

This open access document is published as a preprint in the Beilstein Archives with doi: 10.3762/bxiv.2019.31.v1 and is considered to be an early communication for feedback before peer review. Before citing this document, please check if a final, peer-reviewed version has been published in the Beilstein Journal of Organic Chemistry.

This document is not formatted, has not undergone copyediting or typesetting, and may contain errors, unsubstantiated scientific claims or preliminary data.

Preprint Title	Characterization of two new degradation products of atorvastatin- calcium formed upon treatment with strong acids			
Authors	Jürgen Krauß, Monika Klimt, Markus Luber, Peter Mayer and Franz Bracher			
Article Type	Full Research Paper			
Supporting Information File 1	Supporting information file 1.docx; 77.3 KB			
Supporting Information File 2	Supporting information file 2.pdf; 268.2 KB			
ORCID [®] iDs	Franz Bracher - https://orcid.org/0000-0003-0009-8629			

License and Terms: This document is copyright 2019 the Author(s); licensee Beilstein-Institut.

This is an open access publication under the terms of the Creative Commons Attribution License (<u>http://creativecommons.org/licenses/by/4.0</u>). Please note that the reuse, redistribution and reproduction in particular requires that the author(s) and source are credited. The license is subject to the Beilstein Archives terms and conditions: https://www.beilstein-archives.org/xiv/terms.

The definitive version of this work can be found at: doi: https://doi.org/10.3762/bxiv.2019.31.v1

1	Characterization of two new degradation products of atorvastatin-calcium formed upon				
2	treatment with strong acids				
3					
4	Jürgen Krauß ¹ , Monika Klimt ¹ , Markus Luber ¹ , Peter Mayer ² , Franz Bracher ^{1,*}				
5					
6 7	¹ Department of Pharmacy – Center for Drug Research, Ludwig-Maximilians University Munich, Butenandtstr. 5-13, 81377 Munich, Germany				
8 9	² Department of Chemistry, Ludwig-Maximilians University Munich, Butenandtstr. 5-13, 81377 Munich, Germany				
10					
11	Corresponding author: Franz Bracher, Department of Pharmacy – Center for Drug Research,				
12	Ludwig-Maximilians University Munich, Butenandtstr. 5-13, 81377 Munich, Germany, e-mail:				
13	Franz.Bracher@cup.uni-muenchen.de				
14					
15	Dedicated to the memory of Prof. Dr. Gerhard Rücker, Bonn				
16					
17	Abstract: Atorvastatin-calcium (Lipitor®, Sortis®) is a well-established cholesterol synthesis				
18	enzyme (CSE) inhibitor commonly used in the therapy of hypercholesterolemia. This drug is				
19	known to be sensitive to acid treatment, but only little data has been published on the structures				
20	of the degradation products. Here we report on to the identification of two novel degradation				
21	products of atorvastatin, which are formed only under drastic acidic conditions. While treatment				
22	with conc. sulfuric acid lead to a loss of the carboxanilide residue (accompanied by an				
23	expectable lactonization/dehydration process in the side chain), treatment with conc. aqueous				
24	hydrochloric acid gave a complex, bridged molecule under, C,C-bond formation of the lactone				
25	molety with the pyrrole, migration of the isopropyl group and loss of the carboxanilide residue.				
26	I ne novel degradation products were characterized by NMR spectroscopy, HR-MS data and				
27	A-ray crystal structure analysis.				

Key words: Atorvastatin, stress test, degradation products, crystal structure, fragmentation,cyclization

31

33

34 Introduction

35 Over the past decades, the general trend toward globalization of the supply chains for active pharmaceutical ingredients has created new challenges for the authorities in ensuring the 36 safety and quality of the drug supply [1]. Unprecedented impurities can appear, most likely if 37 limited information is available about details (or alterations) of production processes of drugs. 38 39 On the one hand, it is impossible to check drug substances routinely for all imaginable impurities, on the other hand it is desirable to identify as much as possible degradation 40 products of drugs resulting from inappropriate exposition to potentially harmful conditions 41 during production, manufacturing and storage. For being able to provide relevant data in a 42 43 manageable time frame, two kinds of stress tests have found broad application: accelerated storage conditions (higher temperatures, higher humidity, ...) typically provide reliable data on 44 the stability of a drug, but are still time-consuming; on the other hand in "forced degradation 45 46 experiments" the drug is submitted to more drastic conditions (e.g., strong acid or base, strong 47 oxidant, very high temperature), and potential degradation products can be identified in very 48 short time [2, 3]. However, forced degradation experiments are highly artificial in nature, and 49 thus one has to keep in mind that these extremely drastic conditions are prone to lead to results 50 that might be out of proportion for daily quality control [3]. Nevertheless, knowledge about the outcome of stress tests under extreme conditions helps to get insight into the overall reactivity 51 of drug substances. 52

