



Supporting Information

for

Design, synthesis and application of carbazole macrocycles in anion sensors

Alo Rüütel, Ville Yrjänä, Sandip A. Kadam, Indrek Saar, Mihkel Ilisson, Astrid Darnell, Kristjan Haav, Tõiv Haljasorg, Lauri Toom, Johan Bobacka and Ivo Leito

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Experimental part

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Lowest energy conformation of receptor MC014 with benzoate anion	S223
Lowest energy conformation of receptor MC014 with lactate anion	S224
Lowest energy conformation of receptor MC014 with formate anion	S224

General methods

General purification procedures

Purification of the compounds was performed by column chromatography on silica gel (pore size 60 Å, 230–400 mesh for macrocycles **MC001–MC010** and 70–230 mesh for macrocycles **MC011–MC014**). Analytical thin-layer chromatography (TLC) was conducted on TLC plates (silica gel 60 with fluorescent UV₂₅₄ marker on aluminium backed sheets).

Chemicals and raw materials

All starting materials and solvents used in syntheses were acquired from commercial sources (Sigma-Aldrich, Acros, Alfa Aesar, Fluka) with at least 97% purity and used as received. DMSO-*d*₆ for NMR was obtained from Sigma-Aldrich with at least 99.9% purity. Dry solvents were prepared at least 72 h before use in round-bottomed flasks supplied with 3 Å molecular sieves under an argon atmosphere.

All inert gases used for syntheses were of at least 5.0 purity. Water used in this work was obtained from a MilliQ Advantage A10 system or from an ELGA PURELAB® Ultra system.

For sensor development, the following chemicals were of Selectophore™ grade: bis(2-ethylhexyl) sebacate (DOS, ≥97%, Fluka), tridodecylmethylammonium chloride (TDMACl, Fluka), high molecular weight poly(vinyl chloride) (HMW PVC, Fluka), and tetrahydrofuran (THF, ≥99.5%, Sigma-Aldrich). Nitric acid (65%, J.T. Baker) was diluted for use in the electrode cleaning procedure. The remaining chemicals were of analytical or reagent grade with at least 99% purity, except for 3,4-ethylenedioxythiophene (EDOT, 97%), and acquired from Acros Organics, Alfa Aesar, Fluka, Merck, Sigma-Aldrich, and VWR Chemicals.

General procedures for compound characterization

NMR measurements, characterization and assessment of purity of all synthesis products were carried out on a Bruker Avance-III 700 MHz NMR spectrometer. All spectra were recorded using TopSpin 3.2 software and the chemical shifts calibrated against the residual solvent signal.

COSMO-RS calculations were done using COSMOTermX19 parametrization BP_TZVP_C19 (solvent: DMSO with 0.5% water). UV-vis spectrophotometric measurements were carried out using a Thermo Nicolet Evolution 300 spectrophotometer.

For electrospray (ESI) HRMS analysis, the samples were first dissolved in DMSO with a concentration of ≈ 2 mg/ml and then diluted in MeOH containing 0.1% HCOOH ($\text{pH}_{\text{abs}}^{\text{H}_2\text{O}}$ [1] in the range of 3–5) for ESI+ measurements or MeOH containing 0.1% $\text{NH}_3 \cdot \text{H}_2\text{O}$ ($\text{pH}_{\text{abs}}^{\text{H}_2\text{O}}$ in the range of 8–9) for ESI– measurements. Concentrations of compounds were chosen in such way that appropriate signal intensities were achieved and were in the range of 0.5–2 $\mu\text{g/mL}$. HRMS spectra were obtained on a hybrid Varian 910-FT-ICR-MS system with electrospray ion source. Ionization parameters were as follows: spray chamber temperature, 40 °C; spray needle voltage, 5000 V (–4500 V in negative mode); nebulizing gas (N_2) pressure, 30 psi; API-drying gas (N_2), 15 psi at 300 °C; shield voltage, 600 V; capillary voltage 40 V. The ions, selected by quadrupole, were guided into the FT-ICR analyser cell. Ion guide parameters, FT-ICR ion guide and excitation parameters were optimized for the mass range ($m/z = 100$ –1000). Ion collection time varied from 300 to 1000 ms. FT-ICR analyser cell parameters were: DAC rate 8000 kHz for m/z range of 100–800 direct (broadband); ADC rate 4 MHz; transient length was either 1024 K (262.144 ms) or 2048 K (524.288 ms). For the calibration of the mass axis, samples were spiked with an in-house prepared internal calibration solution, which contained ions with the following exact m/z values [2]: $\text{C}_{16}\text{H}_{36}\text{N}^+$ ($m/z = 242.28423$); $\text{C}_{19}\text{H}_{29}\text{N}_4\text{PF}_3^+$ (m/z

= 401.20765); $\text{C}_{26}\text{H}_{45}\text{N}_7\text{P}_2\text{Cl}^+$ ($m/z = 552.28947$); $\text{C}_{26}\text{H}_{64}\text{N}_{13}\text{P}_4^+$ ($m/z = 682.43526$) in ESI+ mode. For ESI- measurements, a calibration mixture containing anions of perfluorinated Brønsted superacids [3] was used. The ions used for calibration were: $\text{C}_{12}\text{F}_{10}\text{NO}_4\text{S}_2^-$ ($m/z = 475.91145$); $\text{C}_8\text{F}_{17}\text{NO}_2\text{SH}^-$ ($m/z = 497.94620$); $\text{C}_8\text{F}_{18}\text{NO}_4\text{S}_2^-$ ($m/z = 579.89868$); $\text{C}_{12}\text{F}_{26}\text{NO}_4\text{S}_2^-$ ($m/z = 779.88591$). Concentrations of the calibrants in the infused solutions remained within 0.5–1.0 μM .

For nano-ESIMS analysis, the solution was infused using a borosilicate emitter (Thermo Fisher Scientific) to an LTQ Orbitrap XL (Thermo Fisher Scientific) mass spectrometer using a spray voltage of 1.8 kV (tube lens 110 V, capillary temperature 200 °C). The instrument was operated in positive polarity. Ten mass spectra were collected at a resolution setting of 100,000 at 400 m/z , spectra were averaged, and the presence of expected compounds was confirmed by mass accuracy within ± 5 ppm and by similarity to theoretical isotopic distribution.

The reference electrode that was used in all electrochemical measurements was a 6.0726.100 double-junction Ag/AgCl reference electrode (Metrohm AG, Switzerland). Glassy carbon (GC) rods (SIGRADUR® G, HTW Hochttemperatur-Werkstoffe GmbH, Germany) were used as counter electrodes and to prepare working electrodes in-house by fitting them into PVC rods (Simona AG, Germany). All electrochemical polymerisations of solid contacts and impedance measurements were performed with Autolab PGSTAT20 or PGSTAT30 potentiostats (EcoChemie BV, Netherlands) equipped with a FRA2 modules. An EMF16 multichannel interface (Lawson Labs, Inc., United States of America) was used for all potentiometric measurements. A pair of 800 Dosino pumps controlled by a 905 Titrando (Metrohm AG, Switzerland) were used for automated dilutions. pH measurements were performed with a Thermo Scientific Orion Star A111 pH-meter and a Thermo Scientific Orion 9157BNMD pH-electrode. All measurements were performed at room temperature (23 ± 2 °C).

Relative binding affinity measurements

Binding of anions in solution was studied using our previously published relative ^1H NMR titration method.[4] For sample preparation, a few milligrams of receptors were weighed into a NMR tube alongside an anchor molecule, to which binding had previously been quantified using absolute binding measurements.[4–6] The compounds were dissolved in 700 μL $\text{DMSO-}d_6$ with a water content of either 0.5% or 10.0% (m/m) H_2O . Solutions of carboxylates were prepared by weighing the corresponding tetrabutylammonium salt into a vial and dissolving it in 1 mL $\text{DMSO-}d_6$ with a water content of either 0.5% or 10.0% (m/m) H_2O . For each analyte, two solutions were prepared: a dilute (0.17–0.44 M) and a concentrated (0.65–1.41 M) solution. During the titration, a blank spectrum of pure receptors was initially obtained. Then, the analyte was added using an automatic titration syringe. The chemical shifts of the formed host–guest complexes were recorded for each titration step. The dilute solution was used during the first part of the titration, where complexation was partial. The concentrated sample was used towards the end of the titration to ensure full complexation of all host molecules. The recorded chemical shifts of all titration steps were used to calculate the differences of affinity to the anchor molecules, as described in reference [4]. Spectra of all titration experiments are available as supporting information.

Preparation of the membrane cocktails

The solvent polymeric membranes were prepared using PVC, DOS as the plasticiser, and TDMACl as the anion exchanger. The ion-selective membranes (ISMs) were prepared with the following composition as the target: 2 wt % ionophore, 50 mol % TDMACl relative to the ionophore, 65 wt % DOS, and 32 wt % PVC. A control membrane was prepared with no ionophore, 0.7 wt % TDMACl, 66 wt % DOS, and 33 wt % PVC. The membrane components were dissolved in THF to produce membrane cocktails containing 83 wt % THF. See Table S1 for the final compositions of the membrane cocktails.

Table S1: Compositions of the membrane cocktails used in this study

component	wt. %			
	CTRL	MC005	MC009	MC012
ionophore	0.00	0.33	0.33	0.33
TDMACl	0.12	0.12	0.11	0.12
PVC	5.77	5.40	5.30	5.45
DOS	11.22	10.75	11.03	10.80
dry (total)	17.11	16.61	16.78	16.69
THF	82.89	83.39	83.22	83.31

Preparation of the sensor prototypes

Glassy carbon rods (3 mm diameter) encased in PVC (8 mm diameter) were used as the electrode bodies for the sensors. The electrodes were polished using sandpapers, diamond pastes (1–15 μm), and alumina paste (0.3 μm). The electrodes were cleaned prior to electrochemical polymerisation of the solid contact by ultrasonication in deionized water and in ethanol and soaking in 1 M nitric acid. A layer of PEDOT doped with chloride was deposited as the solid contact onto the exposed GC surface ($A \approx 0.07 \text{ cm}^2$) of each electrode. The polymerisations were performed one sensor at a time with a three-electrode cell using a double-junction Ag/AgCl/3 M KCl/1 M KCl reference electrode, a bare GC rod as the counter electrode, and the sensor as the working electrode. The monomer solution, which contained 0.01 M EDOT and 0.1 M KCl, was deaerated by bubbling N_2 through the solution prior to the polymerisations and then blanketed with N_2 during the polymerisations. The polymerisations were performed galvanostatically by applying a constant current of 14 μA for 714 s (10 mC polymerisation charge). The sensors were conditioned in 0.1 M sodium acetate at least overnight and then air-dried prior to drop-casting the membranes. The membranes were drop-cast onto the sensors in two passes of 50 μL with approximately 30 minutes between passes for a total of 100 μL of membrane cocktail applied to each sensor. The THF was allowed to evaporate overnight and then the sensors were conditioned in 0.1 M sodium acetate for at least three days prior to characterisation. Three sensors were prepared with each membrane for this study.

Electrochemical impedance spectroscopy

The sensors were conditioned in 0.1 M sodium acetate prior to the electrochemical impedance spectroscopic (EIS) measurements. EIS measurements were performed in 0.1 M sodium acetate with a three-electrode cell using a double-junction Ag/AgCl/3 M KCl/1 M KCl reference

electrode, a GC rod as the counter electrode, and the sensors as the working electrodes. The measurements were performed potentiostatically at 0 V versus the open-circuit potential using a sinusoidal excitation signal with an amplitude of 100 mV (RMS). Sixty-one measurement points were measured in the frequency range 100 mHz–100 kHz.

Potentiometric measurements

The sensors were conditioned in 0.01 M sodium acetate prior to all of the potentiometric measurements. Liquid junction potentials were estimated using the Henderson equation. Activity coefficients were estimated using the extended Debye–Hückel equation.

Calibration

Potentiometric calibrations were performed with 0.1 M sodium acetate solutions that were diluted with deionized water in half-decade steps. The dilutions were performed at seven-minute intervals with pre-programmed pumps until reaching concentrations where the sensors' potentiometric responses levelled out. Mixing of the solutions was left to the pumping action that occurred during the dilutions. Multiple calibrations were performed with each sensor over the course of their life spans.

pH Sensitivity

The pH sensitivity of the sensors was assessed by measuring their potentiometric responses in 0.01 M acetic acid that was titrated with a mixture of 0.25 M sodium hydroxide and 0.01 M sodium acetate. The pH of the samples was monitored during the measurements and the samples were titrated from pH 3.3 to 10 with constant stirring.

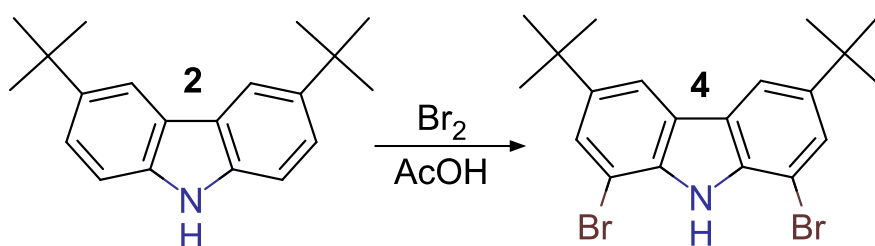
Selectivity

Potentiometric selectivity coefficients, $K_{\text{acetate},j}^{\text{pot}}$, were determined according to the separate solution method (SSM) using solutions with activities of interfering ions equal to that of acetate in a sodium acetate solution with the concentration 0.01 M. The solutions were deaerated by bubbling N_2 through each solution for at least 15 minutes prior to each measurement and then letting N_2 blanket the solution during each measurement. The deaeration was done as a precaution to minimise possible interference due to species originating from dissolved atmospheric carbon dioxide. Constant stirring was used throughout each measurement. The reference electrode was placed in the sample solution after rinsing the sensors with deionized water and drying them. The measurement was started, the sensors were immersed in the solution, and the potentials were measured for at least seven minutes. The measurements were performed with the interfering anion solutions in the order of increasing lipophilicity. The selectivity coefficients were calculated using the experimental slopes determined for each electrode and the potentials that were recorded seven minutes into each selectivity measurement.

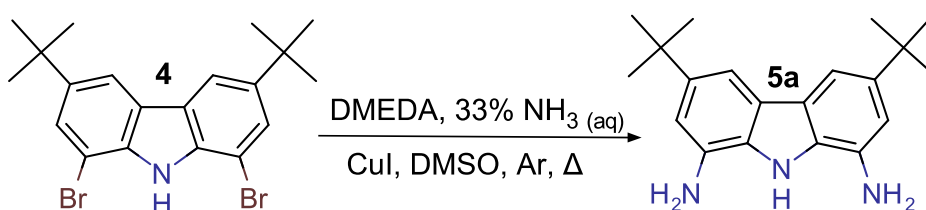
Synthesis and characterisation of compounds

Compound **2** was prepared as in reference [7]. Compounds **6–8** were prepared based on reference [8], but with slight modifications (please refer to Scheme 1 in main text).

1,8-Dibromo-3,6-di-*tert*-butyl-9*H*-carbazole (4) was prepared according to known procedure [9].



For preparation of 3,6-di-*tert*-butyl-9*H*-carbazole-1,8-diamine (**5a**), a general procedure from the work of Jung *et al.* [10] was modified as follows:



A 100-mL pressure tube (withstanding a maximum pressure of 10 bar at 120 °C) was charged with a stirring bar, 1,8-dibromo-3,6-di-*tert*-butyl-9*H*-carbazole (compound **4**, 2.00 g, 4.57 mmol) and CuI (261 mg, 1.37 mmol). DMSO (24 mL) was added and argon gas was bubbled through for 5 minutes while the mixture was magnetically stirred. Under argon counterflow, *N,N*'-dimethylethylenediamine (221 μL , 2.06 mmol) was added, and argon was bubbled through the mixture for an additional minute. After that, the argon flow was stopped and the flask was closed with a screwcap. The contents were magnetically stirred for 20 minutes and then cooled in a water bath (10–15 °C). The stirring was stopped and the screwcap removed, then under an argon counterflow, 12 mL of 33% aqueous NH_3 solution were carefully

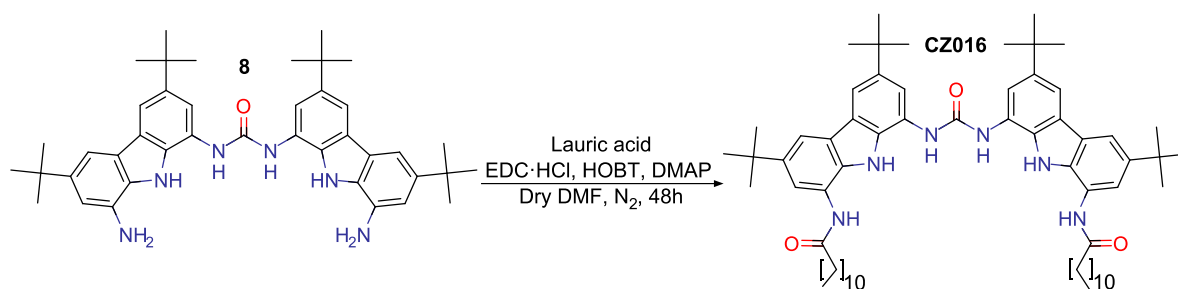
added to the mixture by tube sidewalls without disturbing the contents. Argon flow was removed and the pressure tube was tightly closed with a screwcap. The mixture was vigorously stirred in an oil bath at 130 °C for 18 h. After that, the tube was cooled to approximately 15 °C and its contents were diluted with 100 mL of ethyl acetate. The mixture was filtrated and brine (250 mL) was added to the filtrate. Phases were separated and the aqueous phase was extracted with ethyl acetate (3 × 50 mL). The combined organic phases were washed with brine, dried over anhydrous Na₂SO₄, filtered and solvents removed by evaporation. The resulting residue was triturated with 10 mL DCM, then 100 mL of *n*-hexane was added and the mixture cooled in an ice bath. The precipitate was filtered and washed three times with cold *n*-hexane/DCM 10:1 mixture to yield tan crystals (1.20 g, 3.88 mmol, 85%), the structure of which was confirmed to be identical with literature data [8].

General procedure for preparation of diacyl chlorides (ClCO(CH₂)_{*n*}COCl, *n* = 11–14):



An oven-dried 10-mL round-bottomed flask was charged with a dicarboxylic acid (1 mmol) and a magnetic stirring bar. Then, 2 mL of dry DCM and 1 drop of dry DMF were added. The flask was flushed with argon and closed with a septum through which an argon-filled balloon was connected. The mixture was stirred and thionyl chloride (0.29 mL, 4 mmol) was added dropwise through the septum via syringe. After 4 h of stirring at room temperature the reaction was complete by ¹H NMR analysis. Solvents and residual SOCl₂ were evaporated, the residue dried in vacuo and used in further experiments without additional purification.

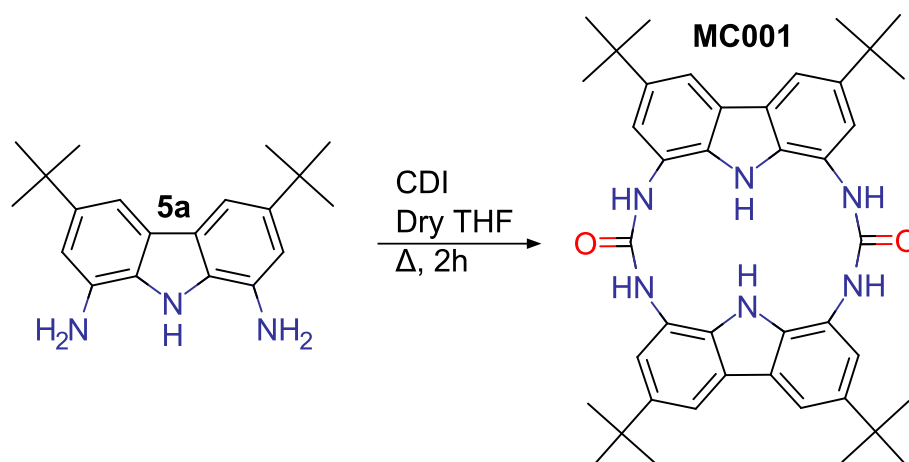
Preparation of compound CZ016



Under N₂ atmosphere, compound **8** (0.050 g, 0.08 mmol), lauric acid (0.038 g, 0.08 mmol), EDC·HCl (0.036 g, 0.19 mmol), HOBT (0.029 g, 0.19 mmol) and DMAP (0.023 g, 0.19 mmol) were dissolved in dry DMF (4 mL). The reaction mixture was stirred for 48 h at room temperature. After disappearance of starting material (monitored by TLC), the reaction mixture was quenched in water (50 mL). The formed precipitate was filtered and washed with water (50 mL). The crude product was purified by column chromatography eluting with 7–9% ethyl acetate in hexane to get compound **CZ016** (0.050 g, 0.05 mmol, 63.8%) as brown solid.

Data for **CZ016**. ¹H NMR (700.1 MHz, DMSO-*d*₆, +25 °C) δ: 10.10 (bs, 2H, amide NH), 10.00 (bs, 2H, carb. NH), 8.89 (bs, 2H, urea NH), 7.95 (s, 2H, CH-5), 7.93 (s, 2H, CH-4), 7.68 (s, 2H, CH-7), 7.53 (s, 2H, CH-2), 2.40-2.35 (m, 4H, -CO-CH₂), 1.61-1.56 (m, 4H, -CO-CH₂-CH₂), 1.42 (s, 18H, *t*-butyl-CH_{3a} or CH_{3b}), 1.40 (s, 18H, *t*-butyl-CH_{3a} or CH_{3b}), 1.37-1.20 (m, 32H, (CH₂)₈), 0.91-0.81 (m, 6H, -CH₂-CH₃). ¹³C NMR (176.0 MHz, DMSO-*d*₆, +25 °C) δ: 171.5 (amide C=O), 153.7 (urea C=O), 141.8 (C-3), 141.5 (C-6), 131.2 (C-12), 130.5 (C-9), 124.6 (C-10), 124.5 (C-11), 123.0 (C-1), 122.6 (C-8), 116.5 (C-7), 116.1 (C-2), 112.1 (C-5), 111.9 (C-4), 36.1 (-CO-CH₂), 34.5 (*t*-butyl-C_a or C_b), 34.4 (*t*-butyl-C_a or C_b), 31.9, 31.8, 31.3, 29.0, 29.0, 28.9, 28.8, 28.7, 28.7, 25.2, 22.1, 13.9 (-CH₂-CH₃). ESI-FT-ICR (-): solvent ~ 0.01% DMSO:methanol, *m/z* of [M-H]⁻ calculated for C₆₅H₉₅N₆O₃, 1007.74711 found 1007.74706.

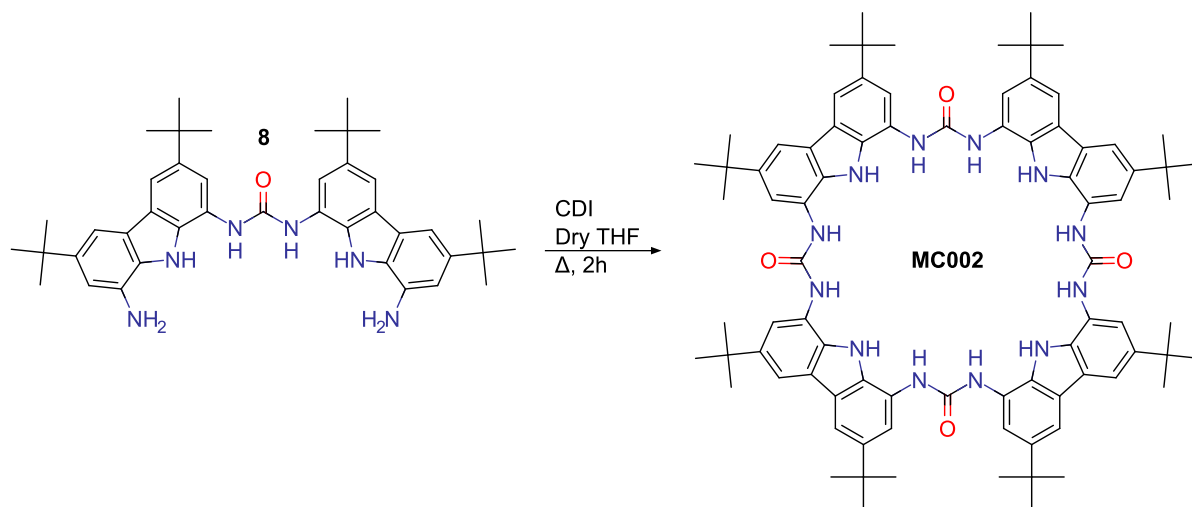
Preparation of compound MC001



Compound **5a** (0.075 g, 0.24 mmol) was dissolved in dry THF (4 mL), then CDI (0.10 g, 0.43 mmol) was added. The reaction mixture was stirred at reflux temperature for 2 h. After disappearance of the starting material (monitored by TLC), the reaction mixture was cooled and concentrated under reduced pressure. The obtained solid was quenched in H₂O (100 mL) and extracted in ethyl acetate (50 mL). The ethyl acetate layer was concentrated under reduced pressure. The crude product was purified by column chromatography eluting with 1% MeOH in DCM to obtain compound **MC001** (0.070 g, 0.10 mmol, 83.3%) as off-white solid.

Data for **MC001**. ¹H NMR (700.1 MHz, DMSO-d₆, +25 °C) δ : 9.87 (bs, 2H, carb. NH), 9.20 (bs, 4H, amide NH), 8.01 (d, $J = 1.4$ Hz, 4H, CH-4), 7.16 (d, $J = 2.1$ Hz, 4H, CH-2), 1.44 (s, 36H, *t*-butyl-CH₃). ¹³C NMR (176.0 MHz, DMSO-d₆, +25 °C) δ : 153.7 (urea CO), 143.2 (C-3), 134.4 (C-6), 126.9 (C-5), 123.9 (C-1), 117.2 (C-2), 113.5 (C-4), 34.9 (*t*-butyl-C), 32.3 (*t*-butyl-CH₃). ESI-ICR (+): solvent ~ 0.01% DMSO:methanol, m/z of [M+H]⁺ calculated for C₄₂H₄₉N₆O₂, 669.39225 found 669.39217.

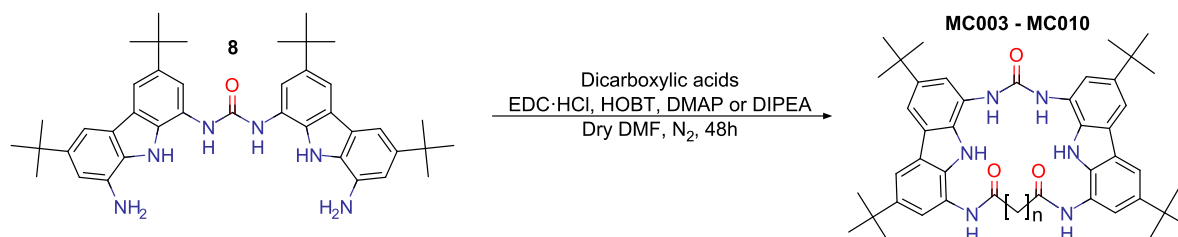
Preparation of compound MC002



Compound **8** (0.10 g, 0.15 mmol), was dissolved in dry THF (25 mL), then CDI (0.04 g, 0.24 mmol) was added. The reaction mixture was stirred at reflux temperature for 6 h. After disappearance of the starting material (monitored by TLC), the reaction mixture was cooled and concentrated under reduced pressure. The obtained solid was quenched in 50 mL H₂O and extracted with 50 mL ethyl acetate. The ethyl acetate layer was concentrated under reduced pressure. The obtained solid was treated with chloroform (20 mL), filtered and washed with chloroform (60 mL). The solid insoluble in chloroform was pure compound **MC002** (0.04 g, 0.03 mmol, 38.4%) isolated as off-white solid.

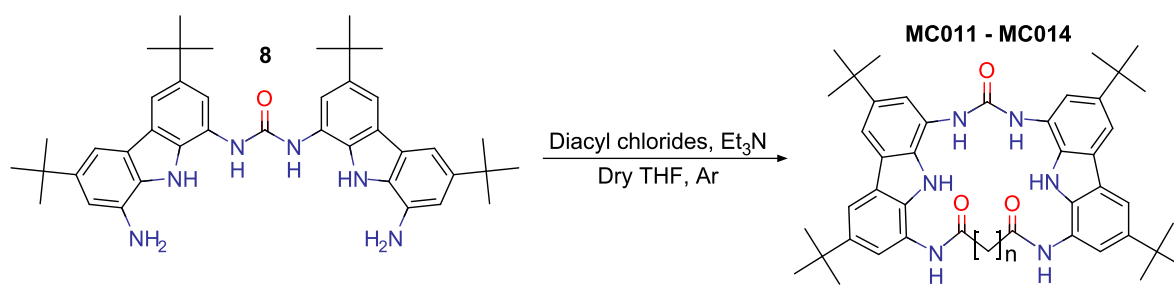
Data for **MC002**. ¹H NMR (700.1 MHz, DMSO-d₆, +25 °C) δ : 9.86 (bs, 4H, carb. NH), 9.21 (bs, 8H, amide NH), 8.01 (d, $J = 1.18$ Hz, 8H, CH-4), 7.16 (d, $J = 1.18$ Hz, 8H, CH-2), 1.44 (s, 72H, (*t*-butyl-CH₃)). ¹³C NMR (176.0 MHz, DMSO-d₆, +25 °C) δ : 153.7 (urea CO), 143.3 (C-3), 134.5 (C-6), 127.0 (C-5), 124.0 (C-1), 117.3 (C-2), 113.6 (C-4), 35.0 (*t*-butyl-C), 32.3 (*t*-butyl-CH₃). ESI-ICR (+): solvent ~ 0.01% DMSO:methanol, m/z of [M+H]⁺ calculated for C₈₄H₁₀₁N₁₂O₄, 1341.80687 found 1341.80722.

