

This open access document is published as a preprint in the Beilstein Archives with doi: 10.3762/bxiv.2020.93.v1 and is considered to be an early communication for feedback before peer review. Before citing this document, please check if a final, peer-reviewed version has been published in the Beilstein Journal of Organic Chemistry.

This document is not formatted, has not undergone copyediting or typesetting, and may contain errors, unsubstantiated scientific claims or preliminary data.

Preprint Title	Vicinal difluorination as a C=C surrogate: an analog of piperine with enhanced solubility, photostability, and acetylcholinesterase inhibitory activity
Authors	Yuvixza Lizarme-Salas, Alexandra D. Ariawan, Ranjala Ratnayake, Hendrik Luesch, Angela Finch and Luke Hunter
Publication Date	12 Aug 2020
Article Type	Full Research Paper
Supporting Information File 1	SI 6.8.20.pdf; 3.0 MB
ORCID [®] iDs	Yuvixza Lizarme-Salas - https://orcid.org/0000-0001-6964-8129; Ranjala Ratnayake - https://orcid.org/0000-0002-5980-2070; Angela Finch - https://orcid.org/0000-0002-4598-6745; Luke Hunter - https://orcid.org/0000-0002-8678-3602

This is an open access publication under the terms of the Creative Commons Attribution License (<u>http://creativecommons.org/licenses/by/4.0</u>). Please note that the reuse, redistribution and reproduction in particular requires that the author(s) and source are credited.

The license is subject to the Beilstein Archives terms and conditions: https://www.beilstein-archives.org/xiv/terms.

The definitive version of this work can be found at: doi: https://doi.org/10.3762/bxiv.2020.93.v1

Vicinal difluorination as a C=C surrogate: an analog of piperine with enhanced solubility, photostability, and acetylcholinesterase inhibitory activity

Yuvixza Lizarme-Salas,^{*a*} Alexandra Daryl Ariawan,^{*a*} Ranjala Ratnayake,^{*b*} Hendrik Luesch,^{*b*} Angela Finch,^{*c*} Luke Hunter^{*a*},*

^a School of Chemistry, University of New South Wales (UNSW), Sydney NSW 2052, Australia
 ^b Department of Medicinal Chemistry and Center for Natural Products, Drug Discovery and Development (CNPD3), University of Florida, Gainesville FL 32610, United States
 ^c School of Medical Sciences, University of New South Wales (UNSW), Sydney NSW 2052, Australia
 * l.hunter@unsw.edu.au

ABSTRACT

Piperine, a natural product derived from peppercorns, has a variety of biological activities that make it an attractive lead compound for medicinal chemistry. However, piperine has some problematic physicochemical properties including poor aqueous solubility and a susceptibility to UV-induced degradation. In this work, we designed an analog of piperine in which the central conjugated hydrocarbon chain is replaced with a vicinal difluoroalkane moiety. We show that this fluorinated analog of piperine has superior physicochemical properties, and it also has higher potency and selectivity towards one particular drug target, acetylcholinesterase. This work highlights the potential usefulness of the *threo*-difluoroalkane motif as a surrogate for *E*-alkenes in medicinal chemistry.

INTRODUCTION

Piperine (1, Figure 1) is a well-known natural product that is derived from peppercorns.^{1–3} Many biological studies of 1 have been carried out, and these studies have led to a diverse array of biological activities being claimed for this compound.^{4–9} For example, 1 is reported to exhibit inhibitory activity towards both acetylcholinesterase (AChE) and β -secretase (BACE-1), which suggests that 1 could hold promise as a dual mechanism-of-action treatment for Alzheimer's disease.^{4,10–13}



Figure 1: The natural product piperine (1) is the inspiration for this work; its crystal structure is shown.¹⁴ In this work, a hypothetical analog (2) was designed to mimic 1. Predicted low-energy rotamers of 2 about F–C–C–F and F–C–C=O are shown; rotamers I and IV give the best mimicry of 1.

However, piperine (1) has some limitations as a drug lead. For example, it has poor solubility, and it is susceptible to photoisomerization of the conjugated system.^{15–17} This prompted us to consider whether there could be another structural motif that could (i) mimic the extended geometry of an *E*-alkene; (ii) impart better solubility; and (iii) be more stable in the presence of UV light. Such a C=C surrogate might offer an improved lead compound for the development of drugs to treat Alzheimer's disease.