53 Atorvastatin-calcium (1; marketed as the trihydrate in Lipitor[®], Sortis[®]), is a well-established 54 drug for treatment of hypercholesterolemia [4]. This drug is monographed in the leading pharmacopoeias (Ph. Eur., USP), and a couple of impurities are listed there. Most of these 55 impurities result from the synthesis process (stereoisomers, products resulting from impure 56 57 starting materials or side reactions), and only one of these impurities, lactone 2, is most likely degradation product, resulting from acid-mediated lactonization of the 58 а 3.5-59 dihydroxyheptanoate side chain.

A couple of previous publications deal with stress tests on atorvastatin (and its salts), and an overview has been published by Sirén [5]. Hereby, atorvastatin was found to be sensitive to acidic, oxidative, photochemical and thermal stress. Acidic degradation of atorvastatin was reported to follow a first order kinetic, but decomposition products were not characterized in this [6] and several other reports, which only determined the downsizing of the atorvastatin peak in HPLC after treatment with acid [7, 8, 9 10]. The most prominent decomposition product

upon acidic treatment, compound 2, results from lactonization of the 3,5-dihydroxyheptanoate 66 side chain under moderately acidic conditions (0.1 M HCl) [11, 12, 13, 14]. Shah et al. [15] 67 identified six additional decomposition products upon treatment with 0.1 M HCl at 80 °C for 24 68 h, among which the dehydrated lactone **3** was dominating, accompanied by minor amounts of 69 products arising from dehydration of the δ -hydroxy group and some epimers resulting from 70 71 acid-catalyzed isomerization reactions. In contrast, Vukkum et al. [13] describe, besides lactones 2 and 3, an α , β -unsaturated carboxylic acid 4. Treatment under more drastic 72 73 conditions (6 M HCl, reflux, 3 h) was reported to result mainly in hydrolysis of the anilide moiety to give carboxylic acid 5 [16, 17]. 74

75

- 76 Figure 1. Atorvastatin-calcium trihydrate (1) and previously published decomposition products
- arising from treatment with acids: lactone **2**, dehydrated lactone **3**, α , β -unsaturated carboxylic
- acid **4**, and carboxylic acid **5** (resulting from postulated anilide hydrolysis).





79

- 81 Here we report on the results of our investigations on the decomposition of atorvastatin-calcium
- 82 (1) under strongly acid conditions.
- 83

84 **Results and Discussion**

85 Stress tests

Since lability of atorvastatin towards moderately acidic conditions is well-documented, we 86 aimed at investigating the outcome of incubation with acids under more drastic conditions. 87 Treatment of atorvastatin-calcium trihydrate (1) with 2 M aqueous hydrochloric acid at room 88 89 temperature gave, in accordance with previous reports, only hydroxylactone 2 (55% yield). 90 This outcome was confirmed by comparison with published NMR data [18, 19]. At elevated 91 temperature (reflux, 4 h) a mixture of lactone 2 and known unsaturated lactone 3 [15] (arising 92 from acid-catalyzed dehydration of 2) was obtained. Under even more drastic acidic conditions (refluxing with 6 M hydrochloric acid for 3 h, with 20% aqueous H₂SO₄ for 2 h, or with p-93 toluenesulfonic acid in toluene for 5 h) unsaturated lactone 3 was formed exclusively and in 94 high to almost quantitative yields (Table 1). 95

96 **Table 1.** Acidic stress conditions and decomposition products formed.

Acidic	Decomposition products (yield)				
conditions	2	3	6	7	
2 M HCl, 20 °C,	55%	-	-	-	
2 h					
2 M HCl, reflux,	65%	14%	-	-	
4 h					
6 M HCl, reflux,	-	70	-	-	
3 h					
20% H ₂ SO ₄ , reflux, 2 h	-	>98%	-	-	
<i>p</i> -toluenesulfonic acid, toluene, reflux, 5 h	-	95%	-	-	
37% HCl, reflux,	-	-	96%	-	
5 h					
conc. H ₂ SO ₄ ,	-	-	-	18%	
60 °C, 2 h					