Preparation of macrocyclic receptors MC003–MC010



1,3-Bis(8-amino-3,6-di-*tert*-butyl-9*H*-carbazol-1-yl)urea (**8**) was dissolved in dry DMF alongside EDC·HCl, HOBT and, for receptors **MC003–MC006**, DMAP or, for receptors **MC007–MC010**, DIPEA. The corresponding dicarboxylic acid was added. The flask was charged with a magnetic stirring bar and closed with a septum, evacuated and backfilled with N₂ after which a N₂-filled balloon was connected through the septum. The mixture was stirred at room temperature. After disappearance of starting material (monitored by TLC) the reaction was quenched in water (50 mL). The formed precipitate was filtered and washed with water (50 mL). The crude product was purified by column chromatography eluting with MeOH in DCM to afford the macrocycles **MC003–MC010** as white to off-white crystalline powders in 29–87% yields. Please refer to Table S2 for exact reaction conditions for **MC003–MC014**.

Preparation of macrocyclic receptors MC011–MC014



An oven-dried 100-mL round-bottomed flask was charged with a magnetic stirring bar and 1,3-bis(8-amino-3,6-di-*tert*-butyl-9*H*-carbazol-1-yl)urea (**8**) hydrochloride salt (100 mg, 0.139 mmol). The flask was closed with a septum, evacuated and backfilled with argon (3 times) after which an argon-filled balloon was connected through the septum. With a syringe, 36 mL of dry THF and 78 μ L of triethylamine (0.56 mmol) were added and the contents were stirred. To the resulting suspension, the diacyl chloride solution in dry THF (0.139 mmol in 4 mL THF) was added dropwise during one hour. The mixture was stirred overnight at room temperature and then quenched by the addition of 0.1 mL of water. The solvent was removed by evaporation and the residue dissolved in 40 mL of DCM. The solution was washed with 20 mL of 10% aqueous K₂CO₃ and the phases were separated. The aqueous phase was extracted with DCM (3 \times 10 mL) and the combined organic phases were dried on anhydrous Na₂SO₄, filtered, and the solvent evaporated. The residue was purified by column chromatography eluting with DCM-MeOH to yield the desired macrocycles as white to off-white crystals in 25–44% yields. Please refer to Table S2 for exact reaction conditions for **MC011–MC014**.

Table S2: Reaction conditions for **MC003–MC014**.

Compound	8	EDC·HCl	HOBt	DMAP/DIPEA ^[a]	Dicarboxylic acid
MC003	80 mg; 0.12 mmol	59 mg; 0.31 mmol	42 mg; 0.31 mmol	39 mg; 0.31 mmol	16 mg; 0.12 mmol
MC004	60 mg; 0.09 mmol	53 mg; 0.28 mmol	37 mg; 0.28 mmol	23 mg; 0.19 mmol	14 mg; 0.09 mmol
MC005	80 mg; 0.12 mmol	71 mg; 0.37 mmol	50 mg; 0.37 mmol	45 mg; 0.37 mmol	20 mg; 0.12 mmol
MC006	85 mg; 0.13 mmol	76 mg; 0.40 mmol	53 mg; 0.40 mmol	48 mg; 0.40 mmol	23 mg; 0.13 mmol
MC007	91 mg; 0.14 mmol	81 mg; 0.42 mmol	57 mg; 0.42 mmol	55 mg; 0.42 mmol	26 mg; 0.14 mmol
MC008	100 mg; 0.16 mmol	89 mg; 0.47 mmol	63 mg; 0.47 mmol	63 mg; 0.47 mmol	31 mg; 0.16 mmol
MC009	110 mg; 0.17 mmol	98 mg; 0.51 mmol	69 mg; 0.51 mmol	66 mg; 0.51 mmol	37 mg; 0.17 mmol
MC010	100 mg; 0.16 mmol	91 mg; 0.48 mmol	54 mg; 0.40 mmol	61 mg; 0.48 mmol	36 mg; 0.16 mmol

Compound	Solvent ^[b]	Reaction time ^[c]	Column eluent	Yield
MC003	5 ml DMF	60 h	1-2% MeOH/DCM ^[e]	70 mg; 0.09 mmol; 76%
MC004	5 ml DMF	45 h	1-2% MeOH/DCM ^[e]	50 mg; 0.07 mmol; 71%
MC005	5 ml DMF	45 h	1-2% MeOH/DCM ^[e]	80 mg; 0.10 mmol; 87%
MC006	5 ml DMF	45 h	1-2% MeOH/DCM ^[e]	50 mg; 0.06 mmol; 48%
MC007	5 ml DMF	40 h	1-2% MeOH/DCM ^[e]	60 mg; 0.08 mmol; 53%
MC008	5 ml DMF	40 h	1-2% MeOH/DCM ^[e]	75 mg; 0.09 mmol; 64%
MC009	5 ml DMF	40 h	1-2% MeOH/DCM ^[e]	40 mg; 0.05 mmol; 29%
MC010	5 ml DMF	40 h	1-2% MeOH/DCM ^[e]	50 mg; 0.06 mmol; 37%
MC011^[d]	40 ml THF	overnight	1% MeOH/DCM ^[f]	52 mg; 0.06 mmol; 44%
MC012^[d]	40 ml THF	overnight	1% MeOH/DCM ^[f]	47 mg; 0.05 mmol; 39%
MC013^[d]	40 ml THF	overnight	1% MeOH/DCM ^[f]	48 mg; 0.05 mmol; 39%
MC014^[d]	40 ml THF	overnight	1% MeOH/DCM ^[f]	31 mg; 0.04 mmol; 25%

^a - DMAP for **MC001–MC010**, DIPEA for **MC011–MC014**; ^b - all solvents dried before use;

^c - all reactions at room temperature; ^d – amount of **8** for reaction was 100 mg; 0.14 mmol; ^e - gradient elution; ^f - isocratic elution.

Data for **MC003**. ^1H NMR (700.1 MHz, DMSO- d_6 , +25 °C) δ : 10.24 (bs, 2H, carb. NH), 9.96 (bs, 2H, amide NH), 8.95 (bs, 2H, urea NH), 7.99 (d, $J = 1.4$ Hz, 2H, CH -5), 7.97 (d, $J = 1.4$ Hz, 2H, CH -4), 7.91 (d, $J = 1.4$ Hz, 2H, CH -7), 7.15, (d, $J = 1.4$ Hz, 2H, CH -2), 2.52-2.49 (m, 4H, -CO- CH_2), 2.12-2.11 (m, 2H, -CO- CH_2 - CH_2), 1.43 (s, 18H, *t*-butyl- CH_3 _a or CH_3 _b), 1.40 (s, 18H, *t*-butyl- CH_3 _a or CH_3 _b). ^{13}C NMR (176.0 MHz, DMSO- d_6 , +25 °C) δ : 172.1 (amide CO), 154.7 (urea CO), 142.1 (C -3 or C -6), 142.8 (C -3 or C -6), 132.1 (C -12), 131.1 (C -9), 125.2 (C -11), 124.4 (C -10), 123.1 (C -1), 122.9 (C -8), 117.8 (C -7), 117.1 (C -2), 113.0 (C -4 & C -5), 35.3 (CO- CH_2), 34.9 (*t*-butyl- C_a or C_b), 34.8 (*t*-butyl- C_a or C_b), 32.3 (*t*-butyl- CH_3 _a or CH_3 _b), 32.3 (*t*-butyl- CH_3 _a or CH_3 _b), 21.5 (-CO- CH_2 - CH_2). ESI FT-ICR (+): solvent methanol (0.1% HCOOH), m/z of $[\text{M}+\text{H}]^+$ calculated for $\text{C}_{46}\text{H}_{57}\text{N}_6\text{O}_3$, 741.44867 found 741.44826.

Data for **MC004**. ^1H NMR (700.1 MHz, DMSO- d_6 , +25 °C) δ : 10.17 (bs, 2H, amide NH), 9.35 (bs, 2H, carb NH), 8.90 (bs, 2H, urea NH), 8.01 (d, $J = 1.4$ Hz, 2H, CH -5), 7.90 (d, $J = 1.4$ Hz, 2H, CH -4), 7.84 (d, $J = 1.4$ Hz, 2H, CH -2), 7.40 (d, $J = 1.4$ Hz, 2H, CH -7), 2.58-2.52 (m, 4H, -CO- CH_2), 1.93-1.91 (m, 4H, -CO- CH_2 - CH_2), 1.45 (s, 18H, *t*-butyl- CH_3 _a or CH_3 _b), 1.42 (s, 18H, *t*-butyl- CH_3 _a or CH_3 _b). ^{13}C NMR (176.0 MHz, DMSO- d_6 , +25 °C) δ : 171.6 (amide CO), 153.6 (urea CO), 142.6 (C -3 or C -6), 142.4 (C -3 or C -6), 131.8 (C -9), 130.4 (C -12), 125.0 (C -11), 124.1 (C -10), 123.3 (C -1), 122.4 (C -8), 117.1 (C -7), 115.4 (C -2), 113.3 (C -5), 111.4 (C -4), 35.3 (CO- CH_2), 34.6 (*t*-butyl- C_a or C_b), 34.4 (*t*-butyl- C_a or C_b), 32.0 (*t*-butyl- CH_3 _a or CH_3 _b), 31.8 (*t*-butyl- CH_3 _a or CH_3 _b), 24.4 (-CO- CH_2 - CH_2). ESI FT-ICR (+): solvent methanol (0.1% HCOOH), m/z of $[\text{M}+\text{H}]^+$ calculated for $\text{C}_{47}\text{H}_{59}\text{N}_6\text{O}_3$, 755.46432 found 755.46429.

Data for **MC005**. ^1H NMR (700.1 MHz, DMSO- d_6 , +25 °C) δ : 10.07 (bs, 2H, amide NH), 9.93 (bs, 2H, carb. NH), 8.90 (bs, 2H, urea NH), 7.98 (d, $J = 1.4$ Hz, 2H, CH -5), 7.90 (d, $J = 1.4$ Hz, 2H, CH -4), 7.91 (d, $J = 1.4$ Hz, 2H, CH -2), 7.51 (d, $J = 1.4$ Hz, 2H, CH -7), 2.56-2.52 (m, 4H, -CO- CH_2 - CH_2), 1.83-1.79 (m, 4H, (CO- CH_2 - CH_2), 1.60-1.56 (m, 2H, -CO- CH_2 - CH_2 - CH_2), 1.44 (s, 18H, *t*-butyl- CH_3 _a or CH_3 _b), 1.40 (s, 18H, *t*-butyl- CH_3 _a or CH_3 _b). ^{13}C NMR (176.0 MHz,

DMSO-*d*₆, +25 °C) δ: 172.1 (amide CO), 154.0 (urea CO), 142.6 (C-3 or C-6), 142.3 (C-3 or C-6), 131.6 (C-9), 130.9 (C-12), 125.4 (C-11), 124.5 (C-10), 123.9 (C-1), 123.1 (C-8), 117.1 (C-7), 115.6 (C-2), 113.3 (C-5), 111.8 (C-4), 35.6 (-CO-CH₂-CH₂-CH₂), 35.0 (*t*-butyl-C_a or C_b), 34.9 (*t*-butyl-C_a or C_b), 32.4 (*t*-butyl-CH_{3a} or CH_{3b}), 32.3 (*t*-butyl-CH_{3a} or CH_{3b}), 28.4 (-CO-CH₂-CH₂-CH₂), 24.8 (-CO-CH₂-CH₂-CH₂). ESI FT-ICR (+): solvent methanol (0.1% HCOOC), *m/z* of [M+H]⁺ calculated for C₄₈H₆₁N₆O₃, 769.47997 found 769.48020.

Data for **MC006**. ¹H NMR (700.1 MHz, DMSO-*d*₆, +25 °C) δ: 10.11 (bs, 2H, amide NH), 9.66 (bs, 2H, carb. NH), 8.96 (bs, 2H, urea NH), 8.34 (d, *J* = 1.4 Hz, 2H, CH-2), 8.00 (d, *J* = 1.4 Hz, 2H, CH-5), 7.85 (d, *J* = 1.4 Hz, 2H, CH-4), 7.37, (d, *J* = 1.4 Hz, 2H, CH-7), 2.51-2.49 (m, 4H, -CO-CH₂-CH₂), 1.91-1.89 (m, 4H, (-CO-CH₂-CH₂-CH₂), 1.55-1.53 (m, 4H, -CO-CH₂-CH₂-CH₂), 1.45 (s, 18H, *t*-butyl-CH_{3a} or CH_{3b}), 1.41 (s, 18H, *t*-butyl-CH_{3a} or CH_{3b}). ¹³C NMR (176.0 MHz, DMSO-*d*₆, +25 °C) δ: 172.2 (amide CO), 153.2 (urea CO), 142.7 (C-3 or C-6), 142.4 (C-3 or C-6), 131.7 (C-9), 129.1 (C-12), 125.2 (C-8), 124.5 (C-10), 124.5 (C-11), 122.9 (C-1), 117.7 (C-7), 113.7 (C-2), 113.2 (C-5), 110.5 (C-4), 36.6 (-CO-CH₂-CH₂-CH₂), 35.1 (*t*-butyl-C_a or C_b), 34.8 (*t*-butyl-C_a or C_b), 32.4 (*t*-butyl-CH_{3a} or CH_{3b}), 32.3 (*t*-butyl-CH_{3a} or CH_{3b}), 28.4 (-CO-CH₂-CH₂-CH₂), 24.8 (-CO-CH₂-CH₂-CH₂). ESI FT-ICR (-): solvent ~ 0.01% DMSO:isopropanol, *m/z* of [M+H]⁻ calculated for C₄₉H₆₃N₆O₃, 783.4956 found 783.4956.

Data for **MC007**. ¹H NMR (700.1 MHz, DMSO-*d*₆, +25 °C) δ: 10.00 (bs, 2H, amide NH), 9.99 (bs, 2H, carb. NH), 8.82 (bs, 2H, urea NH), 8.10 (d, *J* = 1.4 Hz, 2H, CH-2), 7.97 (d, *J* = 1.4 Hz, 2H, CH-5), 7.89 (d, *J* = 1.4 Hz, 2H, CH-4), 7.60 (d, *J* = 1.4 Hz, 2H, CH-7), 2.51-2.49 (m, 4H, -CO-CH₂-CH₂), 1.80-1.60 (m, 4H, -CO-CH₂-CH₂), 1.55-1.46 (m, 6H, CO-CH₂-CH₂-(CH₂)₂), 1.44 (s, 18H, *t*-butyl-CH_{3a} or CH_{3b}), 1.41 (s, 18H, *t*-butyl-CH_{3a} or CH_{3b}). ¹³C NMR (176.0 MHz, DMSO-*d*₆, +25 °C) δ: 171.9 (amide CO), 153.7 (urea CO), 142.5 (C-3 or C-6), 142.3 (C-3 or C-6), 131.4 (C-9), 130.5 (C-12), 125.2 (C-11), 124.5 (C-10), 124.1 (C-1), 123.0 (C-8), 117.1 (C-7), 115.5 (C-2), 113.2 (C-5), 111.5 (C-4), 36.4 (-CO-CH₂-CH₂-CH₂), 35.0 (*t*-butyl-C_a or

C_b), 34.9 (*t*-butyl-C_a or C_b), 32.4 (*t*-butyl-CH_{3a} or CH_{3b}), 32.2 (*t*-butyl-CH_{3a} or CH_{3b}), 28.2 (-CO-CH₂-CH₂-CH₂-CH₂), 28.1 (-CO-CH₂-CH₂-CH₂-CH₂), 25.0 (-CO-CH₂-CH₂-CH₂-CH₂). ESI FT-ICR (-): solvent ~ 0.01% DMSO:isopropanol, *m/z* of [M+Na]⁺ calculated for C₅₀H₆₄N₆NaO₃, 819.4932 found 819.4932.

Data for **MC008**. ¹H NMR (700.1 MHz, DMSO-*d*₆, +25 °C) δ: 10.10 (bs, 2H, carb. NH), 10.05 (bs, 2H, amide NH), 8.87 (bs, 2H, urea NH), 8.10 (d, *J* = 1.4 Hz, 2H, CH-2), 7.96 (d, *J* = 1.4 Hz, 2H, CH-5), 7.89 (d, *J* = 1.4 Hz, 2H, CH-4), 7.70 (d, *J* = 1.4 Hz, 2H, CH-7), 2.51-2.49 (m, 4H, -CO-CH₂-CH₂), 1.81-1.70 (m, 4H, -CO-CH₂-CH₂), 1.45-1.11 (m, 8H, -CO-CH₂-CH₂-(CH₂)₂), 1.44 (s, 18H, *t*-butyl-CH_{3a} or CH_{3b}), 1.41 (s, 18H, *t*-butyl-CH_{3a} or CH_{3b}). ¹³C NMR (176.0 MHz, DMSO-*d*₆, +25 °C) δ: 171.5 (amide CO), 153.5 (urea CO), 142.0 (C-3 or C-6), 141.8 (C-3 or C-6), 130.3 (C-9), 130.2 (C-12), 124.9 (C-11), 124.1 (C-10), 123.5 (C-1), 122.8 (C-8), 116.0 (C-7), 115.8 (C-2), 112.4 (C-5), 111.1 (C-4), 35.7 (-CO-CH₂-CH₂-CH₂), 34.6 (*t*-butyl-C_a or C_b), 34.4 (*t*-butyl-C_a or C_b), 31.9 (*t*-butyl-CH_{3a} or CH_{3b}), 31.8 (*t*-butyl-CH_{3a} or CH_{3b}), 27.9 (-CO-CH₂-CH₂-CH₂-CH₂), 27.4 (-CO-CH₂-CH₂-CH₂-CH₂), 24.3 (-CO-CH₂-CH₂-CH₂-CH₂). ESI FT-ICR (-): solvent ~ 0.01% DMSO:isopropanol, *m/z* of [M+Na]⁺ calculated for C₅₁H₆₆N₆NaO₃, 833.5088 found 833.5061.

Data for **MC009**. ¹H NMR (700.1 MHz, DMSO-*d*₆, +25 °C) δ: 10.11 (bs, 2H, carb. NH), 9.99 (bs, 2H, amide NH), 8.81 (bs, 2H, urea NH), 7.95 (d, *J* = 1.4 Hz, 2H, CH-5), 7.92 (d, *J* = 1.4 Hz, 2H, CH-4), 7.88 (d, *J* = 1.4 Hz, 2H, CH-2), 7.77 (d, *J* = 1.4 Hz, 2H, CH-7), 2.51-2.49 (m, 4H, -CO-CH₂-CH₂), 1.72-1.69 (m, 4H, -CO-CH₂-CH₂), 1.45-1.11 (m, 10H, -CO-CH₂-CH₂-(CH₂)₃), 1.43 (s, 18H, *t*-butyl-CH_{3a} or CH_{3b}), 1.40 (s, 18H, *t*-butyl-CH_{3a} or CH_{3b}). ¹³C NMR (176.0 MHz, DMSO-*d*₆, +25 °C) δ: 171.9 (amide CO), 154.4 (urea CO), 142.4 (C-3 or C-6), 142.2 (C-3 or C-6), 131.4 (C-9), 130.6 (C-12), 125.1 (C-11), 124.6 (C-10), 123.8 (C-1), 123.3 (C-8), 117.1 (C-2), 116.4 (C-7), 112.8 (C-5), 112.0 (C-4), 36.6 (-CO-CH₂-CH₂-CH₂), 35.0 (*t*-butyl-C_a or C_b), 34.9 (*t*-butyl-C_a or C_b), 32.4 (*t*-butyl-CH_{3a} or CH_{3b}), 32.3 (*t*-butyl-CH_{3a} or CH_{3b}).

CH_{3b}), 28.5 (-CO-CH₂-CH₂-CH₂-CH₂-CH₂), 28.3 (-CO-CH₂-CH₂-CH₂-CH₂-CH₂), 28.0 (-CO-CH₂-CH₂-CH₂-CH₂-CH₂), 25.2 (-CO-CH₂-CH₂-CH₂-CH₂-CH₂). ESI FT-ICR (-): solvent ~ 0.01% DMSO:isopropanol, *m/z* of [M+H]⁺ calculated for C₅₂H₆₉N₆O₃, 825.5425 found 825.5426.

Data for **MC010**. ¹H NMR (700.1 MHz, DMSO-*d*₆, +25 °C) δ: 10.10 (bs, 2H, carb. NH), 10.0 (bs, 2H, amide NH), 8.82 (bs, 2H, urea NH), 7.95 (d, *J* = 1.4 Hz, 2H, CH-5), 7.92 (d, *J* = 1.4 Hz, 2H, CH-4), 7.83 (d, *J* = 1.4 Hz, 2H, CH-2), 7.78 (d, *J* = 1.4 Hz, 2H, CH-7), 2.45-2.49 (m, 4H, -CO-CH₂-CH₂), 1.81-1.70 (m, 4H, -CO-CH₂-CH₂), 1.42 (s, 18H, *t*-butyl-CH_{3a} or CH_{3b}), 1.41 (s, 18H, *t*-butyl-CH_{3a} or CH_{3b}), 1.42-1.11 (m, 12H, -CO-CH₂-CH₂-(CH₂)₃). ¹³C NMR (176.0 MHz, DMSO-*d*₆, +25 °C) δ: 171.9 (amide CO), 154.5 (urea CO), 142.3 (C-3 or C-6), 142.2 (C-3 or C-6), 131.5 (C-9), 130.5 (C-12), 125.1 (C-11), 124.6 (C-10), 123.7 (C-1), 123.3 (C-8), 117.4 (C-2), 116.2 (C-7), 112.8 (C-5), 112.1 (C-4), 36.4 (-CO-CH₂-CH₂-CH₂), 35.0 (*t*-butyl-C_a or C_b), 34.9 (*t*-butyl-C_a or C_b), 32.4 (*t*-butyl-CH_{3a} or CH_{3b}), 32.3 (*t*-butyl-CH_{3a} or CH_{3b}), 28.5 (-CO-CH₂-CH₂-CH₂-CH₂-CH₂), 28.4 (-CO-CH₂-CH₂-CH₂-CH₂-CH₂), 28.3 (-CO-CH₂-CH₂-CH₂-CH₂-CH₂), 25.4 (-CO-CH₂-CH₂-CH₂-CH₂-CH₂). ESI FT-ICR (-): solvent ~ 0.01% DMSO:isopropanol, *m/z* of [M+Na]⁺ calculated for C₅₃H₇₀N₆NaO₃, 861.54071 found 861.54028.

Data for **MC011**. ¹H NMR (700.1 MHz, DMSO-*d*₆, +25 °C) δ: 10.11 (s, 2H, carb. NH), 9.98 (s, 2H, amide NH), 8.82 (s, 2H, urea NH), 7.95 (s, 2H, CH-5), 7.93 (s, 2H, CH-4), 7.86 (s, 2H, CH-7), 7.73 (s, 2H, CH-2), 2.46 (t, *J* = 6.6 Hz, 4H, NHCOCH₂), 1.67 (quint, *J* = 6.6 Hz, 4H, NHCOCH₂CH₂), 1.42 (s, 18H, *t*-Bu CH₃), 1.41 (s, 18H, *t*-Bu CH₃), 1.37 – 1.22 (m, 14H, aliphatic CH₂). ¹³C NMR (176.0 MHz, DMSO-*d*₆, +25 °C) δ: 171.35 (amide CO), 154.16 (urea CO), 141.80 (C3/C6), 141.65 (C3/C6), 131.33 (C-12), 129.90 (C-9), 124.54 (C-10), 124.15 (C-11), 123.12 (C-1), 122.88 (C-8), 117.21 (C-2), 115.65 (C-7), 112.19 (C-5), 111.78 (C-4), 35.88 (NHCOCH₂), 34.49 (*t*-Bu CCH₃), 34.45 (*t*-Bu CCH₃), 31.88 (*t*-Bu CCH₃), 31.84 (*t*-Bu CCH₃),

28.2 – 28.1 (4x aliphatic $\underline{\text{CH}}_2$), 24.74 (NHCOCH $\underline{2}\underline{\text{CH}}_2$). ESI-ICR (+): solvent ~ 0.01% DMSO:methanol, m/z of $[\text{M}+\text{H}]^+$ calculated for C₅₄H₇₃N₆O₃, 853.56995 found 853.57361.

Data for **MC012**. ¹H NMR (700.1 MHz, DMSO-d₆, +25 °C) δ : 10.11 (s, 2H, carb. NH), 9.98 (s, 2H, amide NH), 8.82 (s, 2H, urea NH), 7.95 (s, 2H, CH-5), 7.92 (s, 2H, CH-4), 7.81 (s, 2H, CH-7), 7.73 (s, 2H, CH-2), 2.45 (t, $J = 7.3$ Hz, 4H, NHCOCH $\underline{2}$), 1.67 (quint, $J = 7.3$ Hz, 4H, NHCOCH $\underline{2}\underline{\text{CH}}_2$), 1.41 (s, 18H, *t*-Bu CH $\underline{3}$), 1.40 (s, 18H, *t*-Bu CH $\underline{3}$), 1.36 – 1.21 (m, 16H, aliphatic CH $\underline{2}$). ¹³C NMR (176.0 MHz, DMSO-d₆, +25 °C) δ : 171.40 (amide $\underline{\text{CO}}$), 154.06 (urea $\underline{\text{CO}}$), 141.80 ($\underline{\text{C}}3/\underline{\text{C}}6$), 141.64 ($\underline{\text{C}}3/\underline{\text{C}}6$), 131.19 ($\underline{\text{C}}12$), 129.96 ($\underline{\text{C}}9$), 124.56 ($\underline{\text{C}}10$), 124.11 ($\underline{\text{C}}11$), 123.10 ($\underline{\text{C}}1$), 122.81 ($\underline{\text{C}}8$), 116.98 ($\underline{\text{C}}2$), 115.70 ($\underline{\text{C}}7$), 112.27 ($\underline{\text{C}}5$), 111.73 ($\underline{\text{C}}4$), 36.03 (NHCOCH $\underline{2}$), 34.48 (*t*-Bu $\underline{\text{C}}\text{CH}_3$), 34.44 (*t*-Bu $\underline{\text{C}}\text{CH}_3$), 31.87 (*t*-Bu $\underline{\text{C}}\text{CH}_3$), 31.83 (*t*-Bu $\underline{\text{C}}\text{CH}_3$), 28.2 – 28.0 (4x aliphatic $\underline{\text{CH}}_2$), 25.02 (NHCOCH $\underline{2}\underline{\text{CH}}_2$). ESI-ICR (+): solvent ~ 0.01% DMSO:methanol, m/z of $[\text{M}+\text{H}]^+$ calculated for C₅₅H₇₅N₆O₃, 867.58560 found 867.58949.

Data for **MC013**. ¹H NMR (700.1 MHz, DMSO-d₆, +25 °C) δ : 10.08 (s, 2H, carb. NH), 9.97 (s, 2H, amide NH), 8.82 (s, 2H, urea NH), 7.95 (s, 2H, CH-5), 7.93 (s, 2H, CH-4), 7.82 (s, 2H, CH-7), 7.68 (s, 2H, CH-2), 2.47 – 2.40 (m, 4H, NHCOCH $\underline{2}$), 1.70 – 1.60 (m, 4H, NHCOCH $\underline{2}\underline{\text{CH}}_2$), 1.40 (s, 36H, *t*-Bu CH $\underline{3}$), 1.34 – 1.17 (m, 18H, aliphatic CH $\underline{2}$). ¹³C NMR (176.0 MHz, DMSO-d₆, +25 °C) δ : 171.36 (amide $\underline{\text{CO}}$), 153.97 (urea $\underline{\text{CO}}$), 141.79 ($\underline{\text{C}}3/\underline{\text{C}}6$), 141.63 ($\underline{\text{C}}3/\underline{\text{C}}6$), 131.26 ($\underline{\text{C}}12$), 129.99 ($\underline{\text{C}}9$), 124.53 ($\underline{\text{C}}10$), 124.12 ($\underline{\text{C}}11$), 123.06 ($\underline{\text{C}}1$), 122.81 ($\underline{\text{C}}8$), 116.96 ($\underline{\text{C}}2$), 115.68 ($\underline{\text{C}}7$), 112.28 ($\underline{\text{C}}5$), 111.76 ($\underline{\text{C}}4$), 35.94 (NHCOCH $\underline{2}$), 34.48 (*t*-Bu $\underline{\text{C}}\text{CH}_3$), 34.46 (*t*-Bu $\underline{\text{C}}\text{CH}_3$), 31.88 (*t*-Bu $\underline{\text{C}}\text{CH}_3$), 31.84 (*t*-Bu $\underline{\text{C}}\text{CH}_3$), 28.3 – 28.1 (5x aliphatic $\underline{\text{CH}}_2$), 24.81 (NHCOCH $\underline{2}\underline{\text{CH}}_2$). ESI-ICR (+): solvent ~ 0.01% DMSO:methanol, m/z of $[\text{M}+\text{H}]^+$ calculated for C₅₆H₇₇N₆O₃, 881.60125 found 881.60474.