Stereoselective fluorination is an emerging strategy for controlling the conformations of organic molecules. The highly polarized C–F bond tends to align in predictable ways with adjacent functional groups, due to a combination of hyperconjugative and/or dipolar interactions.^{18–21} This knowledge led us to propose the hypothetical analog **2** (Figure 1) as a potential mimic of **1**. Analog **2** contains a saturated alkyl chain with two vicinal C–F bonds in the α/β positions relative to the amide moiety. The presence of a vicinal difluoride moiety within an alkyl chain is known to favor rotamers in which the C–F bonds align *gauche* (I–II, Figure 1).²² Separately, the presence of fluorine on the α -carbon of a tertiary amide is known to restrict the C α –C(O) bond to small set of low-energy rotamers (III–V, Figure 1).²³ Intriguingly, studies of simple molecules containing either F–C–C–F or F–C–C=O have shown that the rotamer populations can change, depending on the polarity of the solvent.^{22,23} The rotamers I and IV (Figure 1, boxed) would appear to provide the closest structural match with compound **1**, but in a highly polar medium the more polar rotamers II and V might predominate. Of course, the contiguous positioning of the F–C–C–F and F–C–C=O moieties within the same molecule

(2) means that these moieties would not function independently, but rather would likely influence one another, adding another layer of complexity. Finally, it is interesting to consider whether the microenvironment of a protein binding site could also change the relative energies of the various F–C–C–F and F–C–C=O rotamers, offering the possibility that analog 2 might be an effective conformational mimic of 1 in some environments but not in others.

Herein, we describe the optimisation of a synthetic route to 2; the conformational analysis of this molecule by NMR and molecular modelling studies; a comparison of the photostabilities of 1 vs. 2; and a comparison of the inhibitory activities of 1 vs. 2 towards both AChE and BACE-1.

RESULTS AND DISCUSSION

Synthesis

Jacobsen and co-workers have recently described a method for the one-step, diastereoselective 1,2difluorination of alkenes, mediated by a hypervalent iodine catalyst.²⁴ The substrate scope of Jacobsen's method has certain constraints, but their original report did include some examples of α , β unsaturated amides and so we were motivated to investigate the method in this work, using compound 3^{25} as the substrate (Scheme 1). We recognized that the product of this reaction would most likely be the unwanted *erythro*-isomer (6), but we nevertheless deemed it a worthwhile preliminary investigation. Disappointingly, treatment of the precursor 3 with pyridine.HF and either catalyst 4 or 5 at room temperature for two days gave no discernible reaction, and the starting material (3) was recovered intact. Warming the reaction mixture to 50 °C caused the disappearance of 3 but the formation of an intractable mixture of products. We were forced to conclude that the rather complex structure of 3 was incompatible with Jacobsen's catalytic system.



Scheme 1: Attempted synthesis of 6 (a diastereoisomer of 2) via a one-step 1,2-difluorination reaction.²⁴

Since the one-step difluorination method (Scheme 1) was unsuccessful, we decided to pursue a stepwise fluorination approach^{26,27} (Scheme 2). Thus, the allylic alcohol 7^{28} was protected as the benzyl ether then subjected to a Sharpless asymmetric dihydroxylation reaction to furnish the diol 8 in good yield. Diol 8 was then converted into the cyclic sulfate 9, which was ring-opened using TBAF to furnish the fluorohydrin 10 in modest yield. Mosher ester analysis of the fluorohydrin 10 suggested that the earlier dihydroxylation reaction had proceeded with 90% ee. The deoxyfluorination of 10 was then attempted using several reagents including DeoxoFluor; DeoxoFluor in combination with TMS-morpholine;²⁹ and PyFluor.³⁰ The optimal yield of the *threo*-difluoroalkane **11** was obtained with DeoxoFluor at elevated temperature (Scheme 2). A side-product in this fluorination reaction was the tricyclic compound 12, which presumably formed through an electrophilic aromatic substitution reaction of the activated alcohol intermediate. The inclusion of TMS-morpholine²⁹ reduced the formation of this side-product but did not lead to an overall increase in the yield of **11**. Despite the modest optimized yield of the difluoroalkane **11**, a sufficient quantity of this material was secured to continue with the synthesis. Hydrogenolysis of the benzyl ether of **11** provided the primary alcohol 13, which only needed to be oxidized to the carboxylic acid 14 then coupled to piperidine in order to deliver the target compound, 2. However, the oxidation of 13 proved to be unexpectedly troublesome. Mild oxidising agents caused incomplete consumption of 13, while vigorous conditions caused the oxidation not only of the primary alcohol of 13 but also of the electron-rich aryl moiety (see SI).



