **2025**, Article ASAP (DOI: 10.1021/acs.jnatprod.5c00281).