98 When atorvastatin-calcium trihydrate (1) was submitted to extremely strong acidic conditions 99 by refluxing with concentrated (37%) aqueous hydrochloric acid, a new product 6 was formed 100 in almost quantitative yield. The ¹H NMR analysis clearly indicated that the entire carboxanilide partial structure got lost under these conditions. However, no signal was observed which could 101 be attributed to a C-H group at the pyrrole ring. The ¹³C-NMR data showed one carbonyl 102 resonance at 170.33 ppm, assignable to a lactone moiety. The HMBC experiment showed a 103 cross peak between the proposed lactone carbonyl carbon and a neighbouring CH-O group, 104 105 confirming the lactone moiety, and the DEPT spectrum showed a new aliphatic methine 106 resonance at 25.2 ppm. By HR-ESI-MS mass data (found: 404.2020 for [M++H]) a molecular formula of C₂₆H₂₆FNO₂ was confirmed, excluding incorporation of HCl into this artefact. Finally, 107 X-ray crystallography structure analysis (see Figure 2 and Supporting Information) disclosed 108 the structure of 6, bearing a novel, bridged tricyclic 1,5-methanopyrrolo[1,2-e][1,5]oxazonin-3-109 110 one ring system (Scheme 1).

111

112 **Scheme 1.** Formation of novel artefacts **6** and **7** under extremely strong acidic conditions.



- 113
- 114

In contrast, submission of atorvastatin-calcium trihydrate (1) to concentrated sulfuric acid for two hours at 60 °C led to the degradation product **7** in low yield (18%) (Scheme 1). No further decomposition products could be isolated. Here, lactonization and dehydration steps in the side chain took place as observed before in the other acid treatments, however, under these extremely strong, virtually anhydrous acid conditions, the entire carboxanilide residue was

- removed to give the (*S*)-configured 4-unsubstituted pyrrole **7**, as exemplified by a typical CH resonance at 6.20 ppm in the ¹H NMR spectrum. This structure was further confirmed by Xray data (see Figure 2 and Supporting Information).
- 123
- 124 **Figure 2.** Top: Molecular structure of artefact **6**. Shown here is the molecular structure of one
- 125 of three independent molecules in **6** drawn at the 50% ellipsoid probability level. Bottom:
- 126 Molecular structure of artefact **7** (drawn at the 50% ellipsoid probability level).
- 127





130

131 132

133 Discussion

134

In this investigation we first confirmed some pathways of decomposition of atorvastatin under acidic conditions. With dilute mineral acids at room temperature, atorvastatin is conveniently converted into the lactone 2 under retention at the C5-O bond of the aliphatic chain [13, 20], whereas treatment under more drastic conditions (e.g., 6 M HCl or heating) causes expectable subsequent dehydration to give the unsaturated lactone 3 [13, 15]. In contrast to previous reports [16, 17] we could not find any indication for a cleavage of the carboxanilide partial structure to give free pyrrolecarboxylic acid **5** under treatment with 6 M HCl under reflux.

142 However, upon treatment with concentrated sulfuric acid, lactonization/dehydration is 143 accompanied by complete loss of the carboxanilide residue to give pyrrole 7. Complete one-144 step removal of carboxamide residues from aromatic rings has been observed before in investigations of fragmentations of protonated species in mass spectrometry [21, 22]. For 145 146 benzanilides Tu [21] proposed a mechanism involving protonation of the amide oxygen, followed by 1,3 proton shift to the ring carbon next to the amide carbonyl group, followed by 147 148 elimination of protonated phenyl isocyanate under re-aromatization. For the pyrrole derived 149 substrate investigated here, even direct protonation at C-3 of the pyrrole by strong acid is most 150 likely, due to the significant basicity of pyrroles. Delocalization of the positive charge ($A \leftrightarrow A'$) as shown in Scheme 2 will support the initial ring protonation step. The X-ray analysis of 151 152 compound 7 revealed that the asymmetric center in the lactone ring is (S)-configured, 153 indicating that once again the lactonization step took place with retention at the remaining stereocenter (the shift from (R) to (S) configuration is only a nomenclatory result of altered 154 priorities of residues around the stereocenter upon dehydration). A comparable fragmentation 155 has been observed in the CID (collision-induced dissociation) mass spectrum of atorvastatin, 156 where the base peak observed at m/z 440 clearly corresponds to a loss of phenyl isocyanate 157 158 [22].

159

160 **Scheme 2.** Proposed mechanism for the formation of desamidated product **7**.