Scheme 2: Attempted synthesis of 2 via a stepwise fluorination approach (ether series).

In order to circumvent the troublesome oxidation reaction (*i.e.* $13\rightarrow14$, Scheme 2), and to seek higher enantiopurity of the target, we modified the synthetic plan to include an ester moiety throughout (Scheme 3). Thus, the α , β -unsaturated ester 15^{25} was carried through a similar sequence to that previously described, *i.e.* dihydroxylation; cyclic sulfate formation; ring-opening with TBAF (although note the regioselectivity); deoxyfluorination; and deprotection to deliver the difluorinated acid 20. Finally, coupling of 20 with piperidine afforded the target compound (2) in moderate yield (Scheme 3). The enantiopurity of 2 was investigated using chiral HPLC. From a spectroscopically pure sample of 2, two HPLC peaks were observed, with an integral ratio of 99:1. It is assumed that the two peaks correspond to the two enantiomers of 2, and if that assumption is true then the optical purity of this sample is 98% *ee*.



Scheme 3: Synthesis of 2 via a stepwise fluorination approach (ester series).

Conformational analysis

Having completed the synthesis of the target molecule (2), the next task was to investigate the conformational behavior of this molecule. This was achieved by performing a DFT study in the Gaussian software, using the M06-2X level of theory with the 6-311+G(d,p) basis set, parameters similar to those employed by Linclau and co-workers for their studies of vicinal difluoro systems.²² A set of starting structures of **2** were generated by systematically rotating three bonds (*i.e.* F–C–C–F; F–C–C=O; O=C–N–C) in 120° increments, and the starting structures were then geometry optimized and their energies calculated. To enable benchmarking against experiment, NMR spin-spin

coupling constants were calculated for the lowest-energy final structures of 2, using the GIAO method with B3LYP/6-311+G(d,p) level of theory. Chloroform was used as the solvent for both the NMR experiments and the SMD calculations.

The three lowest-energy structures to emerge from the computational analysis (*i.e.* **2a–c**) are shown in Figure 2. The global minimum structure (2a) has a F–C–C–F dihedral angle of 73°, giving an extended zigzag carbon chain. This approximates rotamer I (Figure 1), and in this regard 2a is a good conformational mimic of **1**. However, **2a** has a F–C–C=O dihedral angle of 152°; this approximates rotamer III (Figure 1) and in this regard 2a is a poorer conformational mimic of 1. The next-higher energy structure (2b, Figure 2) is related to 2a through a ring-flip of the piperidine moiety (equivalent to a 180° rotation of the amide bond). Structures **2a/b** are close in energy, suggesting that both puckers of the piperidine moiety would be substantially populated in solution. A similar situation occurs for piperine itself; indirect evidence for this comes from the crystal structure of **1** (Figure 1), where there is a second molecule in the unit cell (not shown) that has the alternative ring pucker.¹⁴ The nexthigher energy calculated structure of 2 (*i.e.* 2c, Figure 2) has a F–C–C–F dihedral angle of –52°; this approximates rotamer II (Figure 1), giving a bent carbon chain that contrasts with the extended chains of 2a/b. Structure 2c has a F–C–C=O dihedral angle of -118° ; this approximates rotamer IV (Figure 1). Together, structures 2a-c seem to dominate the conformer population distribution of 2 in chloroform solution, because the weight-averaged calculated J-values of 2a-c match the experimentally-measured *J*-values quite well (Figure 2). But none of **2a–c** are a close conformational match with piperine (1).