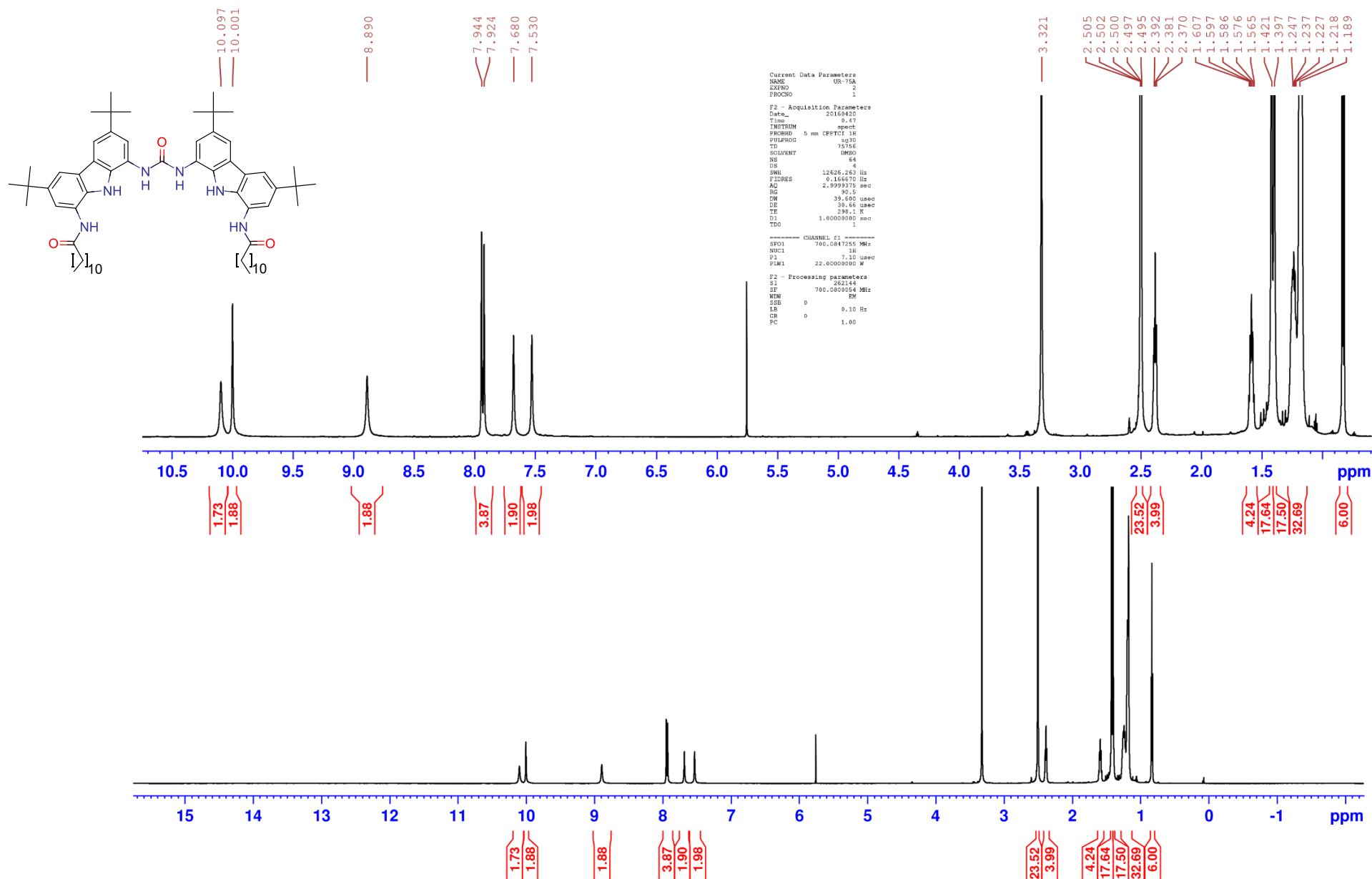
Data for **MC014**. ¹H NMR (700.1 MHz, DMSO-d₆, +25 °C) δ : 10.08 (s, 2H, carb. NH), 9.96 (s, 2H, amide NH), 8.83 (s, 2H, urea NH), 7.95 (s, 2H, CH-5), 7.92 (s, 2H, CH-4), 7.79 (s, 2H,

CH-7), 7.68 (s, 2H, CH-2), 2.47 – 2.40 (m, 4H, NHCOCH₂), 1.70 – 1.60 (m, 4H, NHCOCH₂CH₂), 1.41 (s, 36H, *t*-Bu CH₃), 1.34 – 1.15 (m, 20H, aliphatic CH₂). ¹³C NMR (176.0 MHz, DMSO-d₆, +25 °C) δ: 171.35 (amide CO), 153.89 (urea CO), 141.78 (C3/C6), 141.60 (C3/C6), 131.18 (C-12), 130.09 (C-9), 124.53 (C-10), 124.13 (C-11), 123.03 (C-1), 122.75 (C-8), 116.84 (C-2), 115.78 (C-7), 112.26 (C-5), 111.67 (C-4), 36.00 (NHCOCH₂), 34.45 (*t*-Bu CCH₃), 34.41 (*t*-Bu CCH₃), 31.84 (*t*-Bu CCH₃), 31.81 (*t*-Bu CCH₃), 28.3 – 28.0 (5x aliphatic CH₂), 24.95 (NHCOCH₂CH₂). ESI-ICR (+): solvent ~ 0.01% DMSO:methanol, *m/z* of [M+H]⁺ calculated for C₅₇H₇₉N₆O₃, 895.61690 found 895.62076.

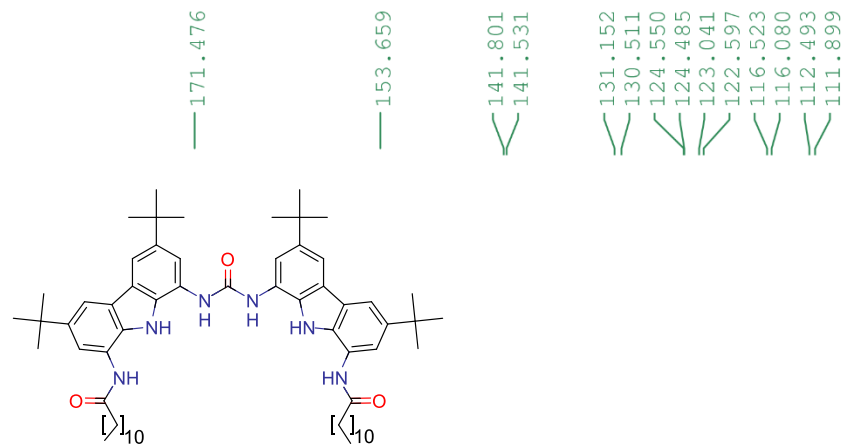
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¹H NMR spectrum (700.1 MHz) of compound **CZ016**



¹³C NMR spectrum (700.1 MHz) of compound **CZ016**

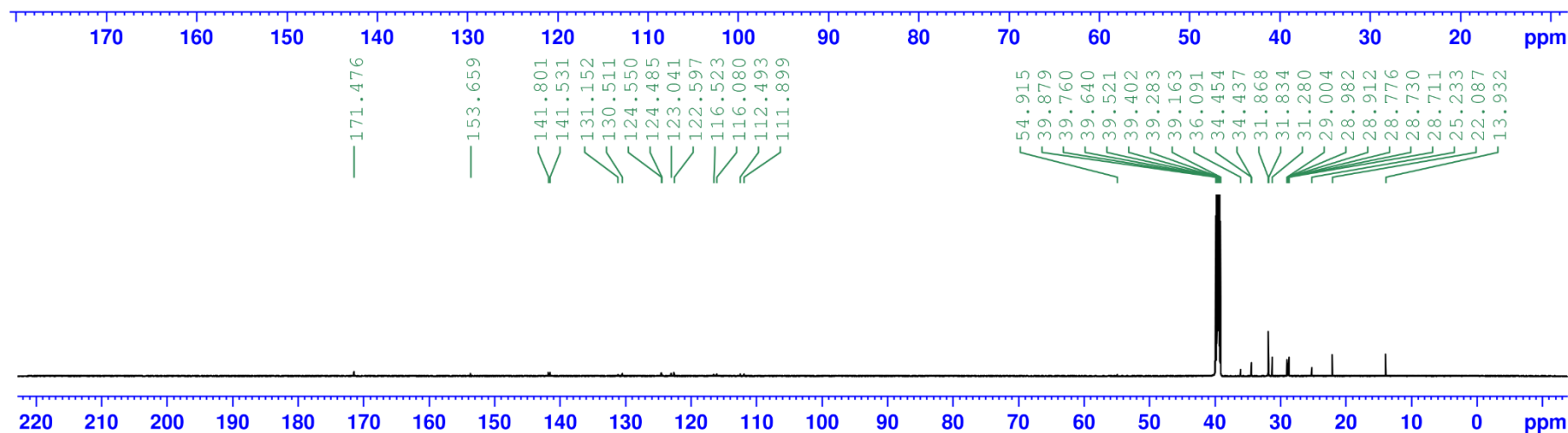
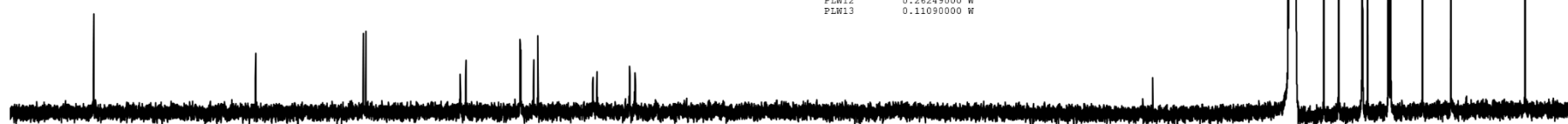


Current Data Parameters
NAME UR-75A
EXPNO 1
PROCNO 1

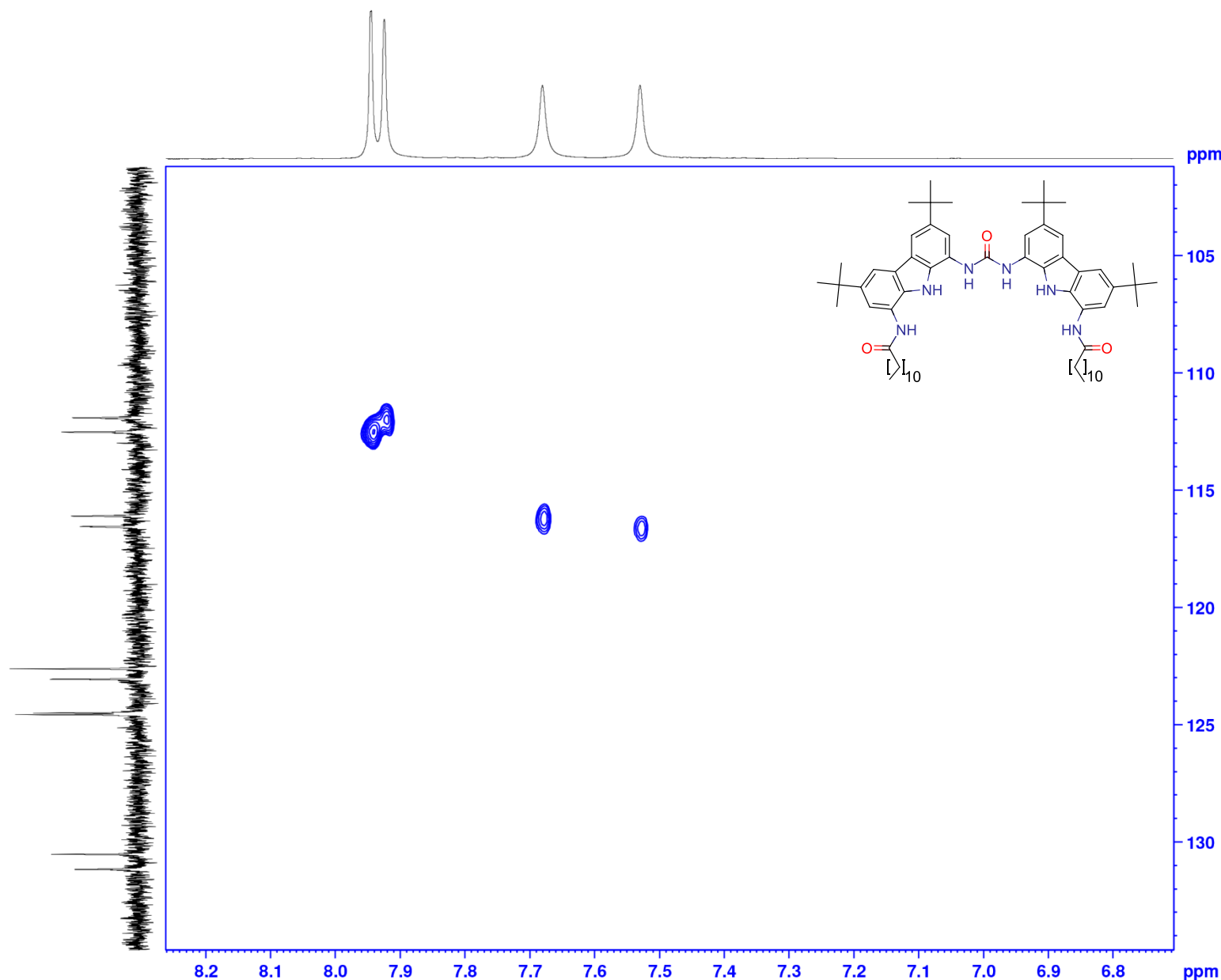
F2 - Acquisition Parameters
Date_ 20160419
Time 22.00
INSTRUM spect
PROBHD 5 mm CPPTCI 1H
PULPROG zgpg30
TD 131072
SOLVENT DMSO
NS 5000
DS 4
SWH 41666.668 Hz
FIDRES 0.317891 Hz
AQ 1.572841 sec
RG 2050
DW 12.000 usec
DE 19.12 usec
TE 298.2 K
D1 2.00000000 sec
D11 0.03000000 sec
TD0 1

===== CHANNEL f1 =====
SFO1 176.0537397 MHz
NUC1 13C
P1 12.40 usec
PLW1 198.00000000 W

===== CHANNEL f2 =====
SFO2 700.0828003 MHz
NUC2 1H
CPDPRG[2] waltz16
PCPD2 65.00 usec
PLW2 22.00000000 W
PLW12 0.26249000 W
PLW13 0.11090000 W



^1H - ^{13}C HSQC spectrum (700.1 MHz) of compound **CZ016**



Current Data Parameters
NAME UR-75A
EXPNO 7
PROCNO 1

F2 - Acquisition Parameters
Date_ 20160420
Time 7.47
INSTRUM spect
PROBHD 5 mm CPPTCI 1H
PULPROG hsqcetdtpasp2.3
TD 1536
SOLVENT DMSO
NS 16
DS 32
SWH 7692.308 Hz
FIDRES 5.008013 Hz
AQ 0.0998400 sec
RG 90.5
DW 65.000 usec
DE 25.00 usec
TE 298.1 K
CNST2 145.0000000
CNST17 -0.5000000
D0 0.00000300 sec
D1 1.50000000 sec
D4 0.00172414 sec
D11 0.03000000 sec
D16 0.00020000 sec
D21 0.00360000 sec
D24 0.00089000 sec
IN0 0.00001670 sec

***** CHANNEL f1 *****
SF01 700.0838154 MHz
NUC1 1H
P1 7.10 usec
P2 14.20 usec
P28 0 usec
PLW1 22.00000000 W

***** CHANNEL f2 *****
SF02 176.0519793 MHz
NUC2 13C
CPDPRG2 bl_p5m4sp_4sp.2
P3 12.40 usec
P14 500.00 usec
P24 2000.00 usec
P31 1600.00 usec
P63 1500.00 usec
PLW0 0 W
PLW2 198.00000000 W
PLW12 10.06400013 W
SFOAL3 Crp60,0.5,20.1
SFOAL3 0 Hz 0.500
SFOFFS3 0 Hz
SFW3 46.51599884 W
SFOAL7 Crp60comp.4
SFOAL7 0 Hz 0.500
SFOFFS7 0 Hz
SFW7 46.51599884 W
SFOAL14 Crp42,1.5,20.2
SFOAL14 0 Hz 0.500
SFOFFS14 0 Hz
SFW14 26.04899979 W
SFOAL18 Crp60_xfilt.2
SFOAL18 0 Hz 0.500
SFOFFS18 0 Hz
SFW18 15.71700001 W
SFOAL31 Crp42,1.5,20.2
SFOAL31 0 Hz 0.500
SFOFFS31 0 Hz
SFW31 6.51219988 W

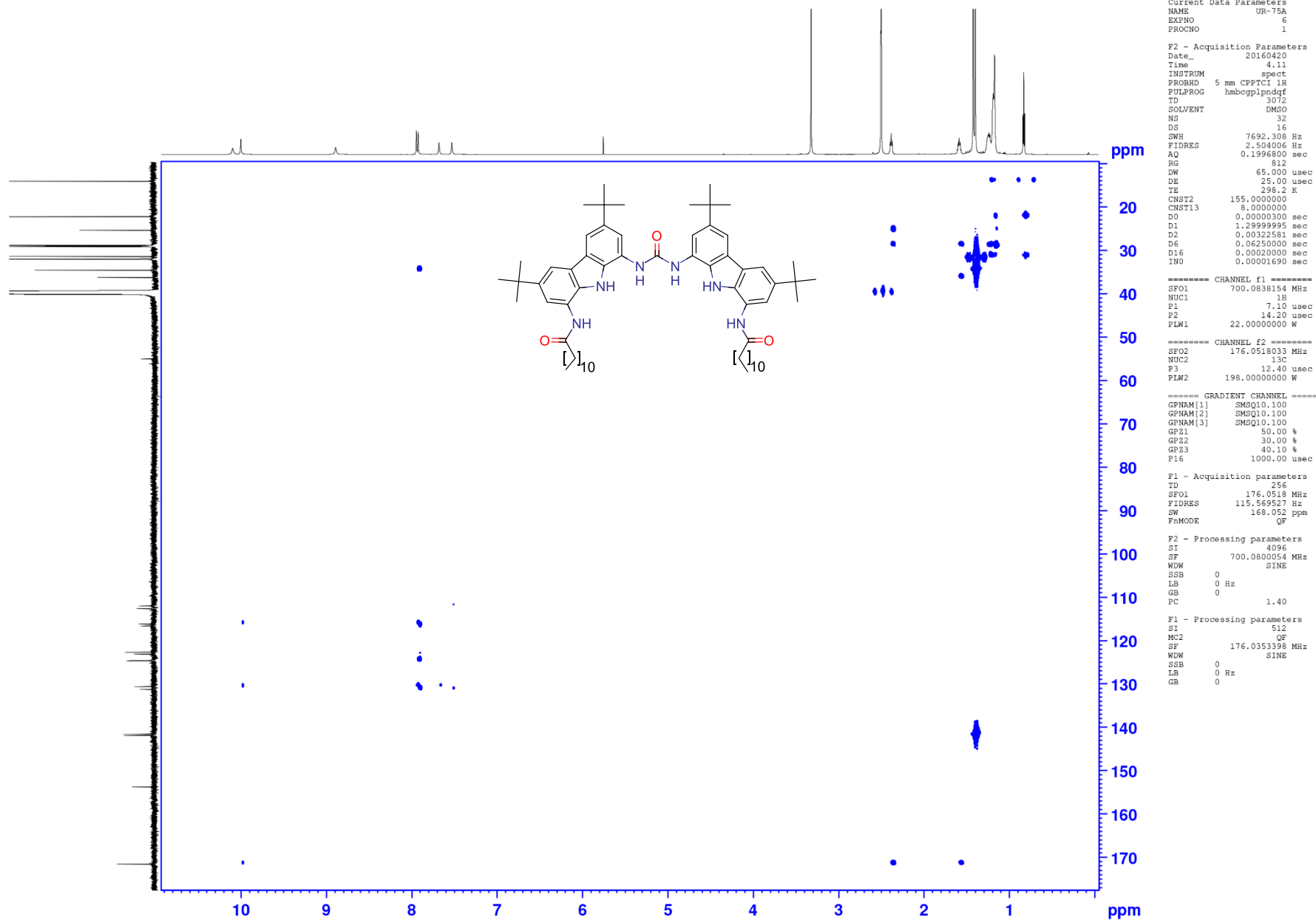
***** GRADIENT CHANNEL *****
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GPNAM[2] SMSQ10.100
GPNAM[3] SMSQ10.100
GPNAM[4] SMSQ10.100
GPZ1 80.00 %
GPZ2 20.10 %
GPZ3 11.00 %
GPZ4 -5.00 %
P16 1000.00 usec
P19 500.00 usec

F1 - Acquisition parameters
TD 160
SF01 176.052 MHz
FIDRES 187.125748 Hz
SW 170.064 ppm
FnMODE Echo-Antiecho

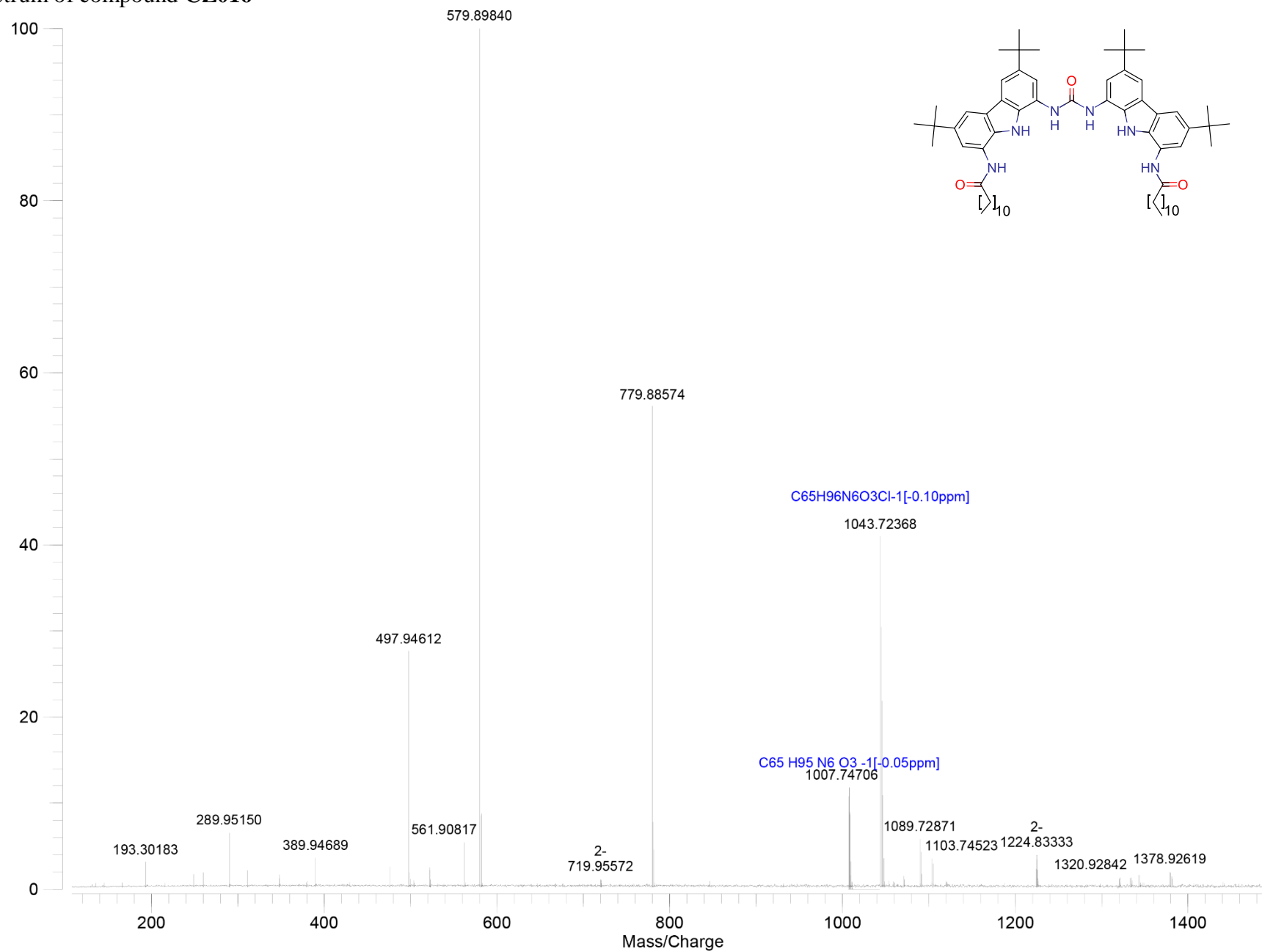
F2 - Processing parameters
SI 4096
SF 700.0799971 MHz
WDW QSINE
SSB 2
LB 0 Hz
GB 0
PC 1.40

F1 - Processing parameters

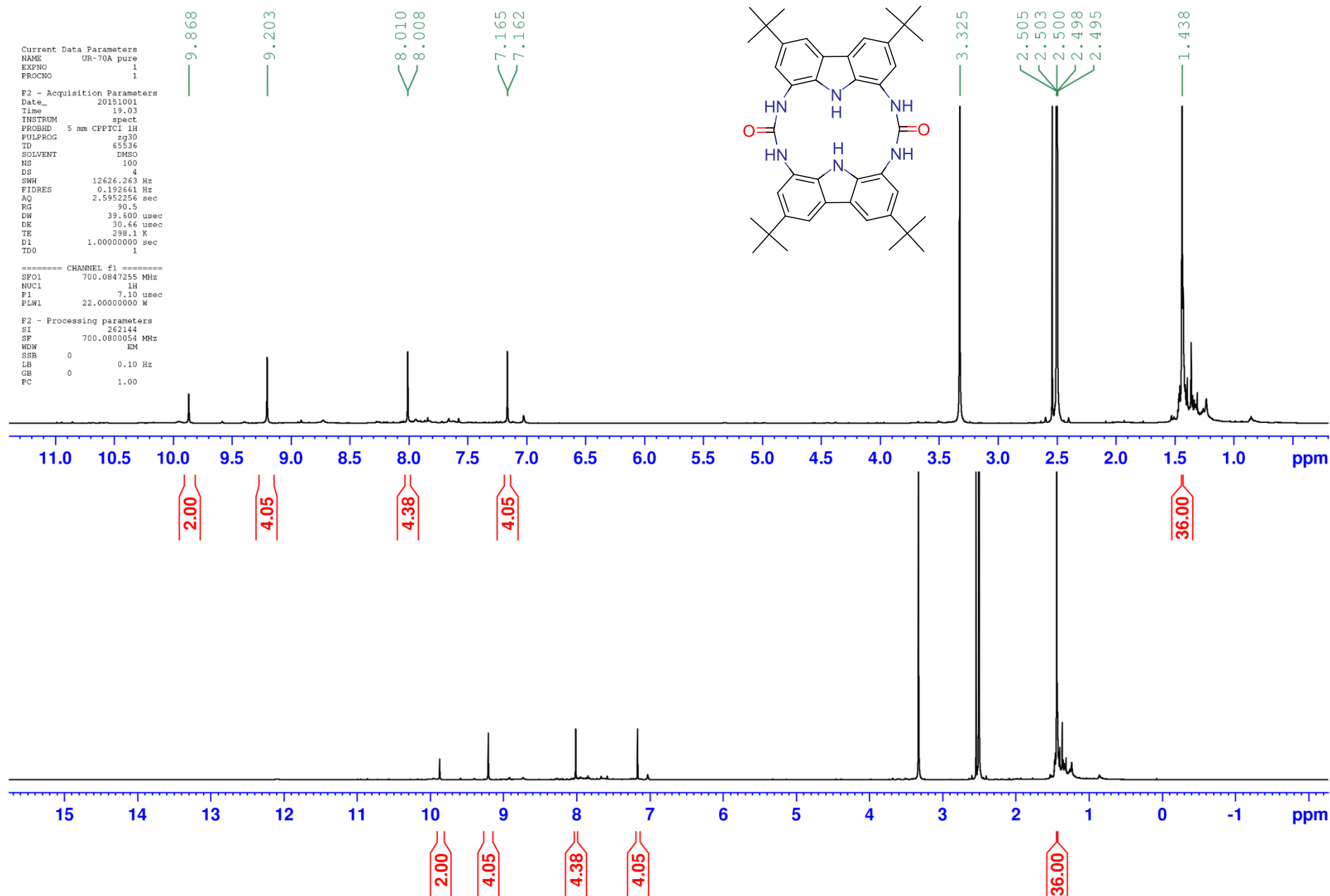
^1H - ^{13}C HMBC spectrum (700.1 MHz) of compound **CZ016**