161



162

163 In contrast, refluxing **2** with concentrated aqueous hydrochloric acid (37%) lead to the 164 formation of the complex, bridged product **6**. Most likely, this decomposition starts again with 165 lactonization of the 3,5-dihydroxyheptanoate side chain, followed by dehydration to give the 166 unsaturated lactone **3**. This hypothesis was confirmed by refluxing pure lactone **3** with 37% 167 hydrochloric acid, resulting in clean conversion into **6** as well. In the following, this unsaturated 168 lactone (Michael system) most likely performs an acid-mediated intramolecular attack at C-2

of the electron-rich pyrrole ring. In a cascade of subsequent reactions, the carboxanilide moiety 169 at C-3 is eliminated and the isopropyl residue, originally located at C-2 of the pyrrole, is shifted 170 to C-3, rendering the annulated, tetrasubstituted pyrrole 6. The structure of 6 was confirmed 171 by X-ray crystal structure analysis. However, most likely cleavage of the carboxanilide moiety 172 (compare formation of 7 from atorvastatin with concentrated sulfuric acid) is not the initial step 173 in the cascade of reactions leading from intermediate 3 to product 6. In a control experiment, 174 175 we treated compound 7 with 37% hydrochloric acid under the above mentioned conditions, but only a complex mixture of decomposition products was obtained, with no indication for 176 177 formation of 6. As the intermediate occurrence of a positive charge at C-2 of the pyrrole ring and a sp³-hybridized C-3 are most likely triggering the elimination of phenyl isocyanate from 178 179 the pyrrole (see postulated mechanism shown in Scheme 2), we propose the formation of an intermediate C, which is formed via initial acid-mediated electrophilic attack of the unsaturated 180 lactone at C-2 of the pyrrole ring. The resulting carbenium ion should be stabilized as shown 181 in Scheme 3 ($\mathbf{B} \leftrightarrow \mathbf{B}$). Subsequent shift of the isopropyl group from C-2 to C-3 then would 182 give carbenium(-iminium) ion D, which can eliminate protonated phenyl isocyanate under 183 formation of bridged pyrrole 6. A comparable shift of the isopropyl group in atorvastatin has 184 previously been observed only under oxidative conditions, giving rise to pyrrolidone-type 185 degradation products [23]. 186

187

Scheme 3. Proposed mechanism for the formation of bridged product 6 under cyclization,
 isopropyl migration and carboxanilide fragmentation.

190

191 192



193 **Conclusion**

Atorvastatin-calcium trihydrate (1) is known to be an acid-labile drug, and incubation with dilute 194 mineral acids gives the known lactone 2, which was further dehydrated to unsaturated lactone 195 3 under more drastic conditions. Treatment with very strong acids gave two hitherto unknown 196 degradation products, whose structures were elucidated by NMR and crystal structure 197 analysis. The bridged tricyclic product 6 was formed with concentrated hydrochloric acid, 198 whereas lactone 7 resulted from treatment with concentrated sulfuric acid. We propose 199 200 mechanisms for the formation of the novel artefacts 6 and 7 here. But it has to be mentioned that these new artefacts, which are formed only under extremely drastic conditions, are not 201 202 likely to be relevant in terms of drug safety and control of impurities in launched atorvastatin 203 batches.

204

205 Acknowledgement

- 206
- 207 The authors thank Prof. Dr. Herbert Mayr for helpful discussions.

208

209 Supporting Information

- 210
- 211 Supporting Information File 1
- 212 Materials and methods; stress tests and analytical data of the products obtained thereby;
- 213 HPLC method for the detection of the novel impurities; details of characterization of 6 and 7 by
- 214 X-ray data; crystallographic data for **6** and **7**.
- 215 [http://www.beilstein-journals.org/bjoc/content/supplementary/.....]

216

217 Supporting Information File 2

218 checkCIF/PLATON report (Structure factors for artefacts 6 (wq033) and 7 (wv633)).

219 [http://www.beilstein-journals.org/bjoc/content/supplementary/.....]

220

221 **References**

- 222 [1] Woo, J.; Wolfgang, S.; Batista, H., Clin. Pharmacol. Ther. **2008**, *83*, 494-497. doi:
- 223 10.1038/sj.clpt.6100493.
- [2] Klick, S.; Muijselaar, P.G.; Waterval, J.; Eichinger, T.; Korn, C.: Gerding, T.K.; Debets,
- A.J.; Vandingenen, J.; Sänger, C.; van den Beld, C.; Somsen, G.; de Jong, G., Pharm.
- 226 Technol. Online **2004**, *29*, 48-66.