Figure 2: Conformational analysis of 2 by DFT and NMR. The numbering scheme for NMR spins is given on structure 2a.

A noteworthy feature of structures **2a/b** (Figure 2) is that both the α -fluorine and the β -fluorine atoms of each structure make close contacts with hydrogen atoms on the piperidine ring (2.08–2.34 Å). This manifests in the observation of a through-space coupling (J = 1.9 Hz) between the α -fluorine and a piperidine hydrogen in the experimental NMR spectrum of **2** (*i.e.* spins 6/7, Figure 2). The attractive F…H interactions might explain why the amide bond is twisted by 11–20° from planarity in **2a/b**. In structure **2c**, only the α -fluorine makes close contacts with hydrogens on the piperidine ring, and the amide is twisted to a lesser degree in that case (8° away from planarity). It is interesting that in the crystal structure of piperine itself (**1**, Figure 1),¹⁴ the amide bond is also twisted by 15° away from planarity; in this case the distortion might be attributable to a H…H clash (1.99 Å).

Structure **2f** (Figure 2), which was among the higher-energy calculated conformers of **2** (see Supporting Information), bears a close resemblance to the solid-state conformation of piperine (**1**, Figure 1). Structure **2f** has a F–C–C–F dihedral angle of 57° and a F–C–C=O dihedral angle of -133° ; these angles approximate rotamers I and IV respectively (Figure 1). The unexpectedly high relative energy that was calculated for **2f** might be partially attributable to the fact that it features only a single F…H contact within 2.50 Å. The calculations suggest that conformer **2f** is not significantly populated in chloroform solution, but it was not possible to verify this by NMR because **2f** is essentially superimposable with **2a/b** within the fluoroalkyl segment. In more polar solvents such as water, the

intramolecular F····H interactions would be expected to decrease in significance³¹ and this might increase the accessibility of conformer **2f**. We reasoned that comparing the biological activities of **1** *vs*. **2** might shed light on this possibility (*vide infra*).

Photostability

A limitation of piperine (1) as a drug lead is its instability under UV light. The conjugated system of 1 is well-known to undergo facile E/Z isomerization upon the absorption of UV photons, leading rapidly to a mixture of all four possible geometric isomers of 1.^{15–17} In the present work, this phenomenon was confirmed by exposing an ethanolic solution of 1 to sunlight for 2.5 h (Figure 3); analysis of the product by ¹H NMR spectrometry revealed a multitude of new signals in the alkenyl region. In contrast, analog 2 lacks conjugation in the central portion of the molecule, and was therefore expected to be more stable to UV light. Indeed, exposure of 2 to sunlight in an identical manner to that described for 1 led to no detectable decomposition (Figure 3, Supporting Information).



Figure 3: Analog 2 has greater stability to UV light than does piperine (1).

Biological activity

The biological activities of piperine (1) and the analog 2 were compared using two different assays, namely the inhibition of either acetylcholinesterase (AChE) or β -secretase (BACE-1).

Inhibition of AChE was measured using a modification of a previously described colorimetric assay (Figure 4a).³² It quickly became apparent that the limited solubility of piperine (**1**) in 50 mM Tris-HCl buffer, even in the presence of methanol co-solvent, was a major problem in this assay: we observed cloudiness or the appearance of a precipitate at higher concentrations of **1**, which affected the reproducibility of the assay and prevented complete inhibition from being achieved (Figure 4a).

The estimated IC₅₀ of **1** was >1,000 μ M. In contrast, analog **2** posed no solubility problems in this assay, generating a reproducible curve all the way to complete inhibition and returning an IC₅₀ value of 51.7 μ M. Thus, the replacement of C=C in **1** with a *threo*-difluoroalkane motif in **2** appears to preserve or enhance the AChE-binding ability, while simultaneously offering the advantage of improved aqueous solubility.



Figure 4: (a) Inihibition of AChE by 1 (IC₅₀ >1000 μ M) and 2 (IC₅₀ = 51.7 μ M); (b) Inhibition of BACE-1 by 1 (IC₅₀ = 59.2 μ M) and 2 (IC₅₀ > 1000 μ M).