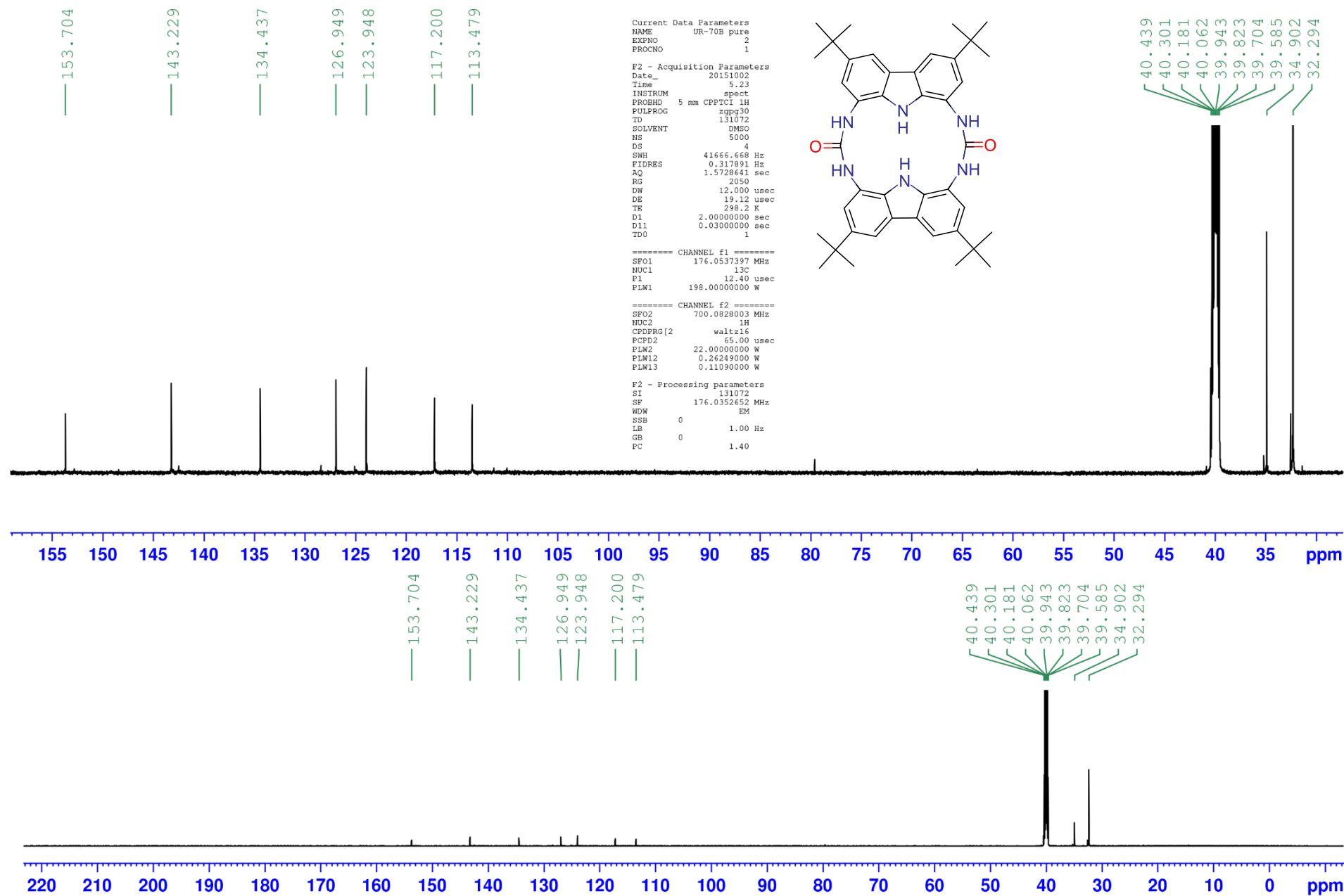
HRMS spectrum of compound **CZ016**



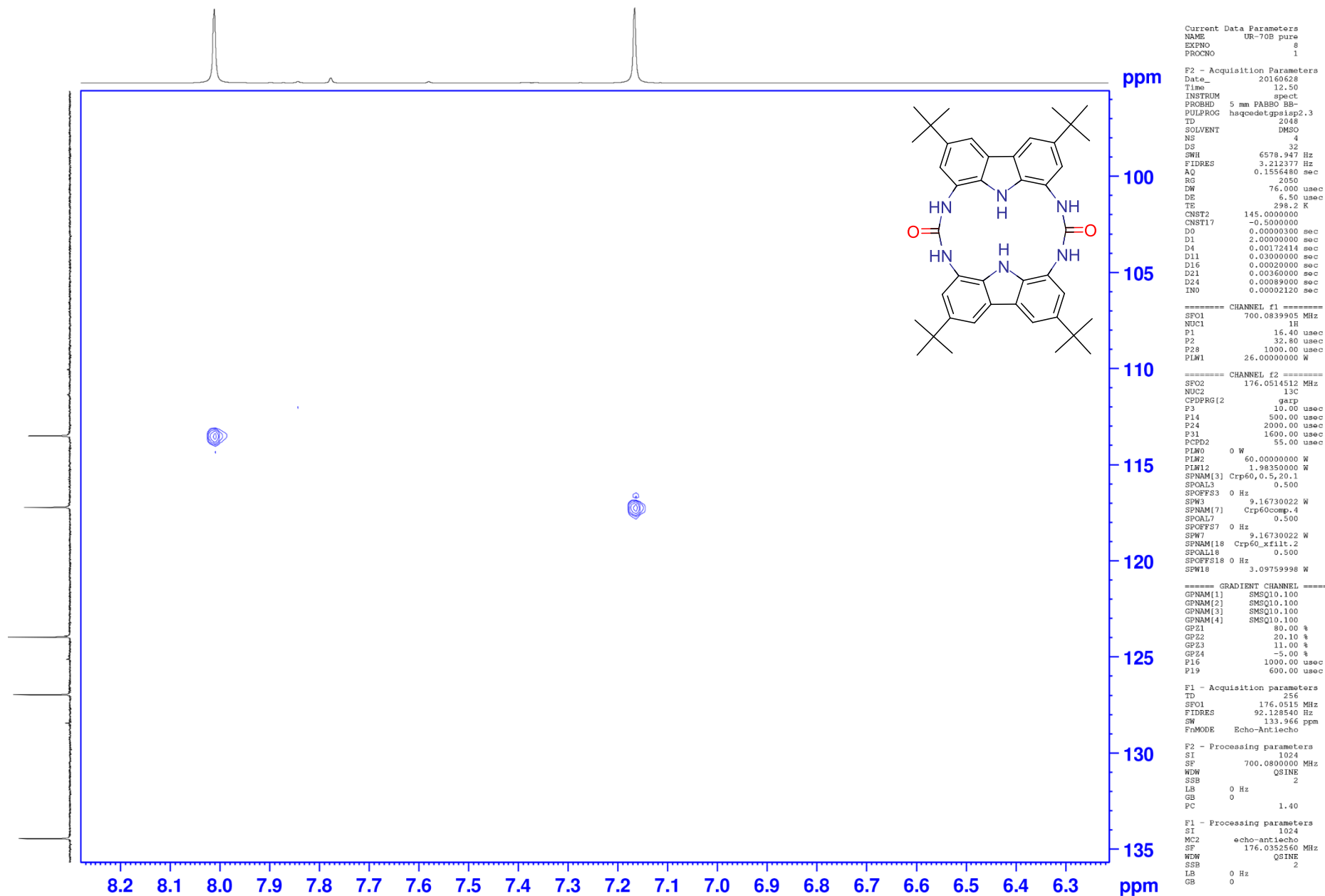
¹H NMR spectrum (700.1 MHz) of compound **MC001**



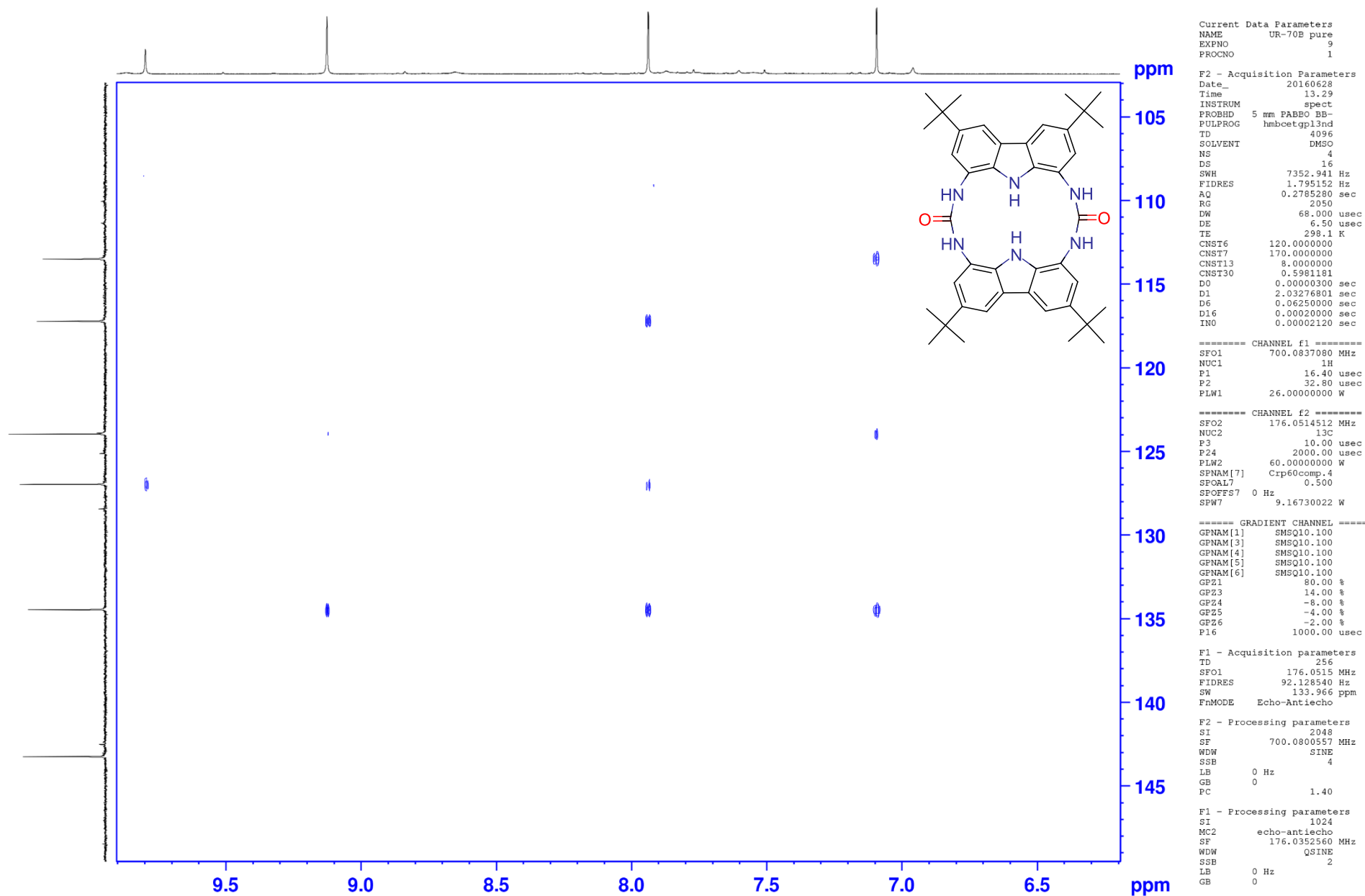
^{13}C NMR spectrum (700.1 MHz) of compound **MC001**



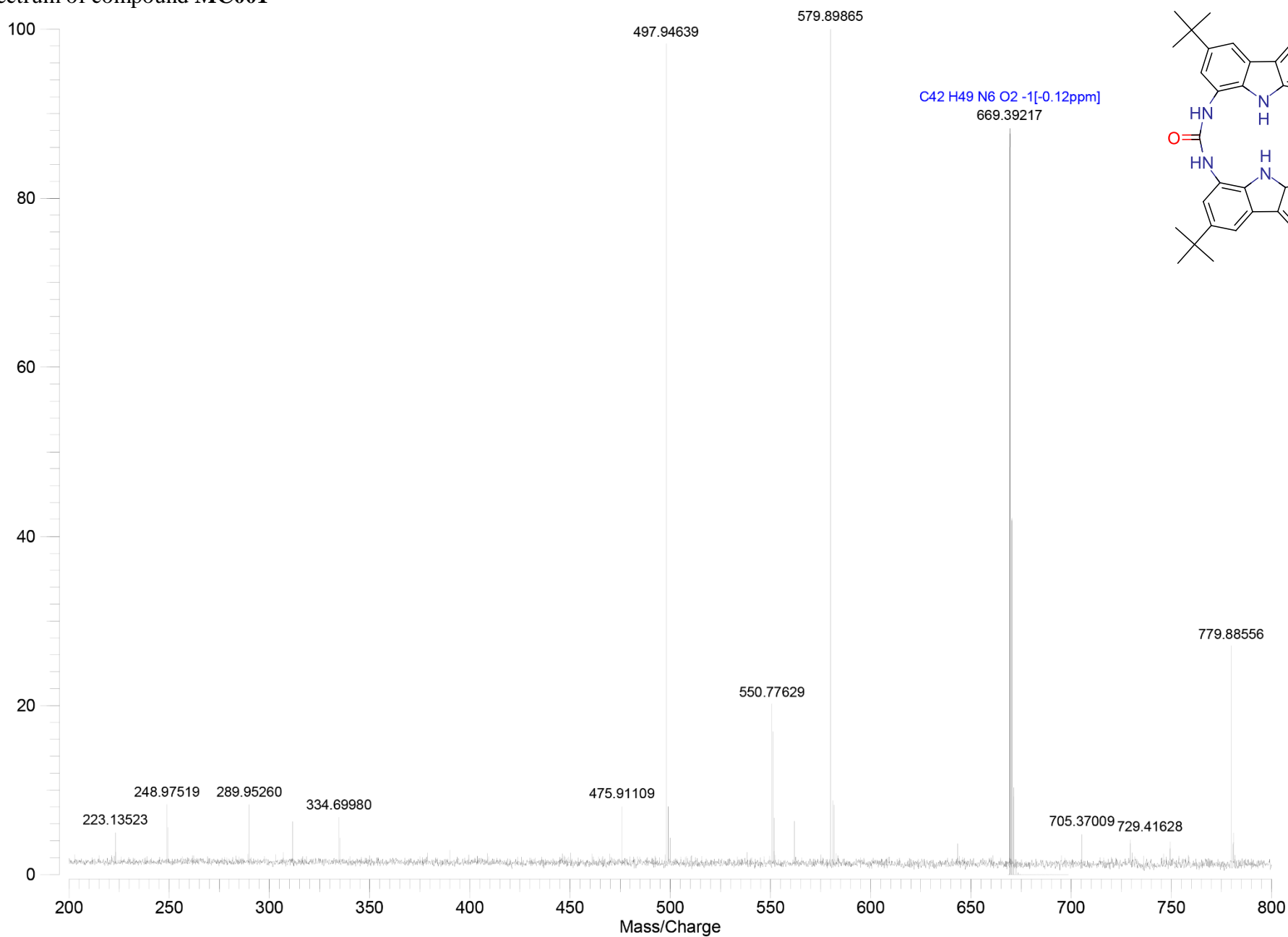
^1H - ^{13}C HSQC spectrum (700.1 MHz) of compound **MC001**



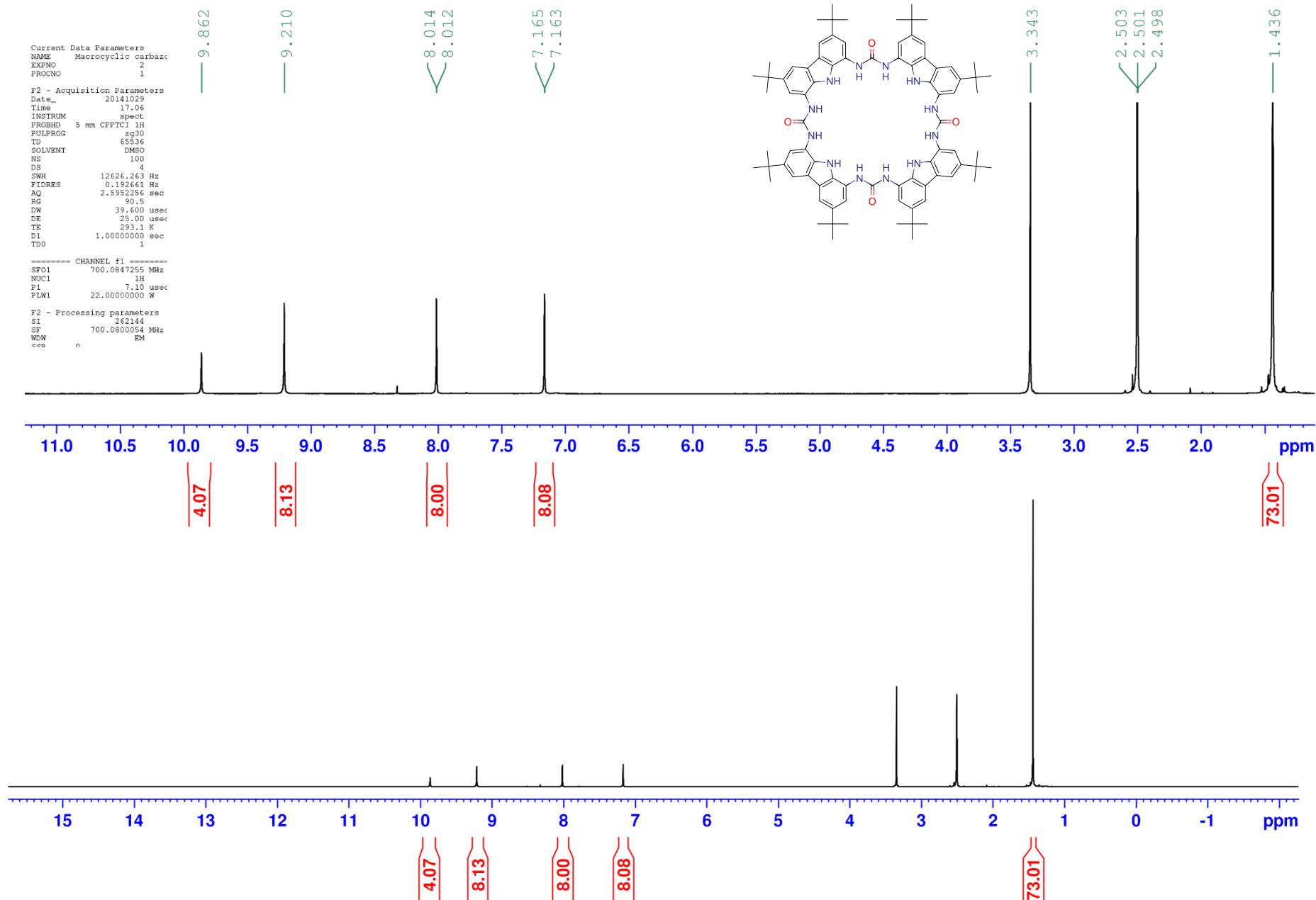
^1H - ^{13}C HMBC spectrum (700.1 MHz) of compound **MC001**



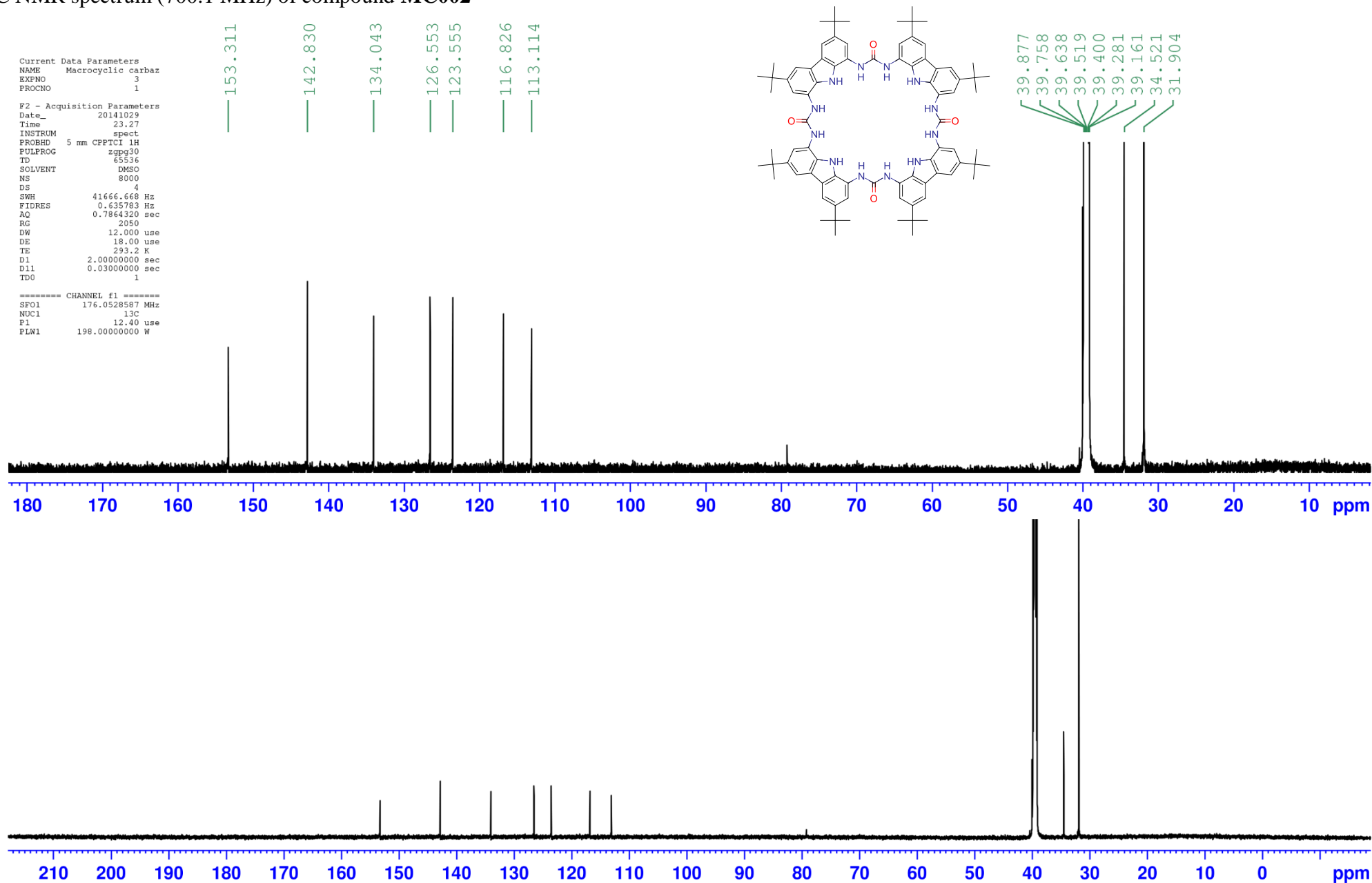
HRMS spectrum of compound MC001



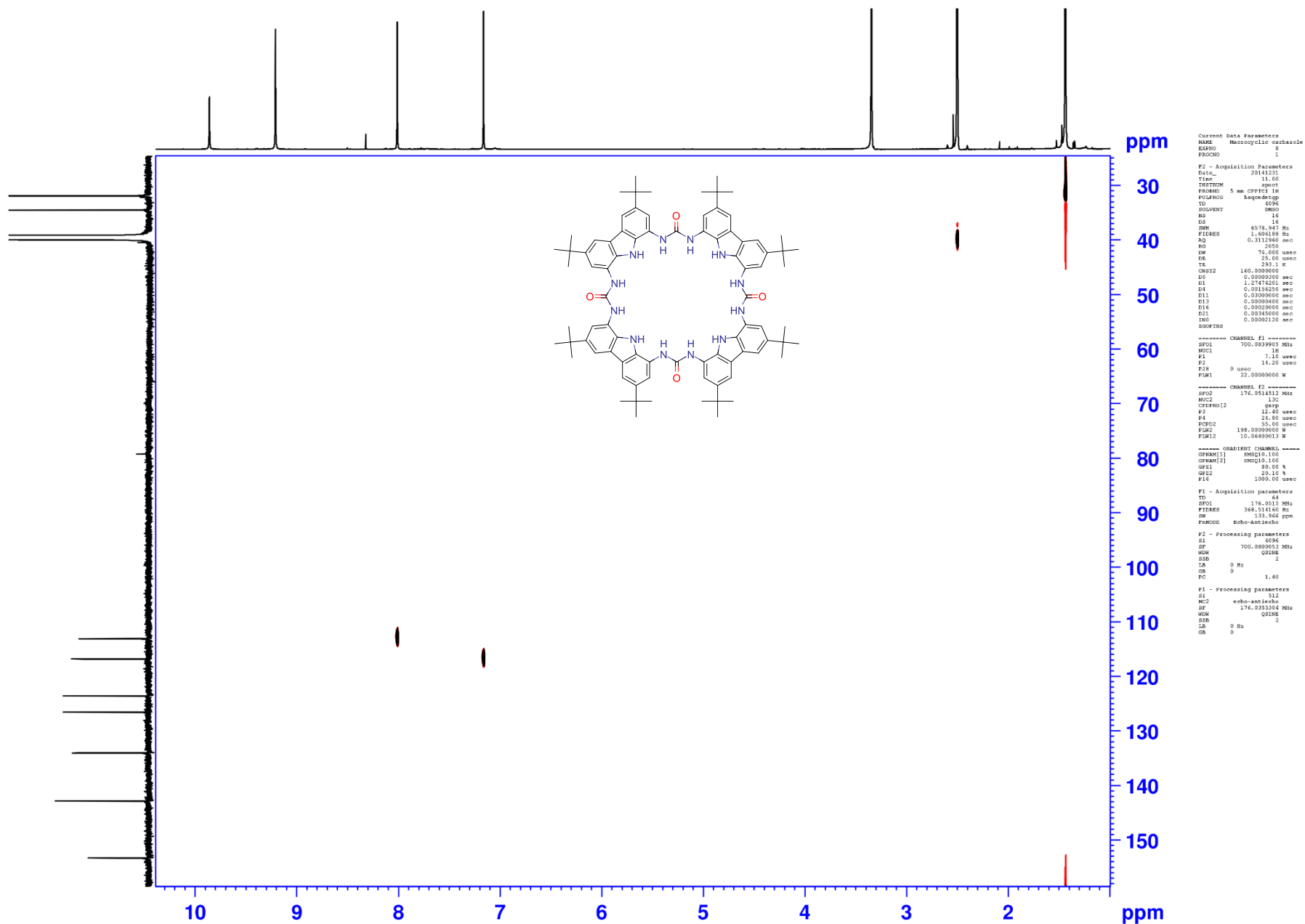
¹H NMR spectrum (700.1 MHz) of compound **MC002**



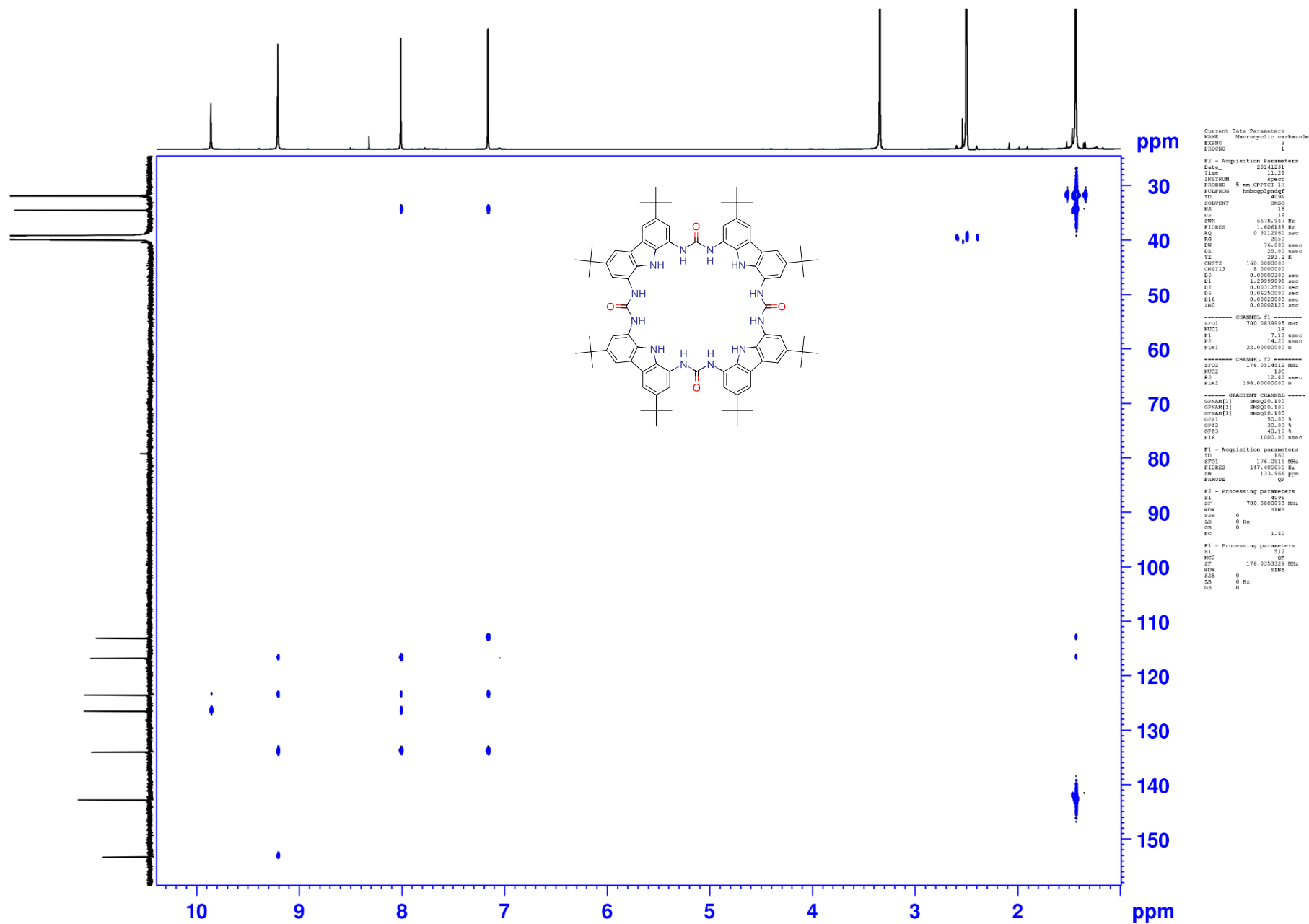
^{13}C NMR spectrum (700.1 MHz) of compound **MC002**



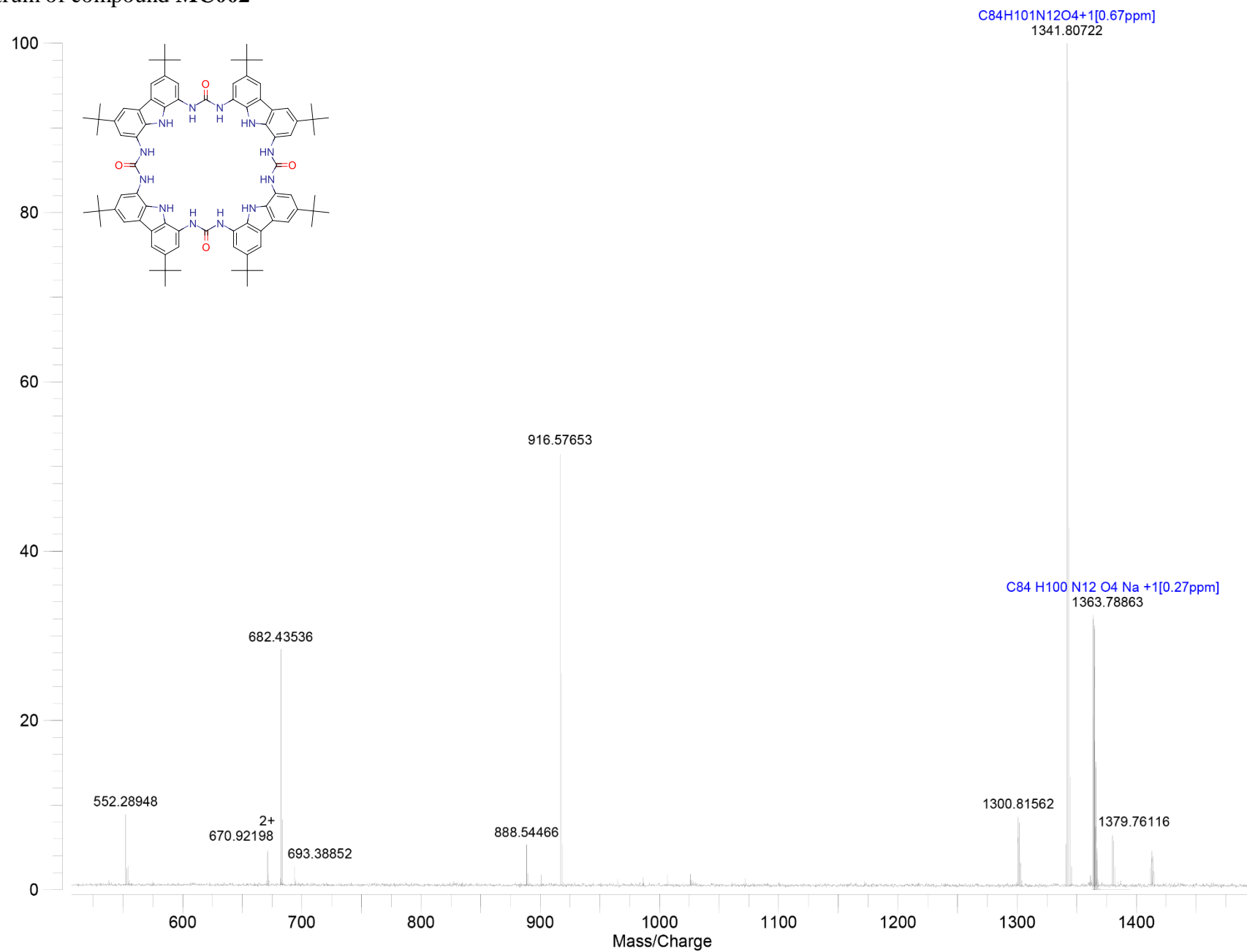
^1H - ^{13}C HSQC spectrum (700.1 MHz) of compound **MC002**