- [3] Blessy, M.; Patel, R.D.; Prajapati, P.N.; Agrawal, Y.K., J. Pharm. Anal. **2014**, *4*,159-165.
- doi: org/10.1016/j.jpha.2013.09.003.
- [4] Haque, T.; Khan, B.V., Clin. Lipidol. **2010**, *5*, 615-625. doi: 10.2217/clp.10.55.
- [5] Sirén, H., Atorvastatin and related compounds: Review on analyses of pharmaceutical,
- blood and environmental samples, J. Biomed. Res. Pract. 1 (2017), 100003.
- 232 http://hdl.handle.net/10138/236735.
- 233 [6] Oliveira, M.A.; Yoshida, M.I.; Belinelo, V.J.; Valotto, R.S., Molecules 2013, 18, 1447-
- 234 1456. doi: org/10.3390/molecules18021447.
- [7] Chowdhury, T.; Jakaria, M.; Bhuiyan, D.; Arifujjaman; Mamur, A., J. Sci. Innov. Res. 2014,
 3, 598-601.
- [8] Zaheer, Z.; Farooqui, M.; Mangle, A.A.; Nikalje, A., Afr. J. Pharm. Pharmacol. 2008, 2,
 204-210.
- 239 [9] Chaudhari, B.G.; Patel, P.; Shah, B., Chem. Pharm. Bull. 2007, 55, 241-246. doi:
- 240 org/10.1248/cpb.55.241.
- [10] Chaudhari, B.G.; Patel, N.M.; Shah, P.B.; Patel, L.J.; Patel, V.P., J. AOAC Int. 2007, *90*,
 1539-1546.
- [11] Mohammadi, A.; Rezanour, N.; Ansari Dogaheh, M.; Ghorbani Bidkorbeh, F.; Hashem,
- 244 M.; Walker, R.B., J. Chromatogr. B 2007, 846, 215-221. doi:
- 245 org/10.1016/j.jchromb.2006.09.007.
- [12] Seshadri, R.K.; Desai, M.M.; Raghavaraju, T.V.; Krishnan, D.; Rao, D.V.; Chakravarthy,
- 247 I.E., Sci. Pharm. 2010, 78, 821-834. doi: org/10.3797/scipharm.1004-14.
- 248 [13] Vukkum, P.; Moses Babu, J.; Muralikrishna, R., Sci. Pharm. 2013, 81, 93-114. doi:
- 249 org/10.3797/scipharm.1208-06.
- 250 [14] Rakibe, U.; Tiwari, R.; Rane, V.; Wakte, P., Acta Chrom. 2018, 31, 33-44. doi:
- 251 org/10.1556/1326.2017.00333.
- 252 [15] Shah, R.P.; Kumar, V.; Singh, S., Rapid Commun. Mass Spectrom. **2008**, *22*, 613-622.
- 253 doi: org/10.1002/rcm.3403.
- [16] Darwish, H.W.; Hassan, S.A.; Salem, M.Y.; El-Zeany, B.A., Spectrochim. Acta A, Mol.
- Biomol. Spectrosc. **2016**, *154*, 58-66. doi: org/10.1016/j.saa.2015.10.007.
- [17] Hassan, S.A.; Elzanfaly, E.S.; El-Zeany, S.B.; Salem, M.Y., Acta Pharm. (Zagreb,
- 257 Croatia) **2016**, 66, 479-490. doi: org/10.1515/acph-2016-0040.
- 258 [18] Sawant, P.; Maier, M.E., Tetrahedron 2010, 66, 9738-9744. doi:
- 259 org/10.1016/j.tet.2010.10.028.
- 260 [19] Stach, J.; Havlíček, J.; Plaček, L.; Rádl, S., Coll. Czech. Chem. Commun. 2008, 73, 229-
- 261 246. doi: org/10.1135/cccc20080229.
- [20] Kearney, A.S.; Crawford, L.F.; Mehta, S.C.; Radebaugh, G.W., Pharm. Res. 1993, 10,
- 263 1461-1465. doi: org/10.1023/A:1018923325359.

- 264 [21] Tu, Y.-P., Rapid Commun. Mass Spectrom. 2004, 18, 1345-1351. doi:
- 265 abs/10.1002/rcm.1475.
- 266 [22] Guo, C.; Jiang, K.; Yue, L.; Xia, Z.; Wang, X.; Pan, Y., Org. Biomol. Chem. 2012, 10,
- 267 7070-7077. doi: org/10.1039/C2OB26011E.
- 268 [23] Cermola, F.; DellaGreca, M.; Iesce, M.R.; Montanaro, S.; Previtera, L.; Temussi, F.,
- 269 Tetrahedron **2006**, *62*, 7390-7395. doi: org/10.1016/j.tet.2006.05.027.