Inhibition of BACE-1 was measured using Fluorogenic Peptide Substrate according to an established method (Figure 4b).³³ Intriguingly, the relative activities of **1** *vs.* **2** were reversed in comparison with the AChE assay. The lead compound (**1**) achieved complete inhibition of BACE-1 under the conditions of the assay, giving an IC₅₀ value of 59.2 μ M. In contrast, analog **2** was a much weaker inhibitor of BACE-1, failing to achieve complete inhibition and giving an estimated IC₅₀ value of >1,000 μ M.

There are several conceivable explanations for the reversed relative activities of 1 vs. 2 in the AChE vs. BACE-1 assays. One possibility is that analog 2 is induced to adopt the "correct" conformation when binding to both targets (*i.e.* 2f, Figure 2), and this structure fits well within the AChE active

site but poorly within the BACE-1 active site, perhaps due to unfavorable interactions of the fluorine substituents of **2** with BACE-1 active site residues. A second possibility is that **2** adopts an "incorrect" conformation upon binding to both targets (*e.g.* **2a**, Figure 2), and this novel molecular shape is readily accommodated by $AChE^{34}$ but not BACE-1, perhaps due to different levels of flexibility within the enzymes' active sites. A third possibility is that analog **2** adopts different conformations upon binding to AChE *vs.* BACE-1; since the microenvironments within these enzymes' active sites could be different (e.g. more / less polar), the "correct" binding geometry of **2** might be favored in AChE but not in BACE-1. These possibilities all remain speculative in the absence of high-resolution structural data of the enzyme-ligand complexes.

CONCLUSIONS

A *threo*-difluorinated piperine analog (2) was successfully synthesised through a stepwise route. The physicochemical properties of 2 were found to be superior to piperine itself (1) in two key respects, namely photostability and aqueous solubility. Conformational analysis of 2 by DFT and NMR revealed that the lowest-energy conformations (2a–c) are imperfect mimics of 1, but that a somewhat higher-energy conformation (2f) is a close match for 1. A preliminary biological investigation revealed that analog 2 displays superior inhibitory activity towards AChE, but inferior activity towards BACE-1, relative to 1. While the inhibition of both AChE and BACE-1 is potentially desirable for the purpose of treating Alzheimer's disease, it should be noted that problems have been encountered in clinical trials with BACE-1 as the target,³⁵ suggesting that analog 2 might still have relevance in the context of an Alzheimer's treatment. More generally, our finding that the *threo*-difluoroalkane motif is sometimes, but not always, an effective surrogate for *E*-alkenes suggests that this bioisosteric switch could be exploited more widely in medicinal chemistry as a means of increasing the selectivity of a lead compound towards its desired target.

REFERENCES

- Ørsted, H. C. On Piperine, a New Plant Alkaloid. Schweigers J. Chem. Phys. 1820, 29, 80– 82.
- Ladenburg, A.; Scholtz, M. Synthesis of Piperic Acid and Piperine. *Chem. Ber.* 1894, 27, 2958–2960.