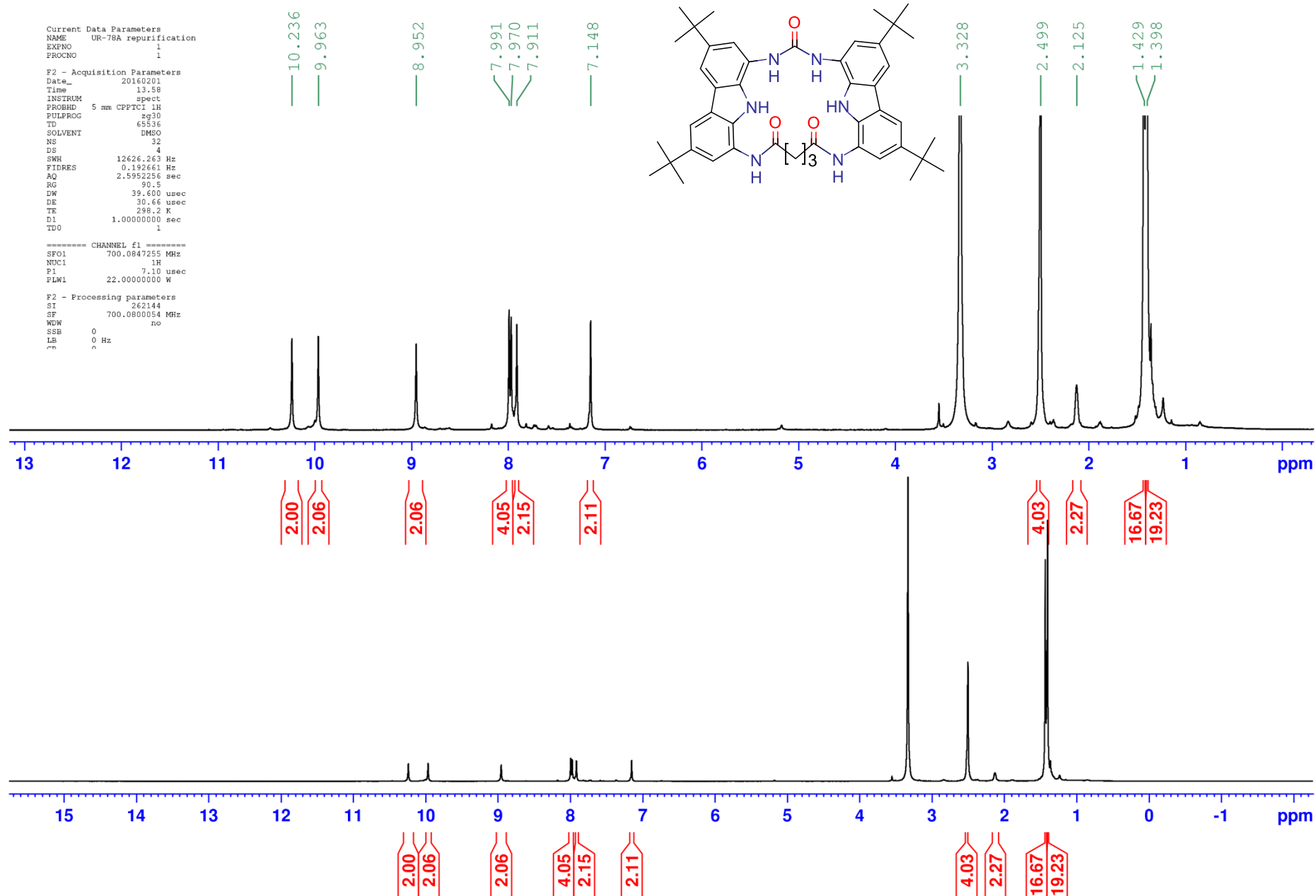
^1H - ^{13}C HMBC spectrum (700.1 MHz) of compound **MC002**



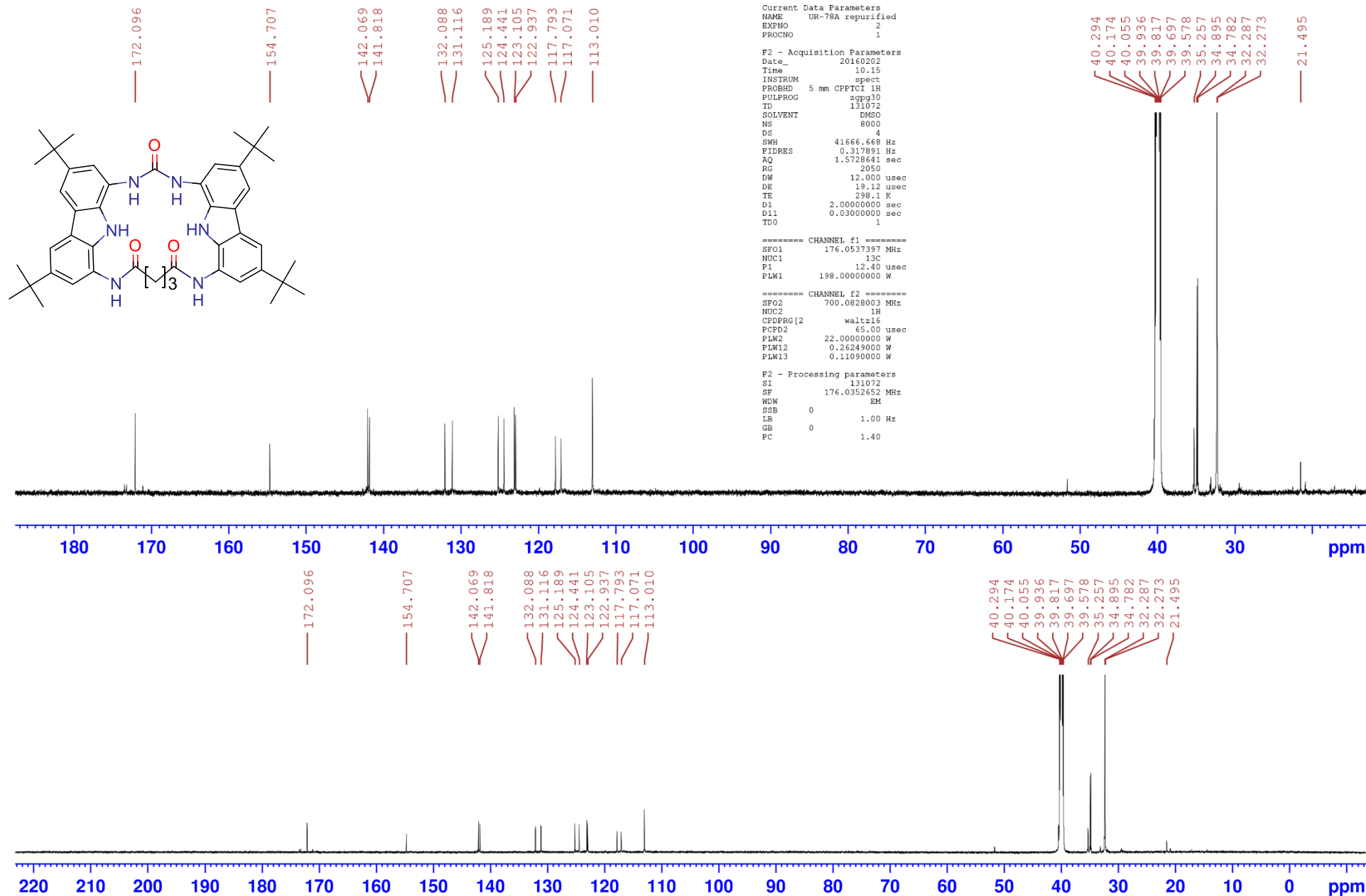
HRMS spectrum of compound **MC002**



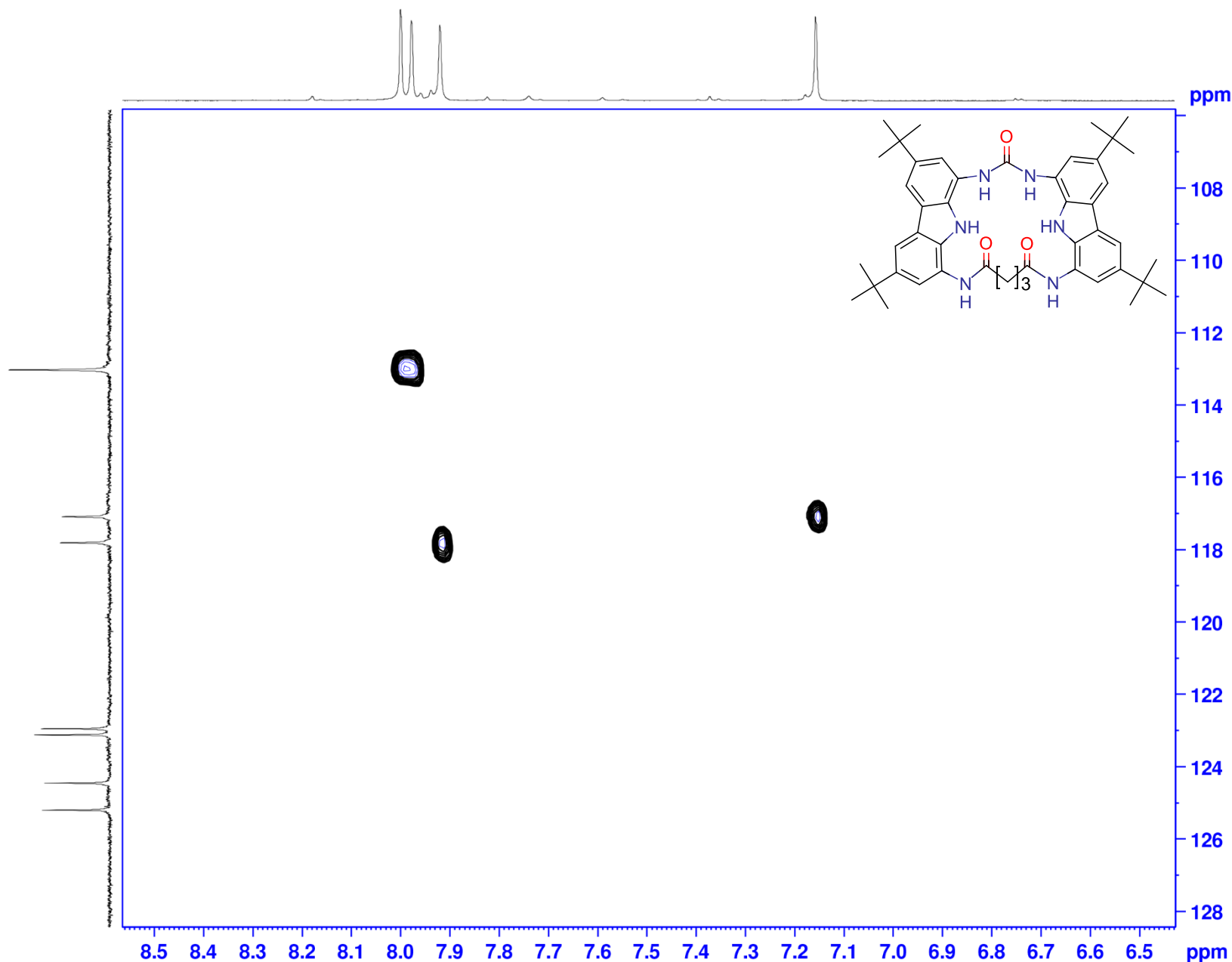
¹H NMR spectrum (700.1 MHz) of compound **MC003**



¹³C NMR spectrum (700.1 MHz) of compound **MC003**



^1H - ^{13}C HSQC spectrum (700.1 MHz) of compound **MC003**



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Current Data Parameters
NAME          UR-78A
EXPNO         4
PROCNO        1

F2 - Acquisition Parameters
Date_         20160602
Time          15.10
INSTRUM       spect
PROBHD        5 mm PABBO BB-
PULPROG       hsqcetdtp
TD            1024
SOLVENT       DMSO
NS            2
DS            16
SWH           7246.377 Hz
FIDRES        7.076540 Hz
AQ            0.0706560 sec
RG            2050
DW            69.000 usec
DE            6.50 usec
TE            298.2 K
CNST2         145.0000000
D0            0.00000300 sec
D1            1.50389099 sec
D4            0.00172414 sec
D11           0.03000000 sec
D13           0.00000400 sec
D16           0.00020000 sec
D21           0.00345000 sec
IN0           0.00001830 sec
ZGPGTNS

===== CHANNEL f1 =====
SFO1          700.0840045 MHz
NUC1           1H
P1            16.40 usec
P2            32.80 usec
P28           1000.00 usec
PLW1          26.00000000 W

===== CHANNEL f2 =====
SFO2          176.0541798 MHz
NUC2           13C
CPDPRG[2]     garp
P3            10.00 usec
P4            20.00 usec
PCPD2         55.00 usec
PLW2          60.00000000 W
PLW12         1.98350000 W

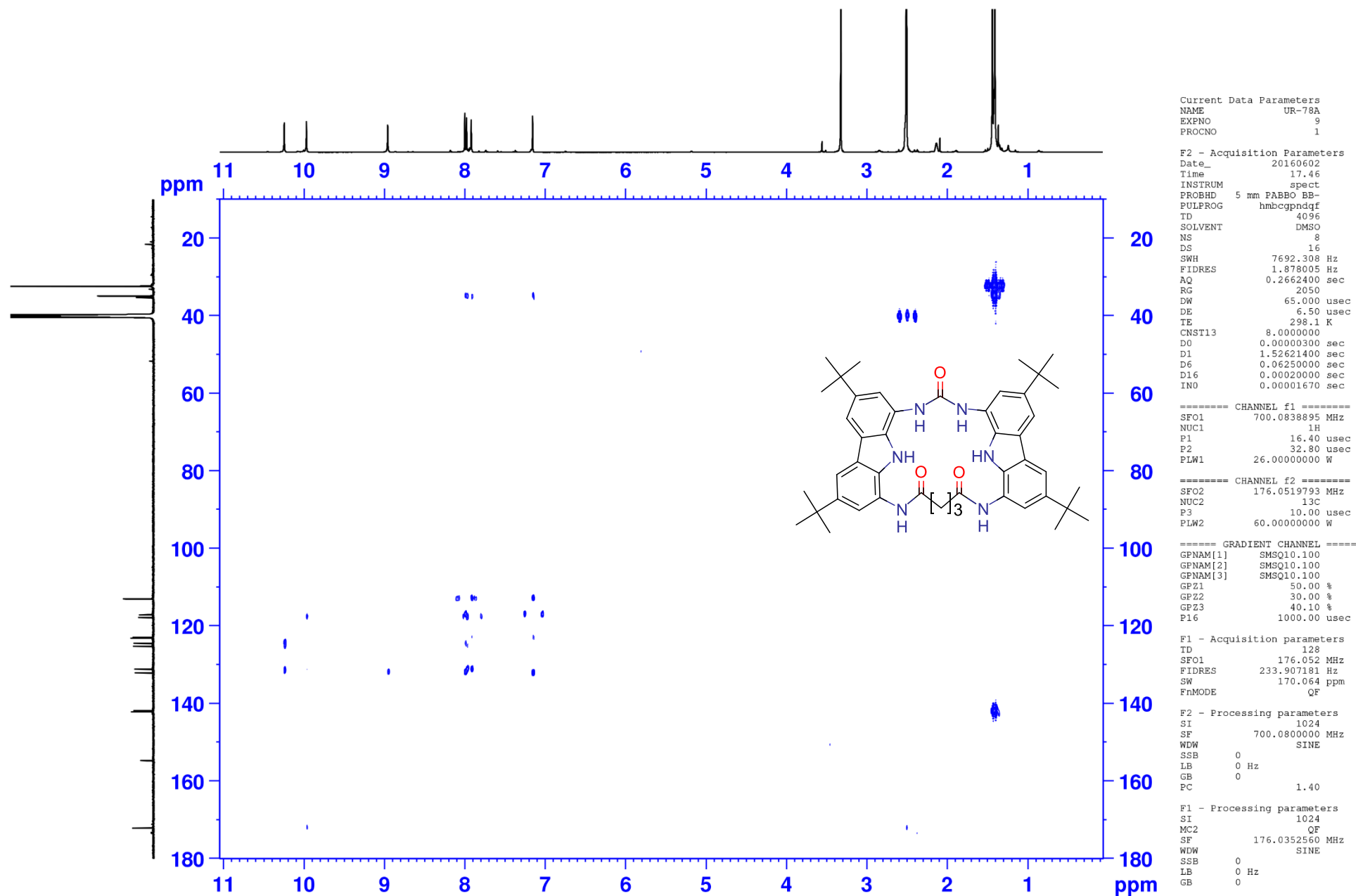
===== GRADIENT CHANNEL =====
GPNAM[1]      SMSQ10.100
GPNAM[2]      SMSQ10.100
GPZ1          80.00 %
GPZ2          20.10 %
P16           1000.00 usec

F1 - Acquisition parameters
TD            256
SFO1          176.0542 MHz
FIDRES        106.728142 Hz
SW            155.193 ppm
FnMODE        Echo-Antiecho

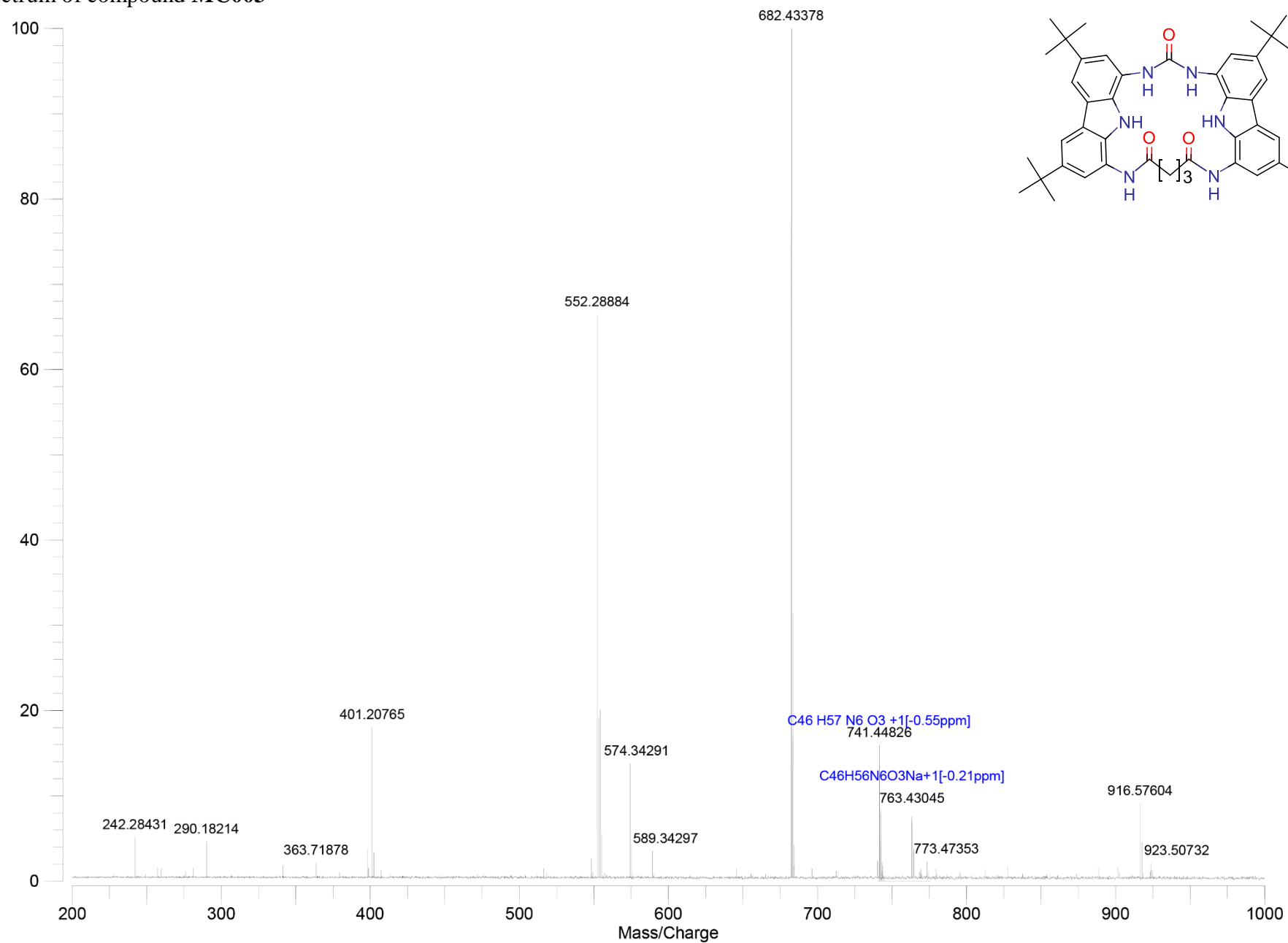
F2 - Processing parameters
SI            1024
SF            700.0800000 MHz
WDW           QSINE
SSB           2
LB            0 Hz
GB            0
PC            1.40