- (3) Epstein, W. W.; Netz, D. F.; Seidel, J. L. Isolation of Piperine from Black Pepper. J. Chem. Educ. 1993, 70, 598–599.
- Meghwal, M.; Goswami, T. K. Piper Nigrum and Piperine: An Update. *Phyther. Res.* 2013, 27, 1121–1130.
- Srinivasan, K. Black Pepper and Its Pungent Principle-Piperine: A Review of Diverse Physiological Effects. *Crit. Rev. Food Sci. Nutr.* 2007, 47, 735–748.
- Manayi, A.; Nabavi, S. M.; Setzer, W. N.; Jafari, S. Piperine as a Potential Anti-Cancer Agent: A Review on Preclinical Studies. *Curr. Med. Chem.* 2018, 25, 4918–4928.
- Yadav, V.; Krishnan, A.; Vohora, D. A Systematic Review on Piper Longum L.: Bridging Traditional Knowledge and Pharmacological Evidence for Future Translational Research. *J. Ethnopharmacol.* 2020, 247, 112255.
- Smilkov, K.; Ackova, D. G.; Cvetkovski, A.; Ruskovska, T.; Vidovic, B.; Atalay, M.
 Piperine: Old Spice and New Nutraceutical? *Curr. Pharm. Des.* 2019, 25, 1729–1739.
- (9) Butt, M. S.; Pasha, I.; Sultan, M. T.; Randhawa, M. A.; Saeed, F.; Ahmed, W. Black Pepper and Health Claims: A Comprehensive Treatise. *Crit. Rev. Food Sci. Nutr.* **2013**, *53*, 875–886.
- (10) Chonpathompikunlert, P.; Wattanathorn, J.; Muchimapura, S. Piperine, the Main Alkaloid of Thai Black Pepper, Protects against Neurodegeneration and Cognitive Impairment in Animal Model of Cognitive Deficit like Condition of Alzheimer's Disease. *Food Chem. Toxicol.* 2010, 48, 798–802.
- (11) Okello, E. J.; Coleman, A.; Seal, C. J. In-Vitro Anti-Cholinesterase Activities By Piperine, an Alkaloid From the Spice Family Piperaceae. *Int. J. Pharm. Sci. Res.* **2015**, *6*, 3726–3732.
- (12) Pai, V.; Chandrashekar, K. S.; Shreedhara, C. S.; Pai, A. In-Silico and in-Vitro Correlation Studies of Natural β-Secretase Inhibitor: An Approach towards Alzheimer's Disease. *Res. J. Pharm. Technol.* 2017, *10*, 3506–3510.
- (13) Čolović, M. B.; Krstić, D. Z.; Lazarević-Pašti, T. D.; Bondžić, A. M.; Vasić, V. M. Acetylcholinesterase Inhibitors: Pharmacology and Toxicology. *Curr. Neuropharmacol.* 2013, *11*, 315–335.
- (14) He, H.; Zhang, Q.; Wang, J. R.; Mei, X. Structure, Physicochemical Properties and

Pharmacokinetics of Resveratrol and Piperine Cocrystals. *CrystEngComm* **2017**, *19*, 6154–6163.

- (15) Grewe, R.; Freist, W.; Neumann, H.; Kersten, S. On the Constituents of Black Pepper. *Chem. Ber.* 1970, *103*, 3752–3770.
- (16) Hashimoto, K.; Yaoi, T.; Koshiba, H.; Maoka, T.; Fujiwara, Y.; Yamamot, Y.; Mori, K. Photochemical Isomerization of Piperine, a Pungent Constituent in Pepper. *Food Sci. Technol. Int.* 1996, 2, 24–29.
- (17) Kozukue, N.; Park, M. S.; Choi, S. H.; Lee, S. U.; Ohnishi-Kameyama, M.; Levin, C. E.;
 Friedman, M. Kinetics of Light-Induced Cis-Trans Isomerization of Four Piperines and Their
 Levels in Ground Black Peppers as Determined by HPLC and LC/MS. *J. Agric. Food Chem.*2007, 55, 7131–7139.
- (18) O'Hagan, D. Understanding Organofluorine Chemistry. An Introduction to the C–F Bond.
 Chem. Soc. Rev. 2008, *37*, 308–319.
- Hunter, L. The C-F Bond as a Conformational Tool in Organic and Biological Chemistry.
 Beilstein J. Org. Chem. 2010, 6, doi:10.3762/bjoc.6.38.
- Meanwell, N. A. Fluorine and Fluorinated Motifs in the Design and Application of Bioisosteres for Drug Design. *J. Med. Chem.* 2018, *61*, 5822–5880.
- (21) Gillis, E. P.; Eastman, K. J.; Hill, M. D.; Donnelly, D. J.; Meanwell, N. A. Applications of Fluorine in Medicinal Chemistry. J. Med. Chem. 2015, 58, 8315–8359.
- (22) Fox, S. J.; Gourdain, S.; Coulthurst, A.; Fox, C.; Kuprov, I.; Essex, J. W.; Skylaris, C. K.; Linclau, B. A Computational Study of Vicinal Fluorination in 2,3- Difluorobutane: Implications for Conformational Control in Alkane Chains. *Chem. Eur. J.* 2015, *21*, 1682–1691.
- (23) Tormena, C. F.; Rittner, R.; Abraham, R. J.; Basso, E. A.; Pontes, R. M. Conformational Analysis. Part 33. An NMR, Solvation and Theoretical Investigation of Conformational Isomerism in N,N-Dimethylfluoroacetamide and N,N-Dimethyl-α-Fluoropropionamide. J. Chem. Soc. Perkin Trans. 2 2000, No. 10, 2054–2059.
- (24) Banik, S. M.; Medley, J. W.; Jacobsen, E. N. Catalytic, Diastereoselective 1,2-Difluorination of Alkenes. J. Am. Chem. Soc. 2016, 138, 5000–5003.

- (25) De Araújo, J. X.; Barreiro, E. J.; Parente, J. P.; Fraga, C. A. M. Synthesis of Piperamides and New Analogues from Natural Safrole. *Synth. Commun.* **1999**, *29*, 263–273.
- (26) Nicoletti, M.; O'Hagan, D.; Slawin, A. M. Z. α,β,γ-Trifluoroalkanes: A Stereoselective Synthesis Placing Three Vicinal Fluorines along a Hydrocarbon Chain. *J. Am. Chem. Soc.* 2005, *127*, 482–483.
- (27) Hunter, L.; Jolliffe, K. A.; Jordan, M. J. T.; Jensen, P.; Macquart, R. B. Synthesis and Conformational Analysis of α,β-Difluoro-γ-Amino Acid Derivatives. *Chem. Eur. J.* 2011, *17*, 2340–2343.
- (28) de Araújo-Júnior, J. X.; de M. Duarte, C.; Maria, M. C.; Parente, J. P.; Fraga, C. A. M.; Barreiro, E. J. Synthesis of Natural Amide Alkaloid Piperdardine and a New Bioactive Analogue. *Synth. Commun.* **2001**, *31*, 117–123.
- Bresciani, S.; O'Hagan, D. Stereospecific Benzylic Dehydroxyfluorination Reactions Using Bio's TMS-Amine Additive Approach with Challenging Substrates. *Tetrahedron Lett.* 2010, 51, 5795–5797.
- (30) Nielsen, M. K.; Ugaz, C. R.; Li, W.; Doyle, A. G. PyFluor: A Low-Cost, Stable, and Selective Deoxyfluorination Reagent. *J. Am. Chem. Soc.* **2015**, *137*, 9571–9574.
- (31) Yamamoto, I.; Jordan, M. J. T.; Gavande, N.; Doddareddy, M. R.; Chebib, M.; Hunter, L. The Enantiomers of Syn-2,3-Difluoro-4-Aminobutyric Acid Elicit Opposite Responses at the GABA C Receptor. *Chem. Commun.* **2012**, *48*, 829–831.
- (32) Mathew, M.; Subramanian, S. In Vitro Screening for Anti-Cholinesterase and Antioxidant Activity of Methanolic Extracts of Ayurvedic Medicinal Plants Used for Cognitive Disorders. *PLoS One* 2014, 9, e86804.
- (33) Liu, Y.; Zhang, W.; Li, L.; Salvador, L. A.; Chen, T.; Chen, W.; Felsenstein, K. M.; Ladd, T. B.; Price, A. R.; Golde, T. E.; et al. Cyanobacterial Peptides as a Prototype for the Design of Potent β-Secretase Inhibitors and the Development of Selective Chemical Probes for Other Aspartic Proteases. *J. Med. Chem.* 2012, *55*, 10749–10765.
- (34) Manap, A. S. A.; Tan, A. C. W.; Leong, W. H.; Chia, A. Y. Y.; Vijayabalan, S.; Arya, A.;
 Wong, E. H.; Rizwan, F.; Bindal, U.; Koshy, S.; et al. Synergistic Effects of Curcumin and
 Piperine as Potent Acetylcholine and Amyloidogenic Inhibitors with Significant
 Neuroprotective Activity in Sh-Sy5y Cells via Computational Molecular Modeling and in

Vitro Assay. Front. Aging Neurosci. 2019, 11, doi: 10.3389/fnagi.2019.00206.

 Moussa-Pacha, N. M.; Abdin, S. M.; Omar, H. A.; Alniss, H.; Al-Tel, T. H. BACE1 Inhibitors: Current Status and Future Directions in Treating Alzheimer's Disease. *Med. Res. Rev.* 2020, 40, 339–384.