F1 - Processing parameters
SI            1024
MC2           echo-antiecho
SF            176.0352560 MHz
WDW           QSINE
SSB           2
LB            0 Hz
GB            0
  
```

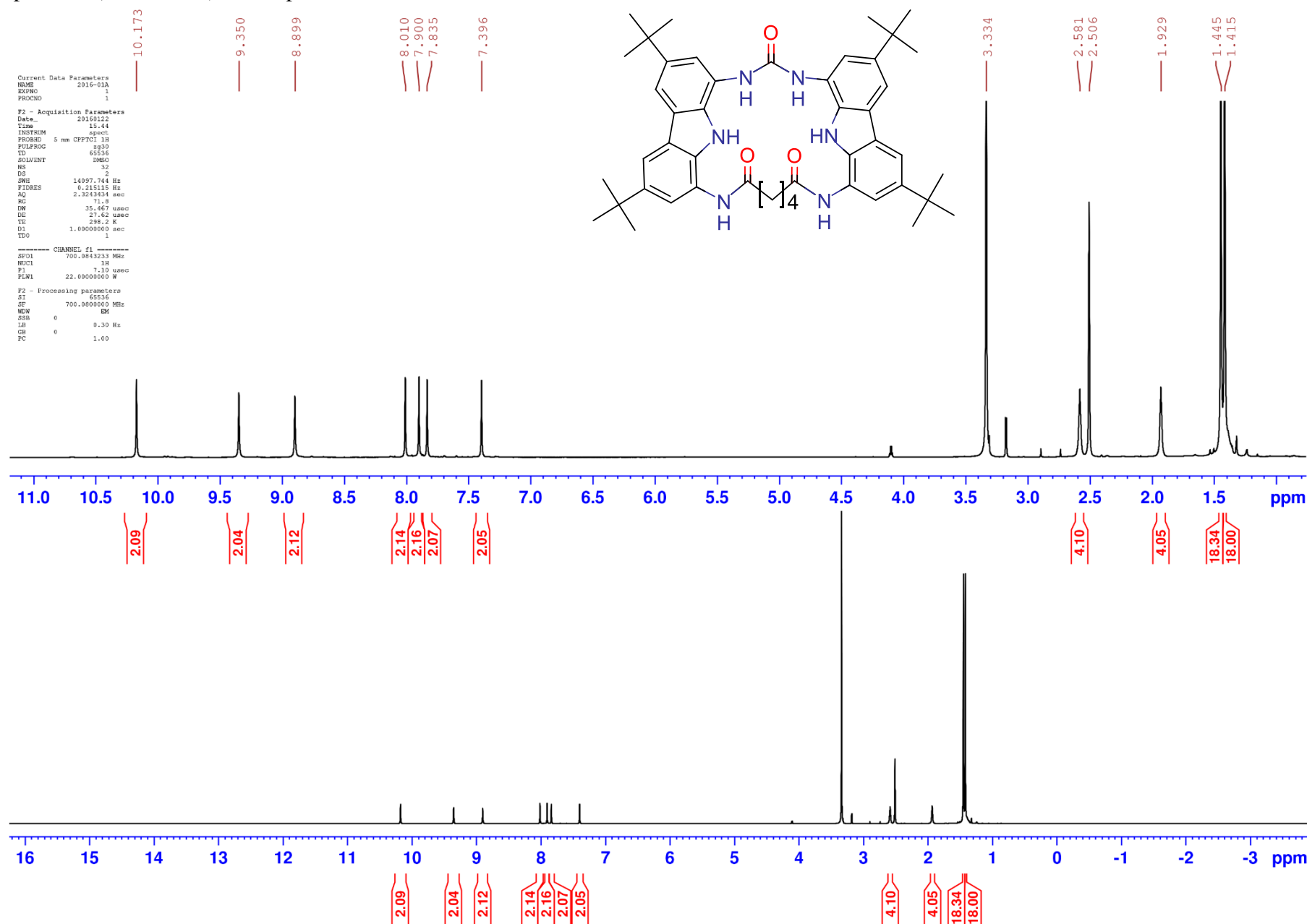

^1H - ^{13}C HMBC spectrum (700.1 MHz) of compound **MC003**



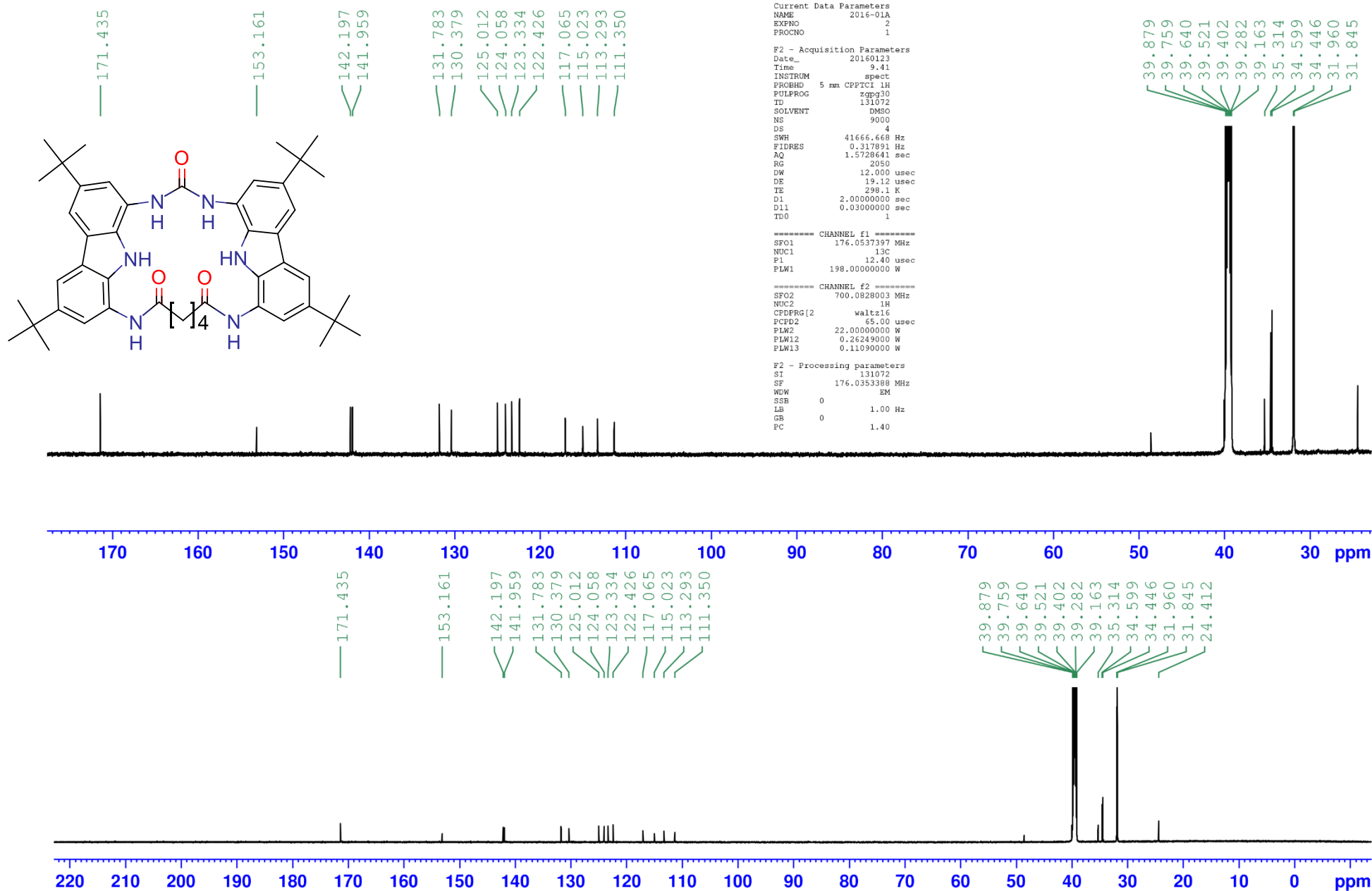
HRMS spectrum of compound MC003



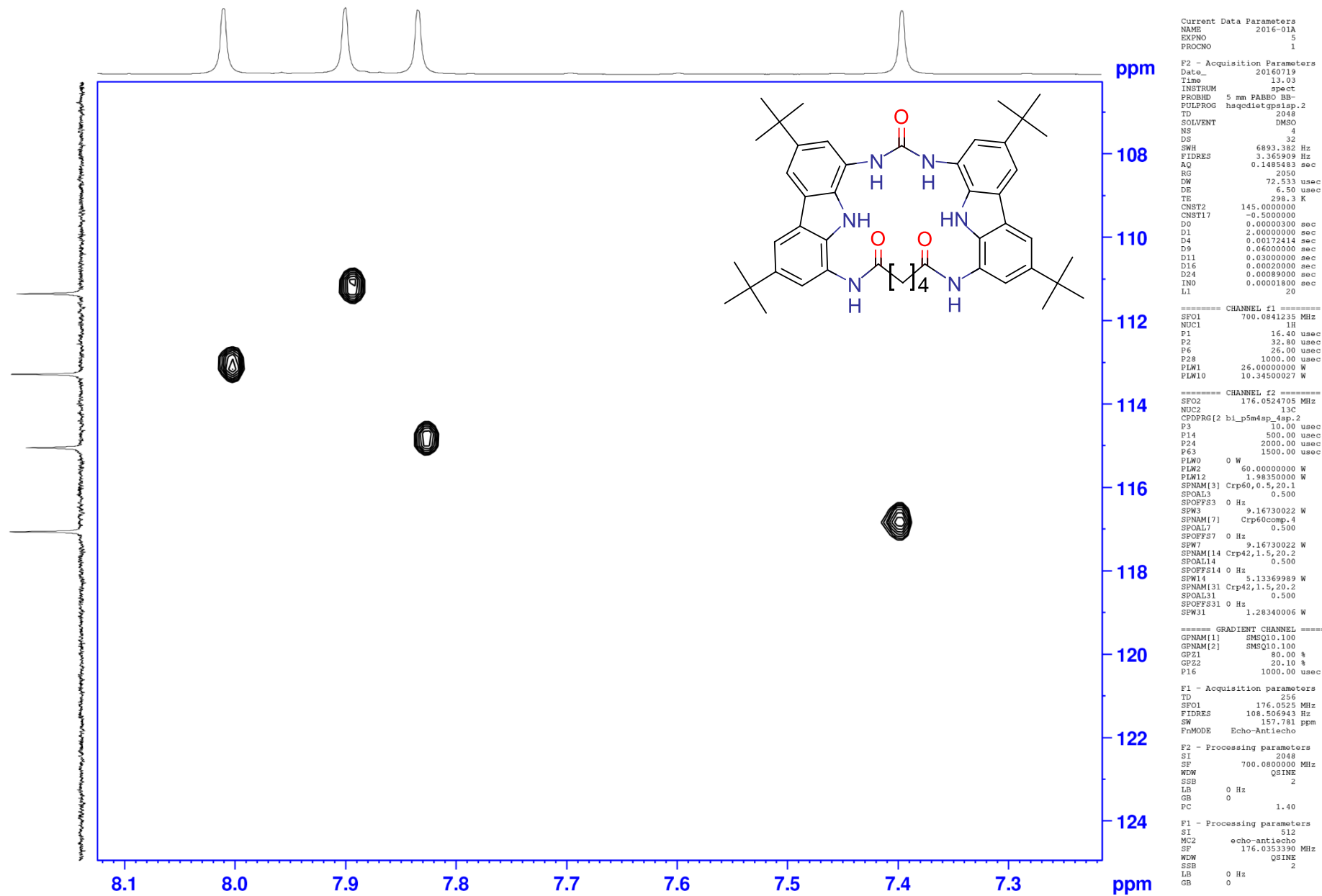
¹H NMR spectrum (700.1 MHz) of compound **MC004**



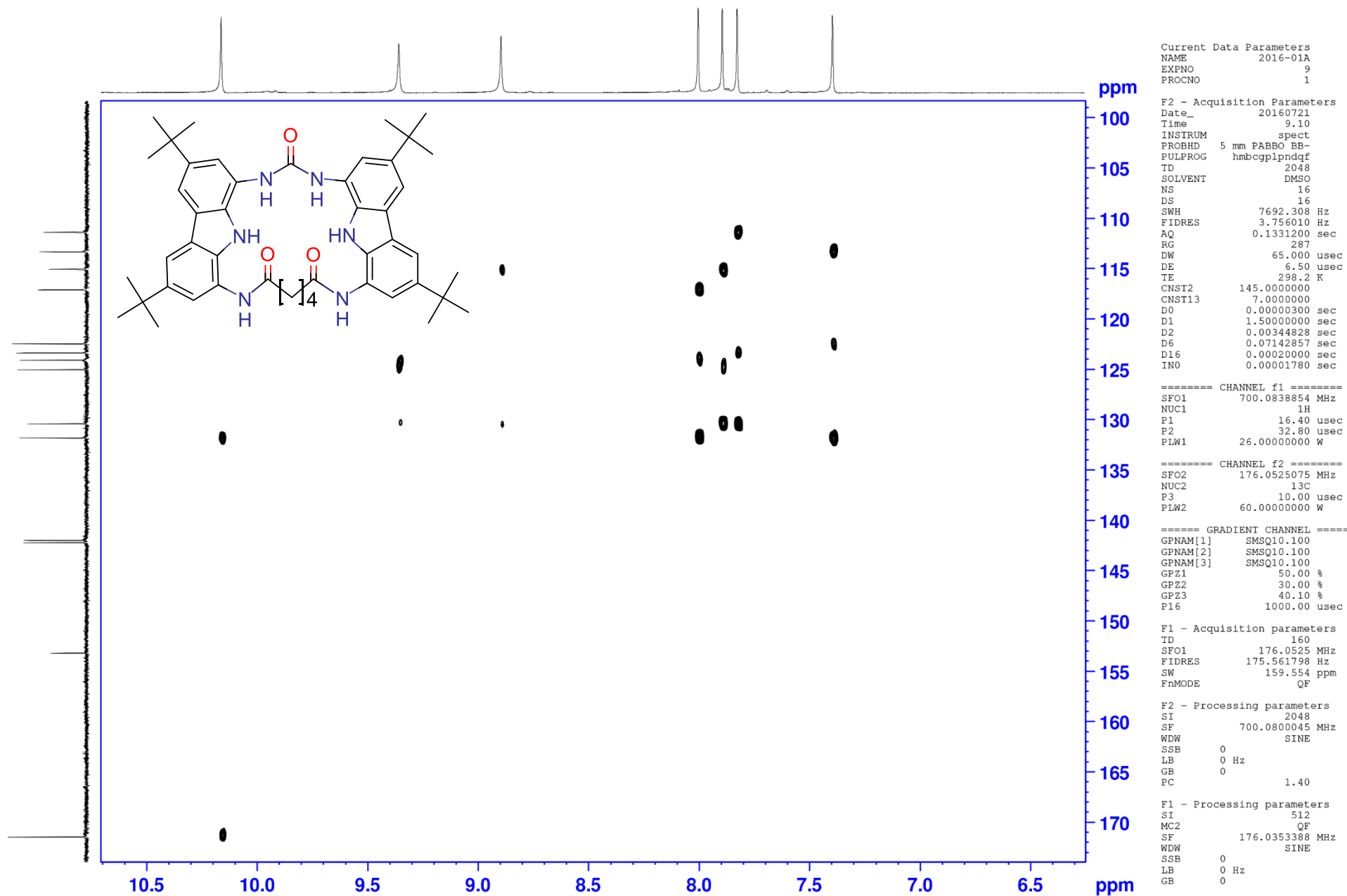
¹³C NMR spectrum (700.1 MHz) of compound **MC004**



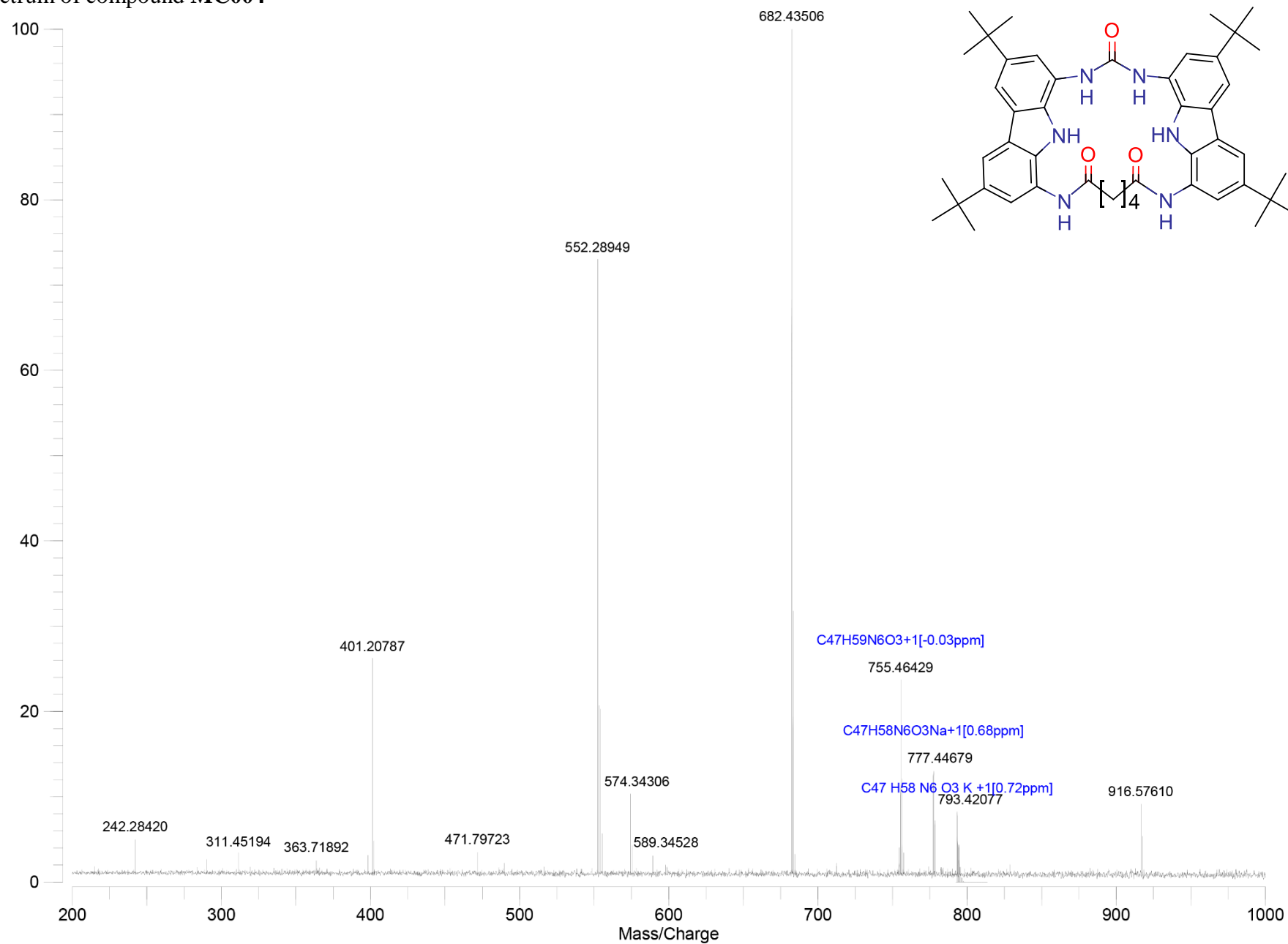
^1H - ^{13}C HSQC spectrum (700.1 MHz) of compound **MC004**



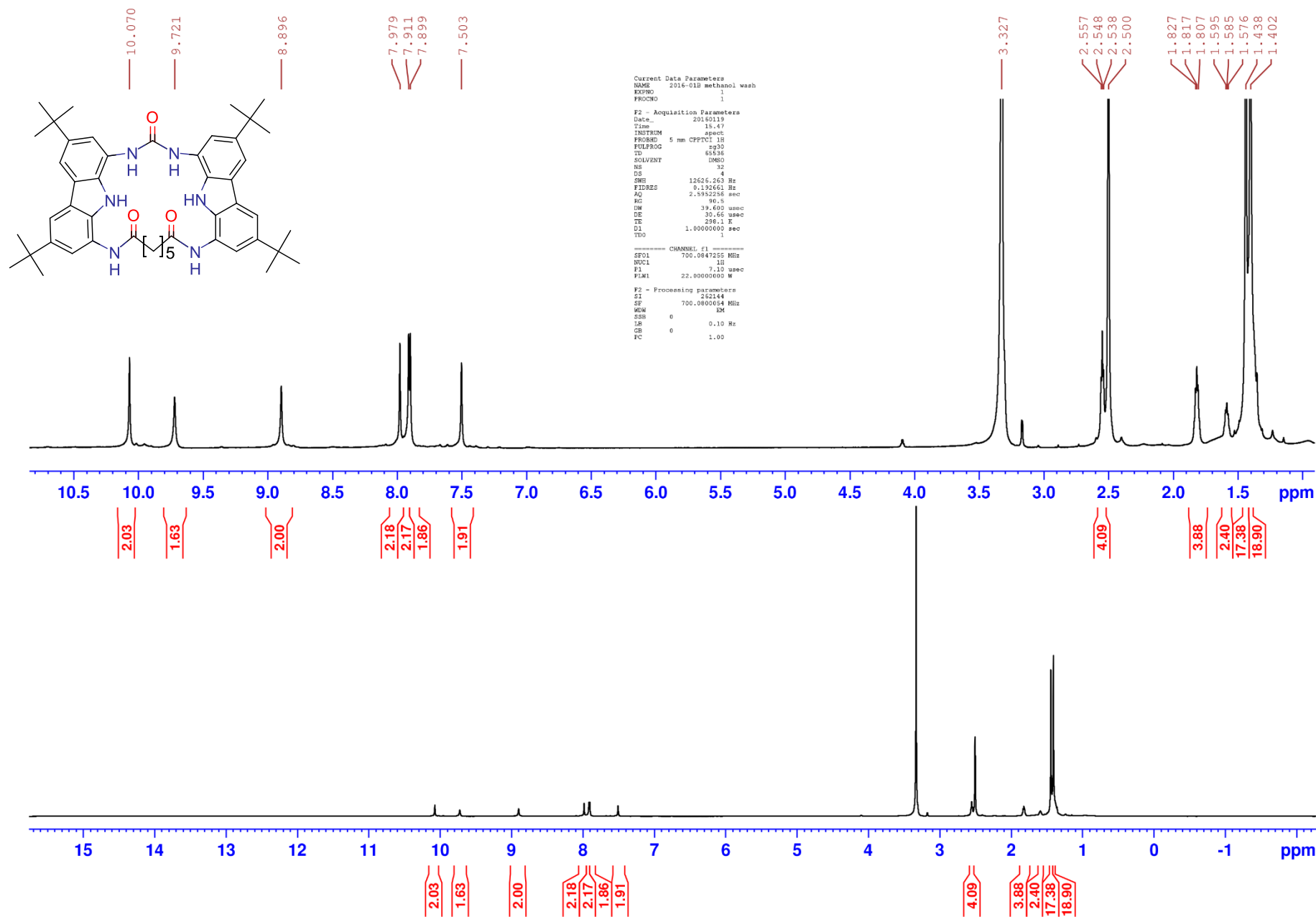
^1H - ^{13}C HMBC spectrum (700.1 MHz) of compound **MC004**



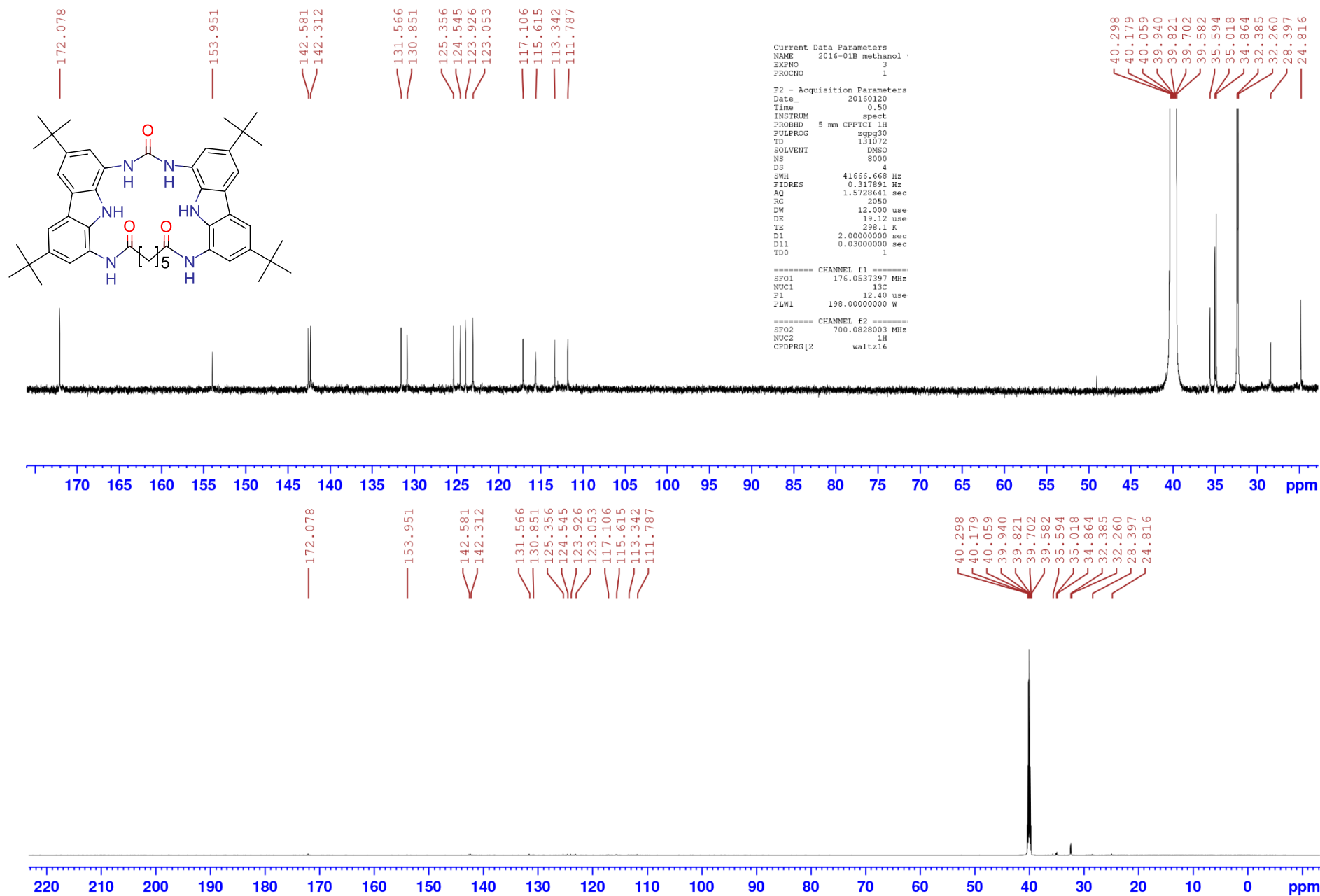
HRMS spectrum of compound MC004



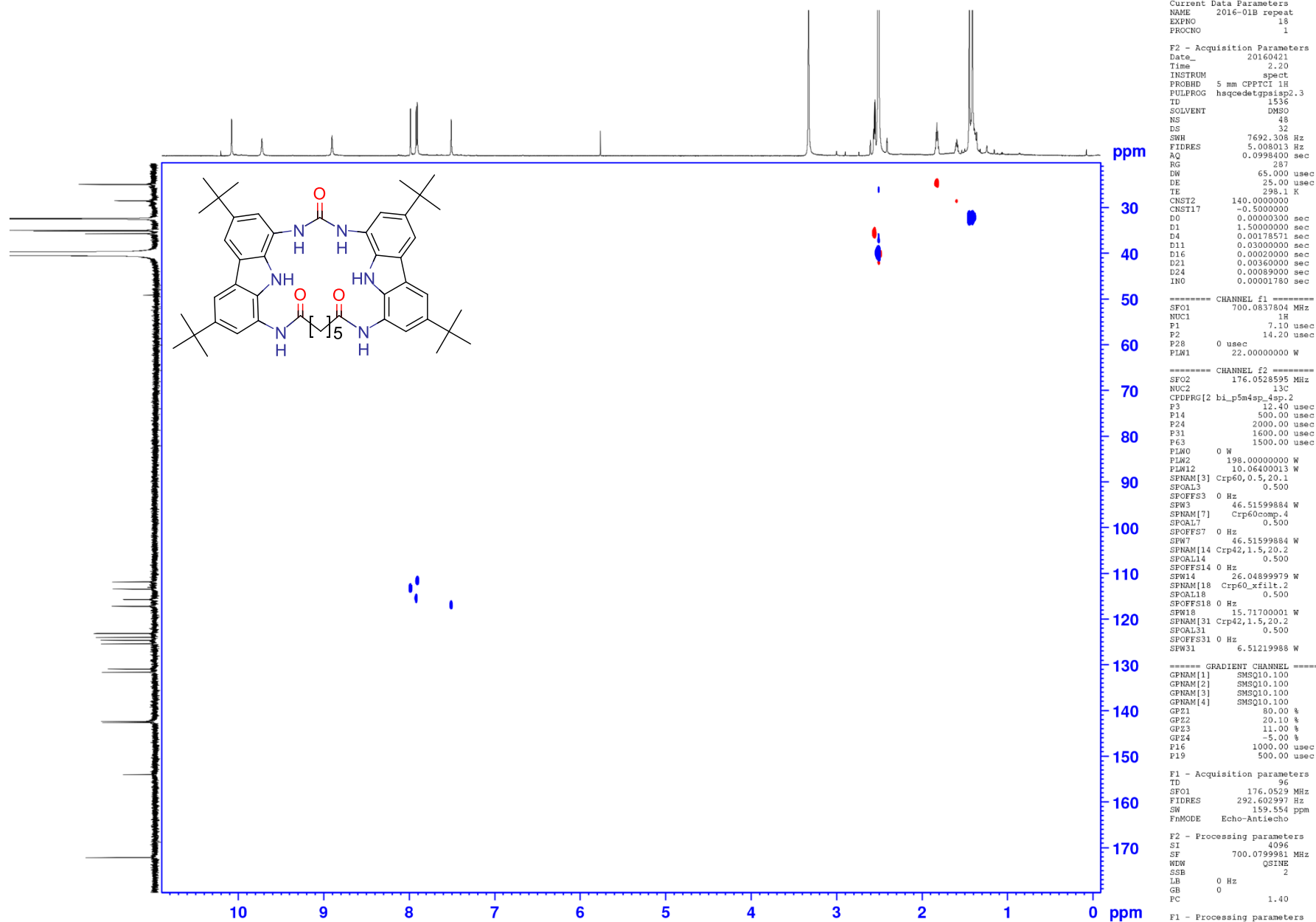
¹H NMR spectrum (700.1 MHz) of compound **MC005**



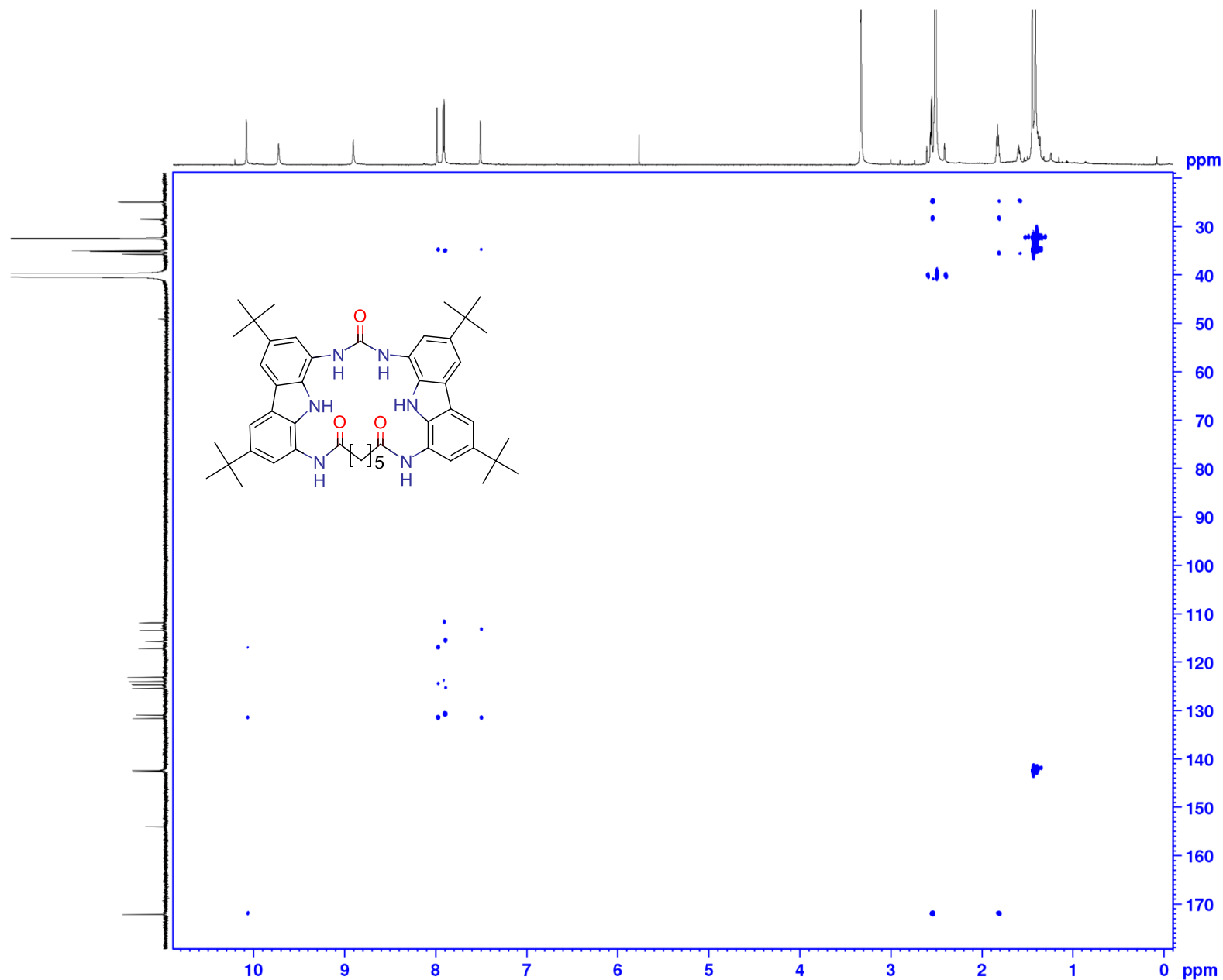
¹³C NMR spectrum (700.1 MHz) of compound **MC005**



^1H - ^{13}C HSQC spectrum (700.1 MHz) of compound **MC005**



^1H - ^{13}C HMBC spectrum (700.1 MHz) of compound **MC005**



Current Data Parameters
NAME 2016-01B repeat
EXPNO 17
PROCNO 1

F2 - Acquisition Parameters
Date_ 20160420
Time 19.08
INSTRUM spect
PROBHD 5 mm CPPTCI 1H
PULPROG hmbcgp1pndqf
TD 3072
SOLVENT DMSO
NS 64
DS 16
SWH 7692.308 Hz
FIDRES 2.504006 Hz
AQ 0.1996800 sec
RG 812
DW 65.000 usec
DE 25.00 usec
TE 298.2 K
CNST2 145.0000000
CNST13 8.0000000
D0 0.00000300 sec
D1 1.29999995 sec
D2 0.00344828 sec
D6 0.06250000 sec
D16 0.00020000 sec
INO 0.00001770 sec

===== CHANNEL f1 =====
SFO1 700.0837804 MHz
NUC1 1H
P1 7.10 usec
P2 14.20 usec
PLW1 22.00000000 W

===== CHANNEL f2 =====
SFO2 176.0526835 MHz
NUC2 13C
P3 12.40 usec
PLW2 198.00000000 W

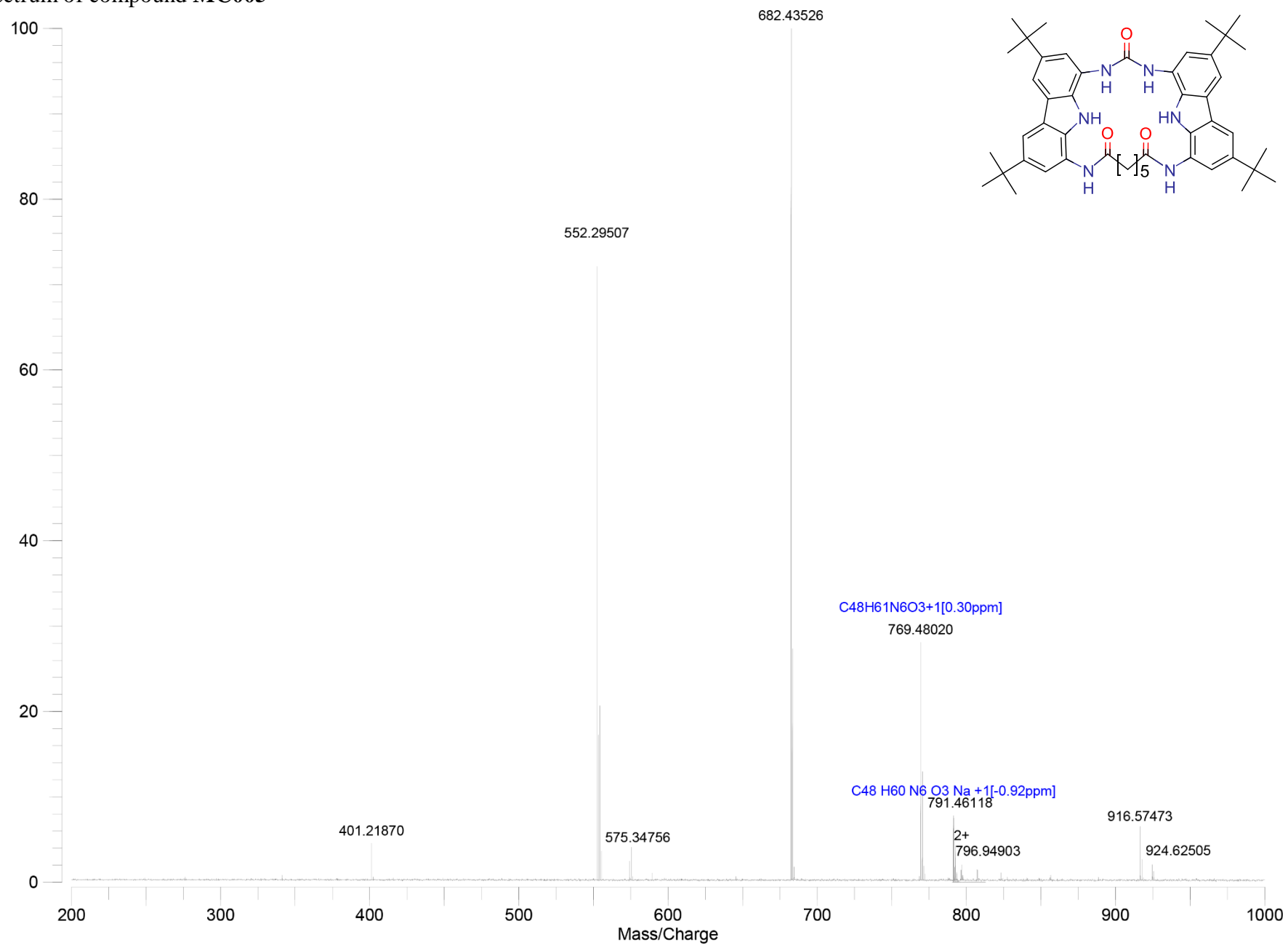
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GPNAM[2] SMSQ10.100
GPNAM[3] SMSQ10.100
GPZ1 50.00 %
GPZ2 30.00 %
GPZ3 40.10 %
FIG 1000.00 usec

F1 - Acquisition parameters
TD 256
SFO1 176.0527 MHz
FIDRES 110.346046 Hz
SW 160.455 ppm
FnMODE QF

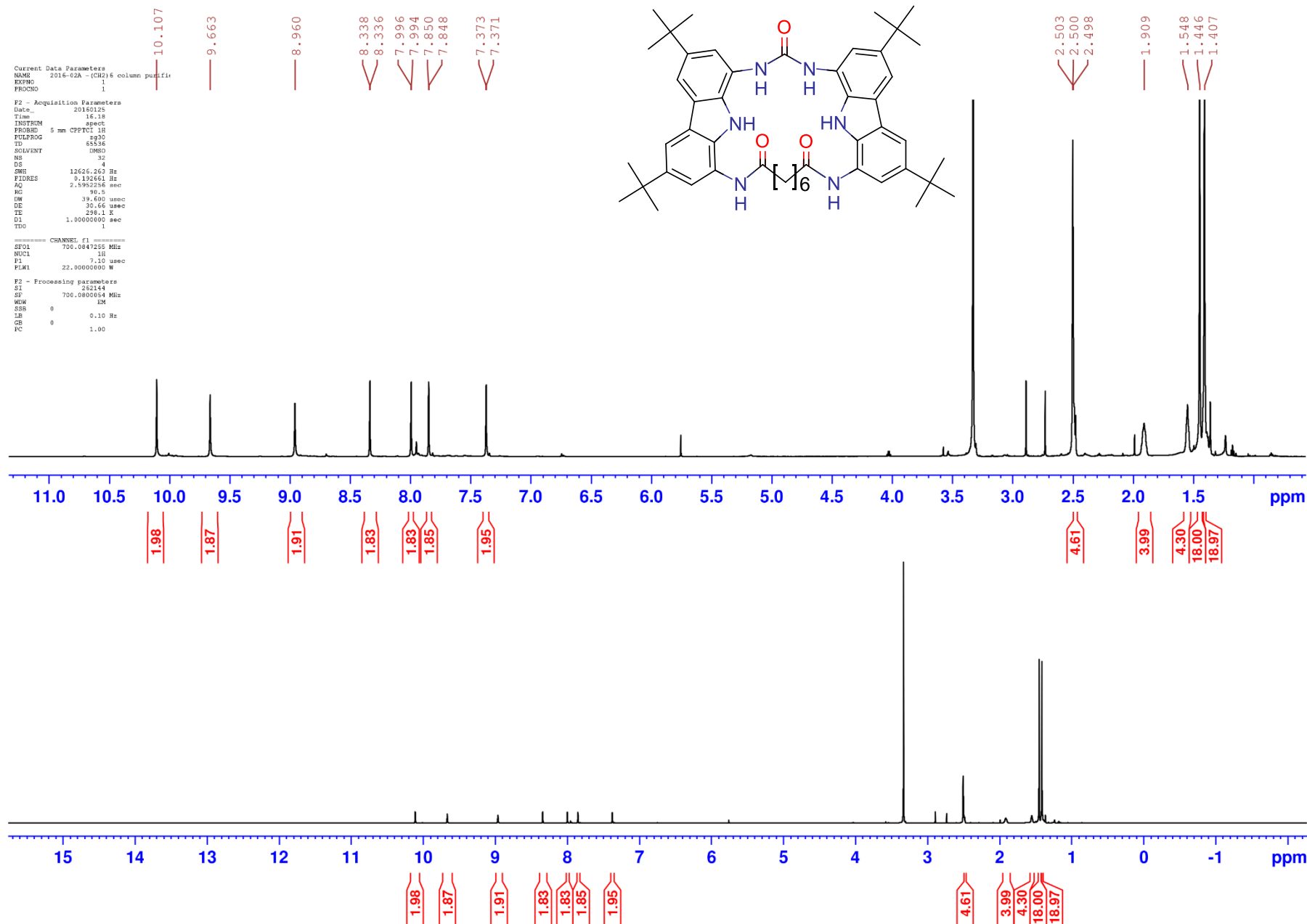
F2 - Processing parameters
SI 4096
SF 700.0800053 MHz
WDW SINE
SSB 0
LB 0 Hz
GB 0
PC 1.40

F1 - Processing parameters
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MC2 QF
SF 176.0352560 MHz
WDW SINE
SSB 0
LB 0 Hz
GB 0

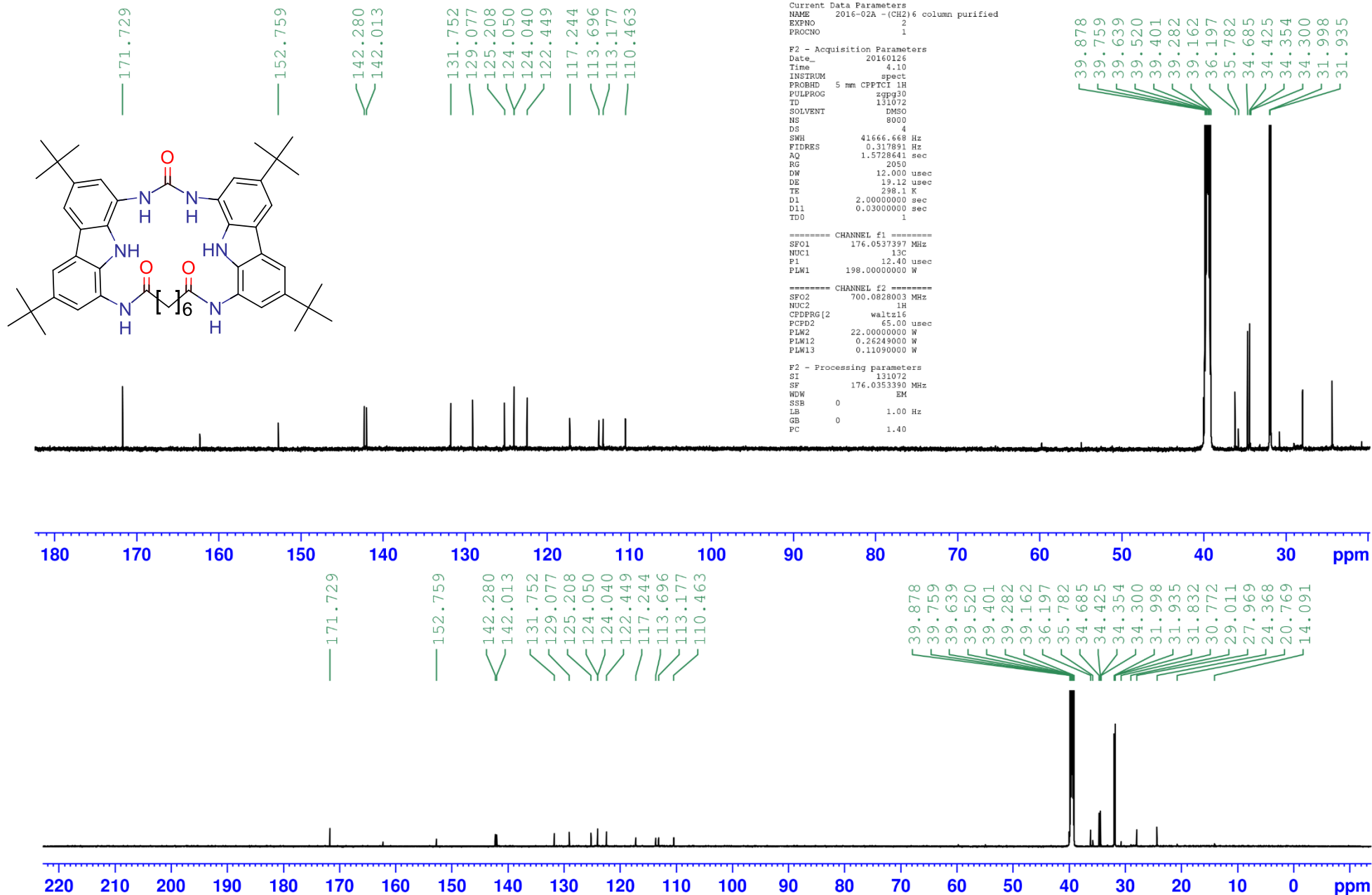
HRMS spectrum of compound **MC005**



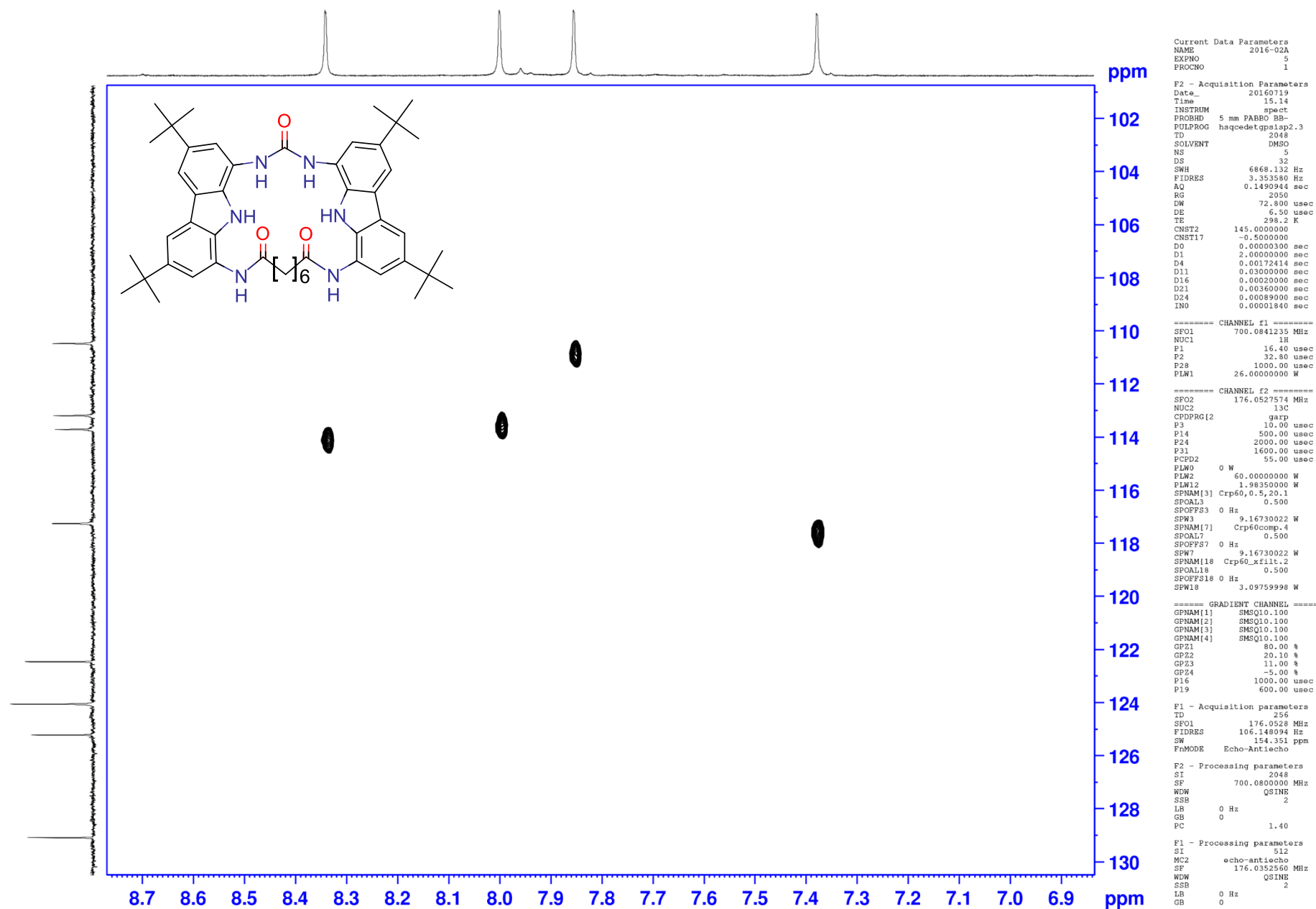
¹H NMR spectrum (700.1 MHz) of compound **MC006**



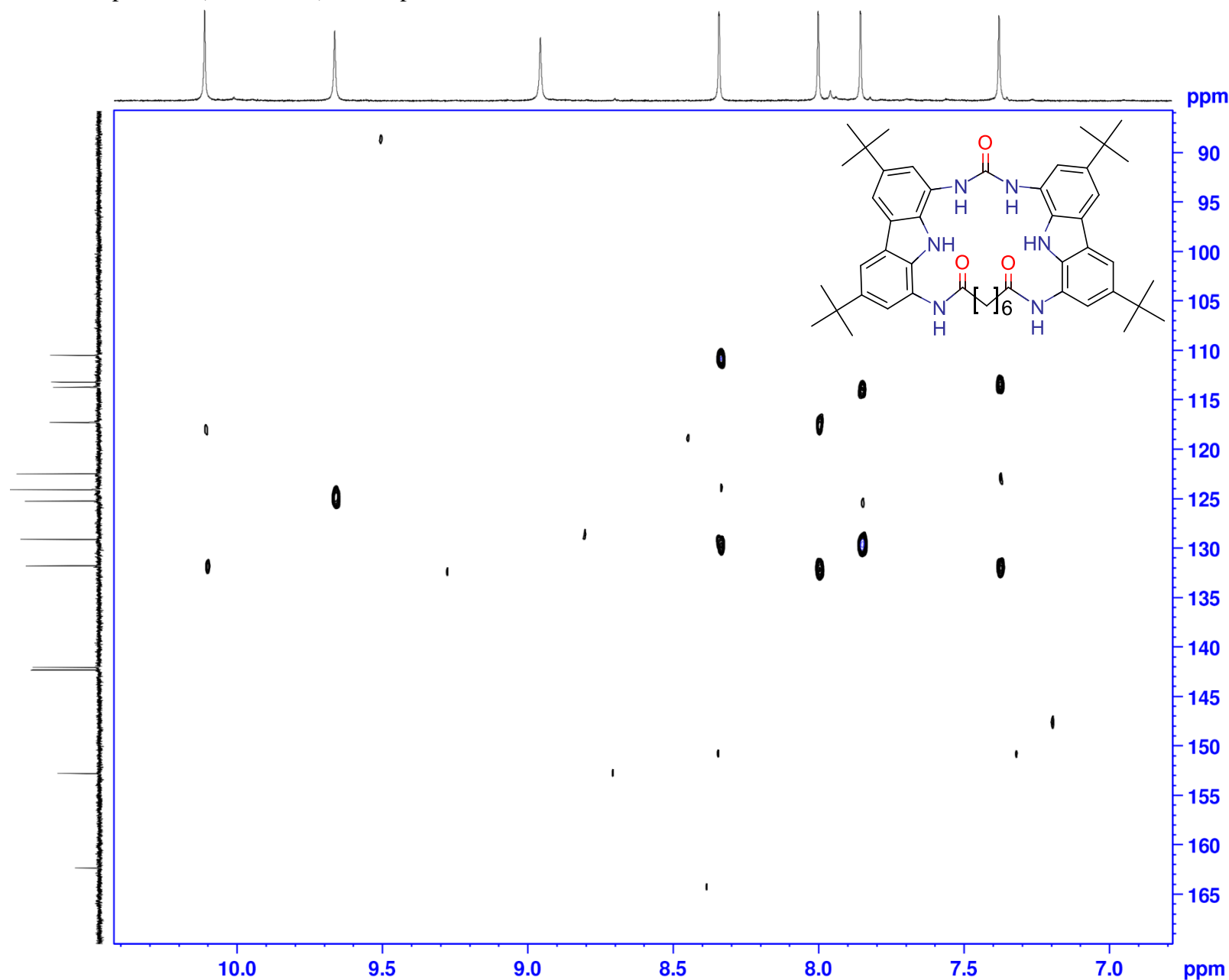
¹³C NMR spectrum (700.1 MHz) of compound **MC006**



^1H - ^{13}C HSQC spectrum (700.1 MHz) of compound **MC006**



^1H - ^{13}C HMBC spectrum (700.1 MHz) of compound **MC006**



Current Data Parameters
NAME 2016-02A
EXPNO 6
PROCNO 1

F2 - Acquisition Parameters
Date_ 20160719
Time 16.02
INSTRUM spect
PROBHD 5 mm PABBO BB-
PULPROG hmbcgp1pndqf
TD 2048
SOLVENT DMSO
NS 16
DS 16
SWH 7142.857 Hz
FIDRES 3.487723 Hz
AQ 0.1433600 sec
RG 2050
DW 70.000 usec
DE 6.50 usec
TE 298.1 K
CNST2 145.0000000
CNST13 10.0000000
D0 0.00000300 sec
D1 1.50573397 sec
D2 0.00344828 sec
D6 0.05000000 sec
D16 0.00020000 sec
INO 0.00001840 sec

===== CHANNEL f1 =====
SFO1 700.0839485 MHz
NUC1 1H
P1 16.40 usec
P2 32.80 usec
PLW1 26.00000000 W

===== CHANNEL f2 =====
SFO2 176.0527574 MHz
NUC2 13C
P3 10.00 usec
PLW2 60.00000000 W

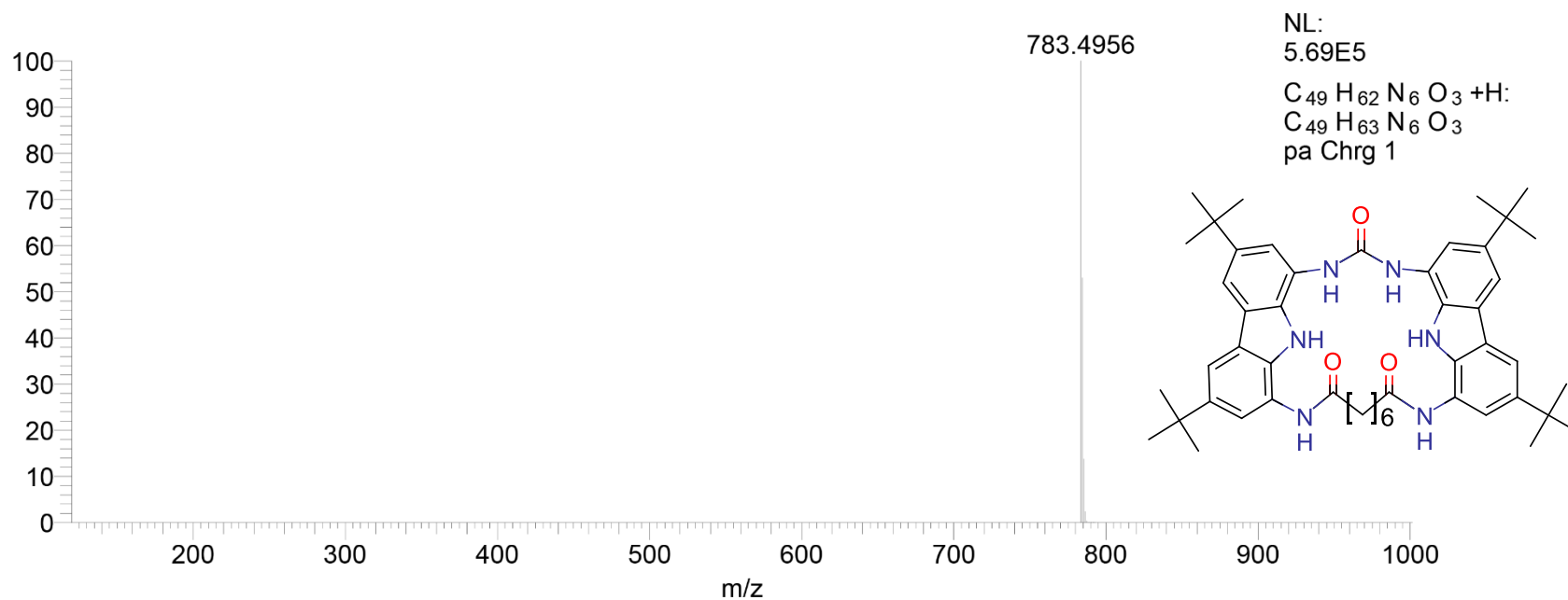
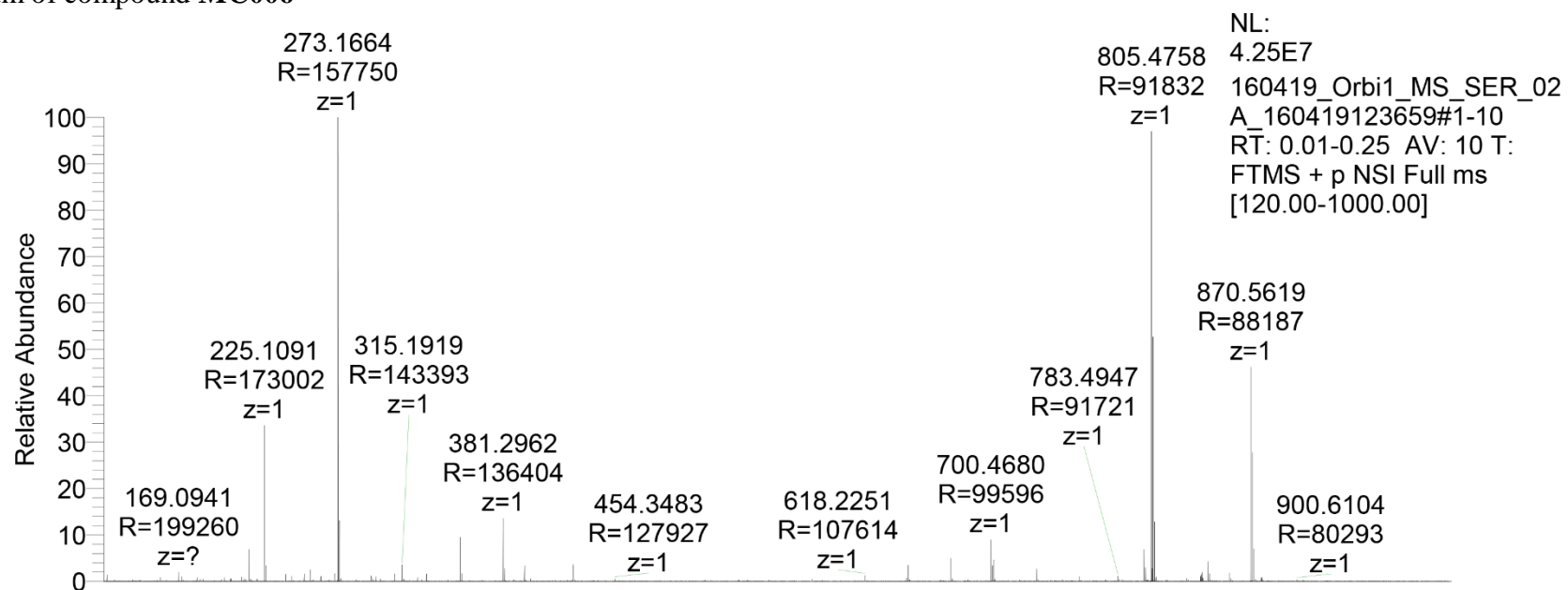
===== GRADIENT CHANNEL =====
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GPNAM[2] SMSQ10.100
GPNAM[3] SMSQ10.100
GPZ1 50.00 %
GPZ2 30.00 %
GPZ3 40.10 %
P16 1000.00 usec

F1 - Acquisition parameters
TD 87
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FIDRES 312.343842 Hz
SW 154.351 ppm
FnMODE QF

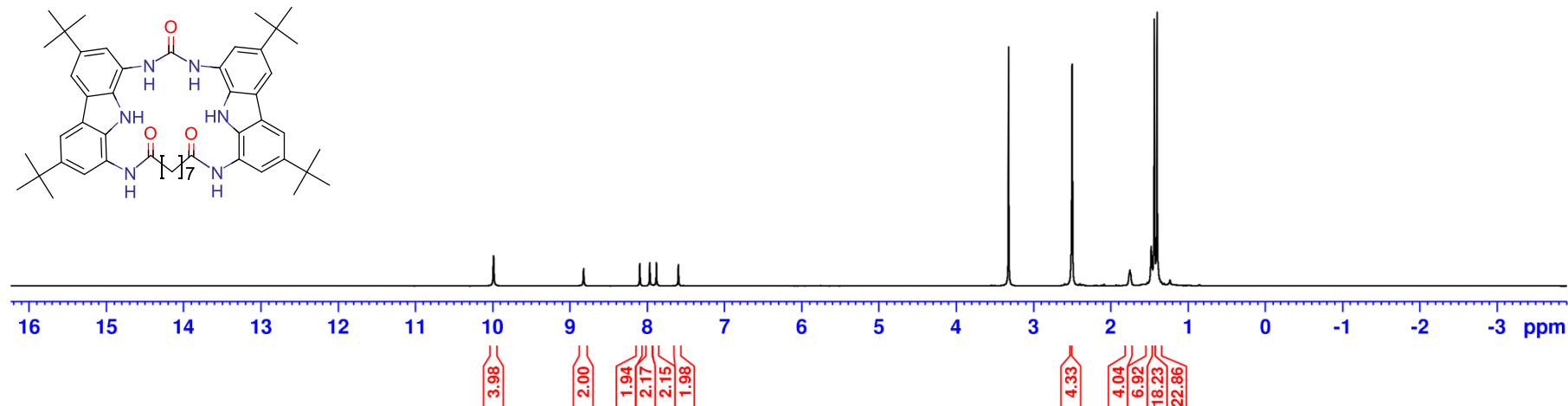
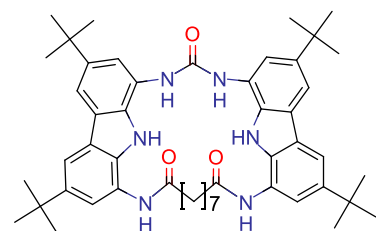
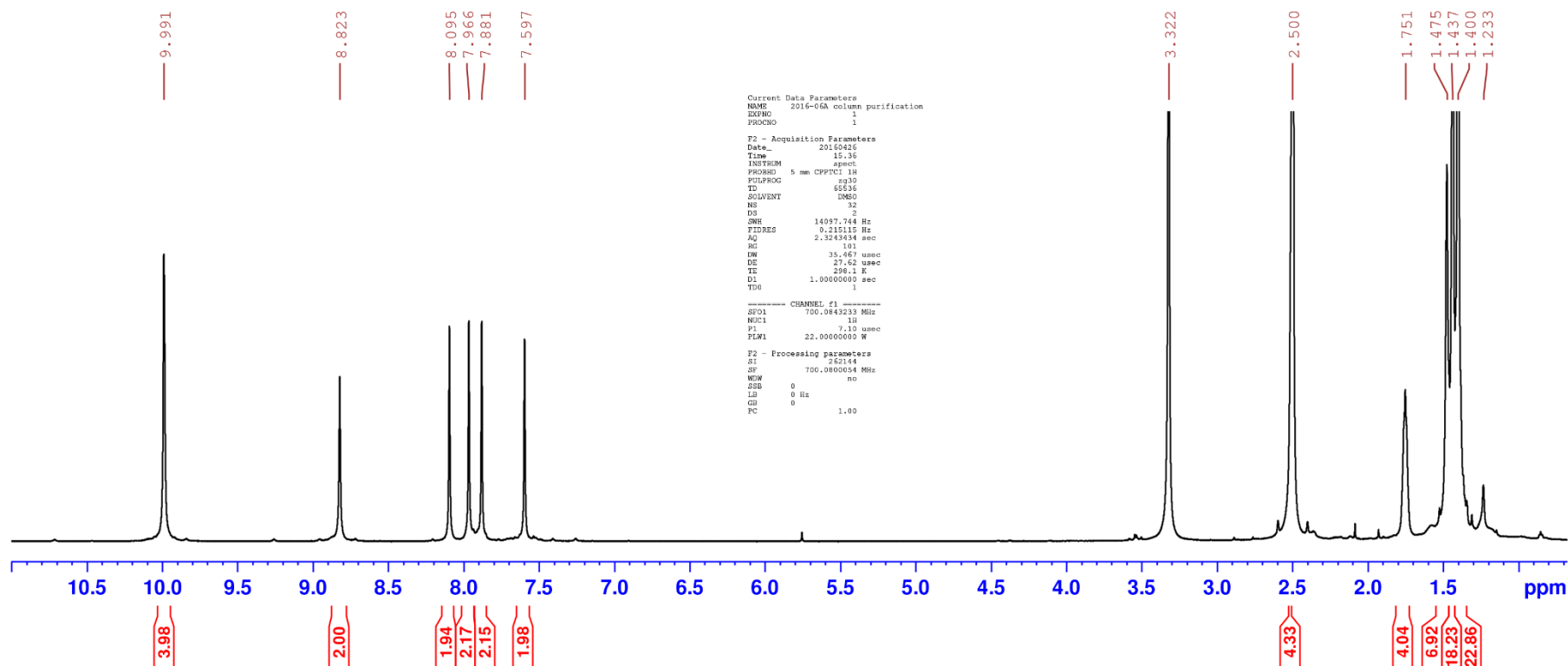
F2 - Processing parameters
SI 2048
SF 700.0800000 MHz
WDW SINE
SSB 0
LB 0 Hz
GB 0
PC 1.40

F1 - Processing parameters
SI 1024
MC2 QF
SF 176.0352560 MHz
WDW SINE
SSB 0
LB 0 Hz
GB 0

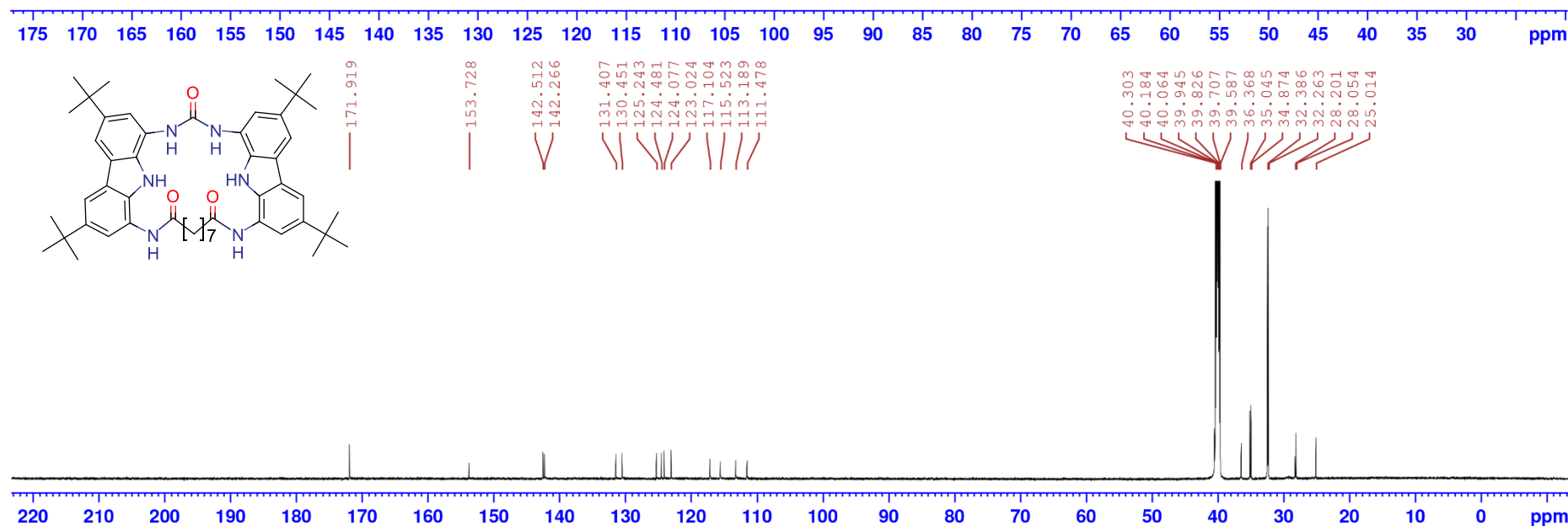
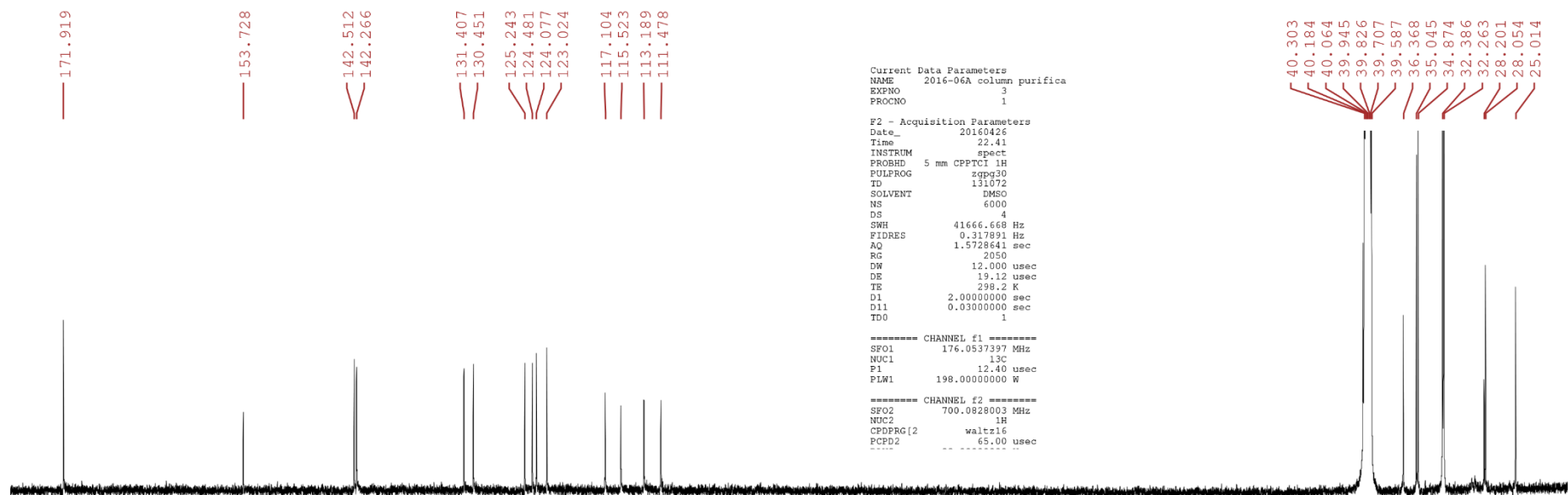
HRMS spectrum of compound MC006



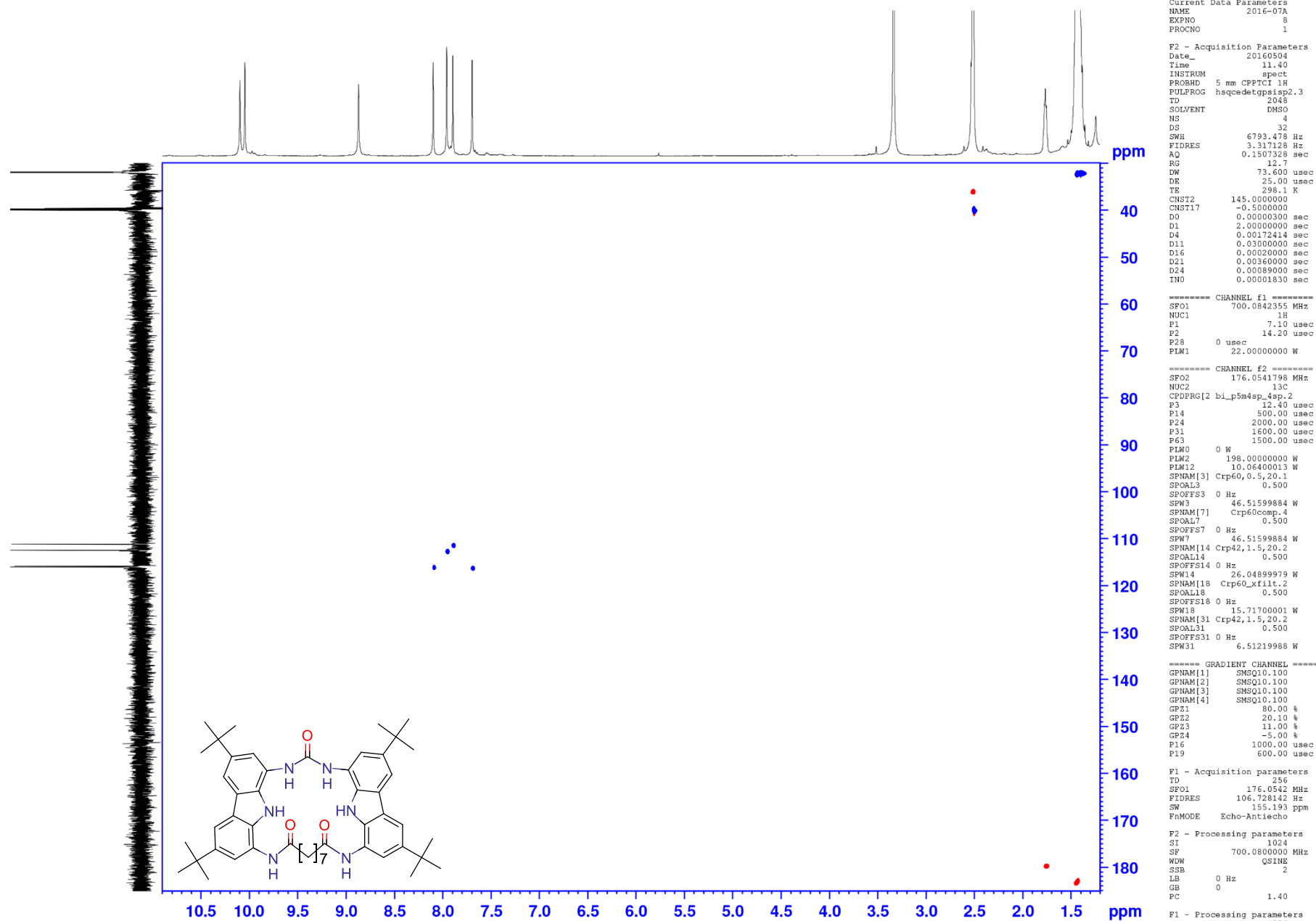
¹H NMR spectrum (700.1 MHz) of compound **MC007**



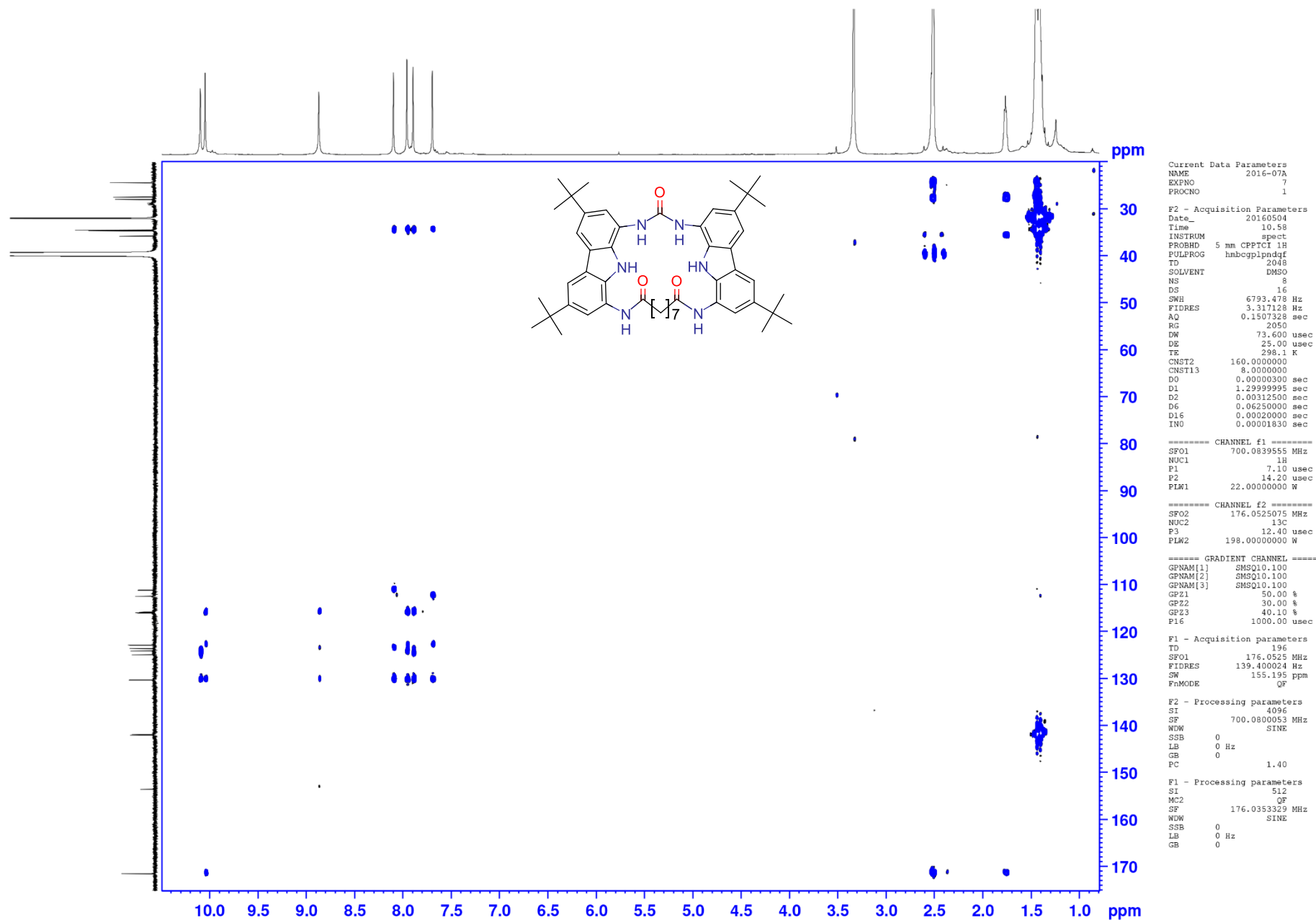
¹³C NMR spectrum (700.1 MHz) of compound **MC007**



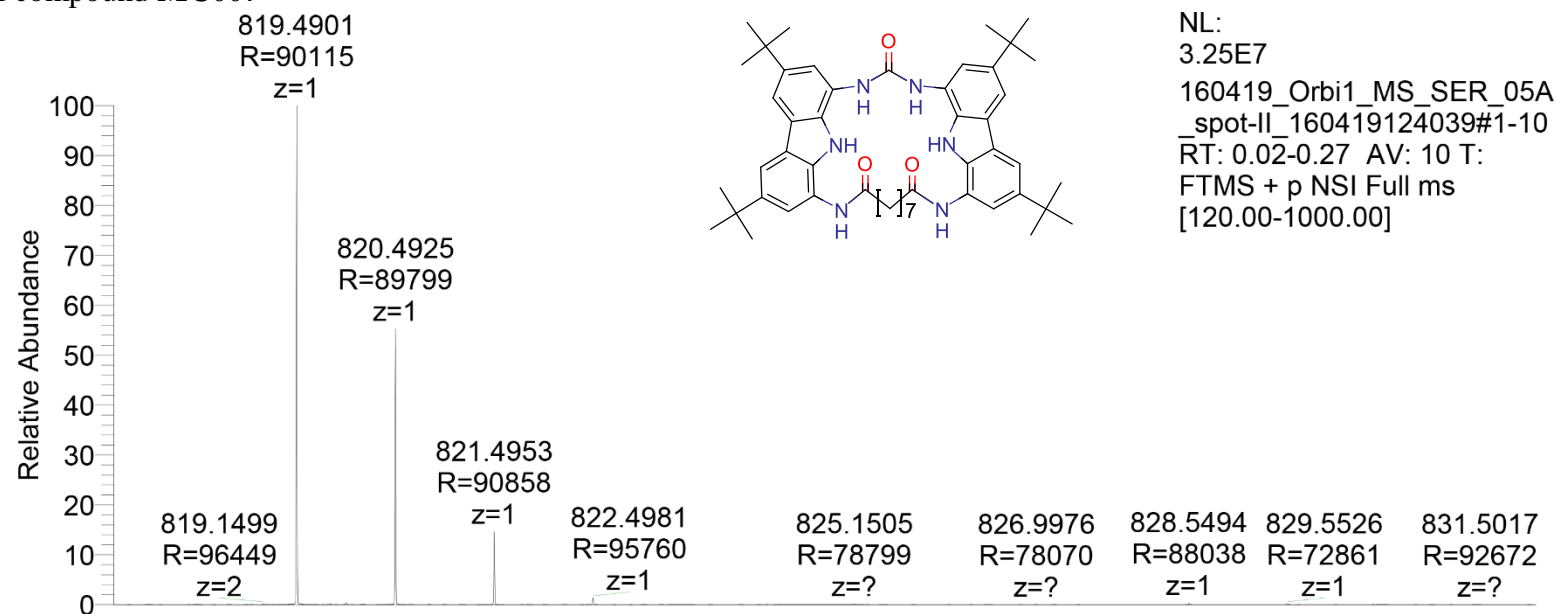
^1H - ^{13}C HSQC spectrum (700.1 MHz) of compound **MC007**



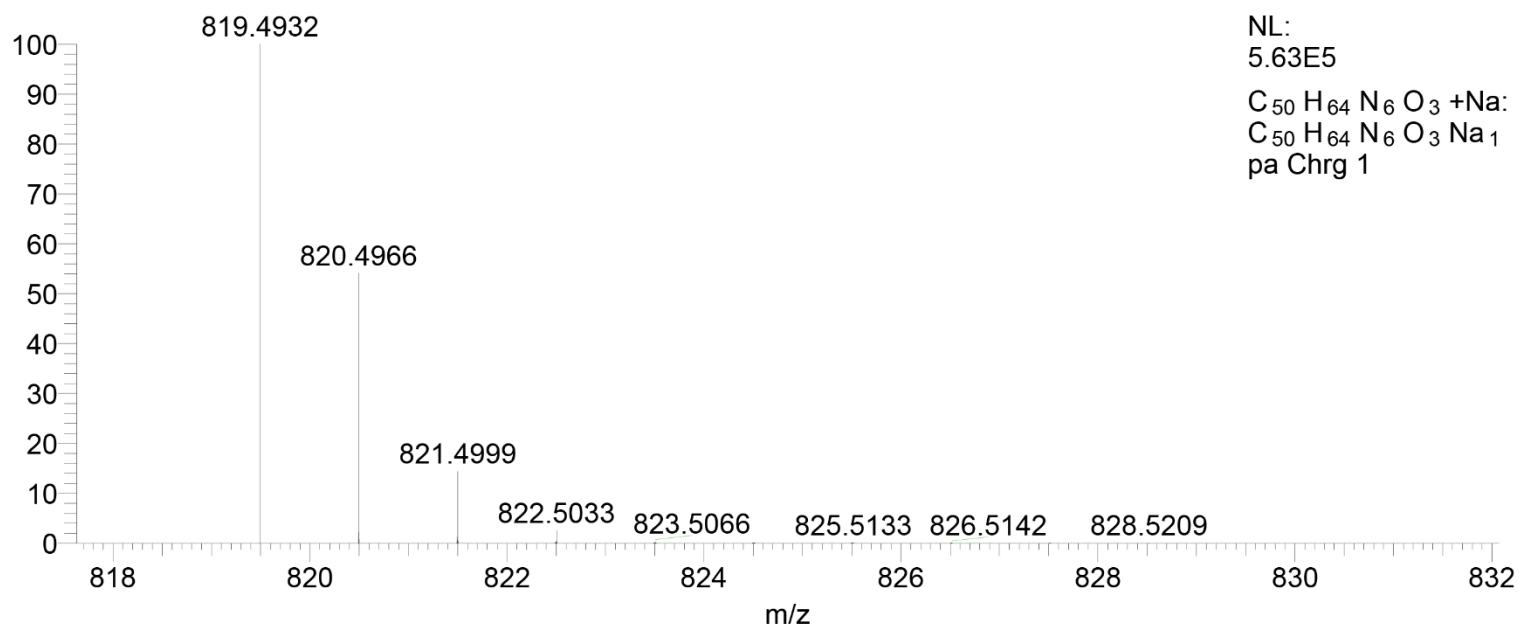
^1H - ^{13}C HMBC spectrum (700.1 MHz) of compound **MC007**



HRMS spectrum of compound **MC007**

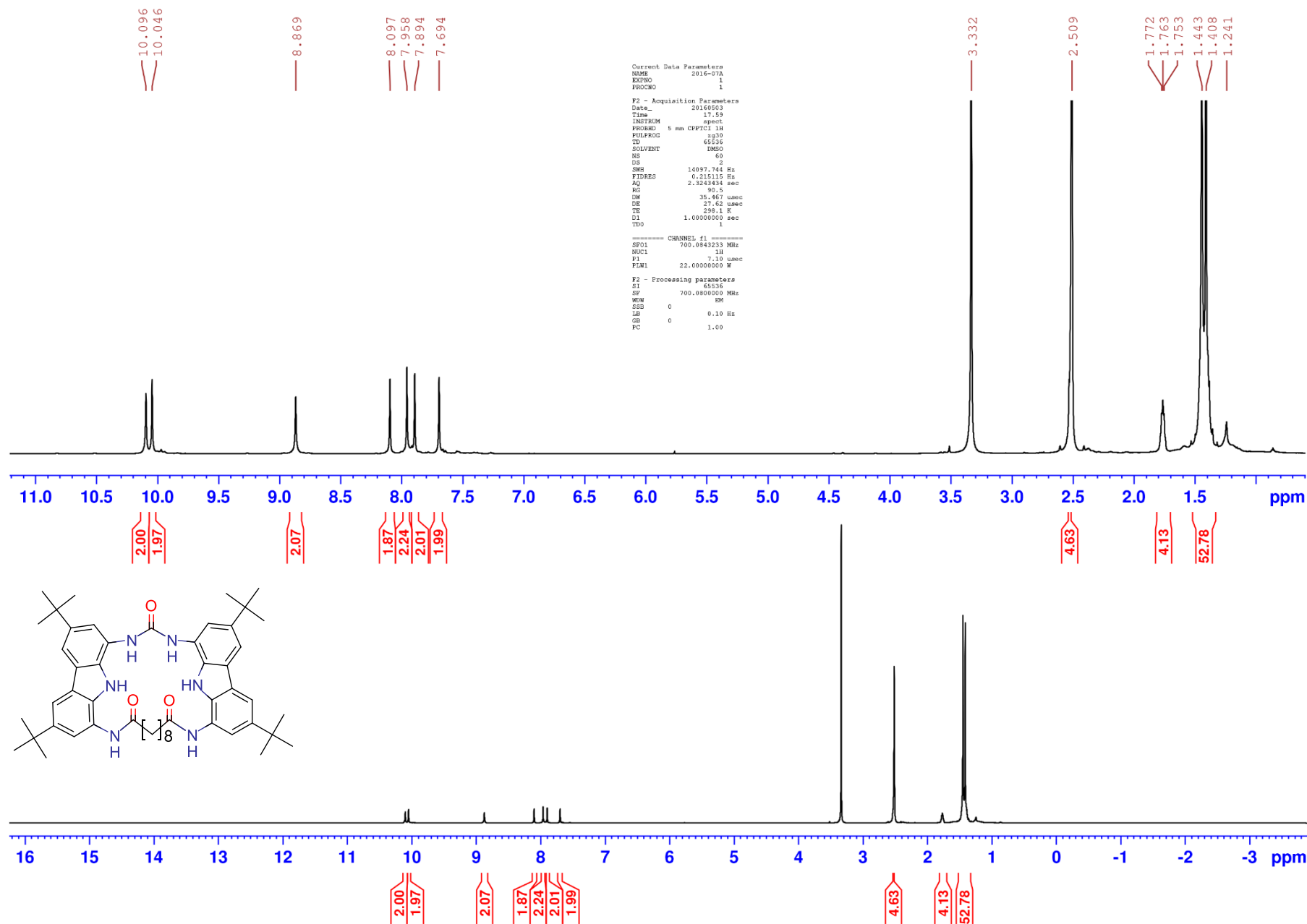


NL:
3.25E7
160419_Orbi1_MS_SER_05A
_spot-II_160419124039#1-10
RT: 0.02-0.27 AV: 10 T:
FTMS + p NSI Full ms
[120.00-1000.00]

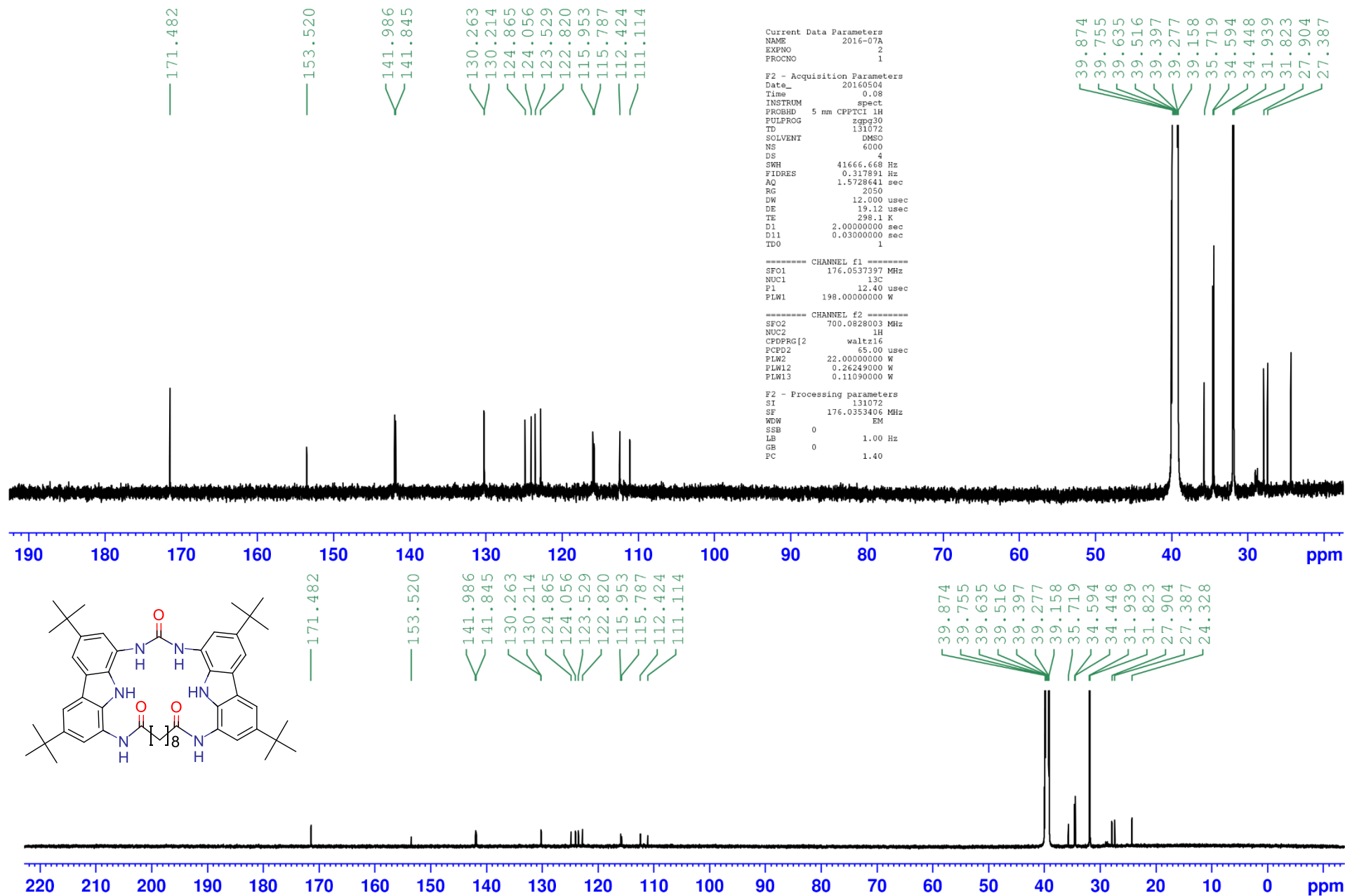


NL:
5.63E5
 $C_{50}H_{64}N_6O_3 + Na$:
 $C_{50}H_{64}N_6O_3Na_1$
pa Chrg 1

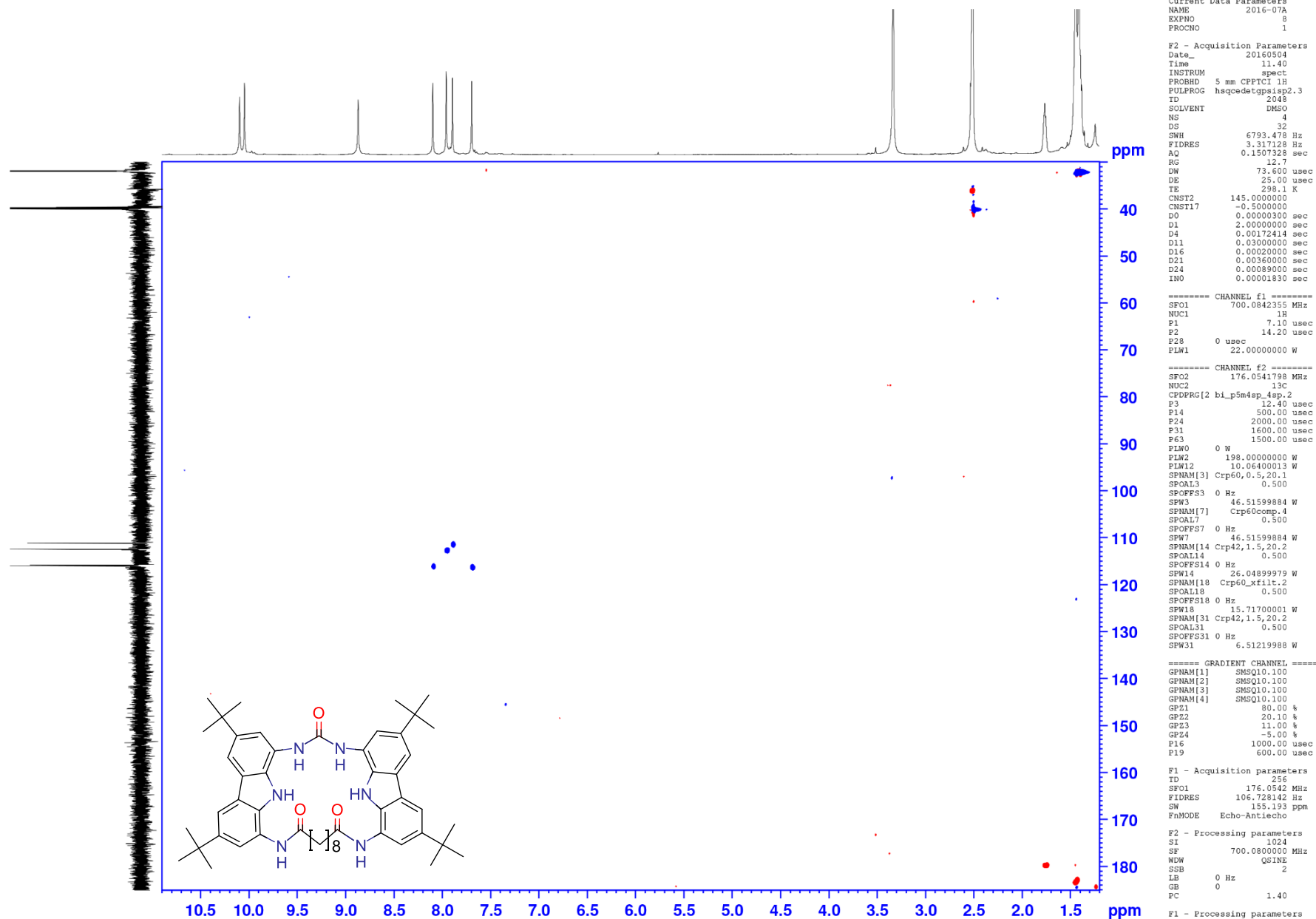
¹H NMR spectrum (700.1 MHz) of compound **MC008**



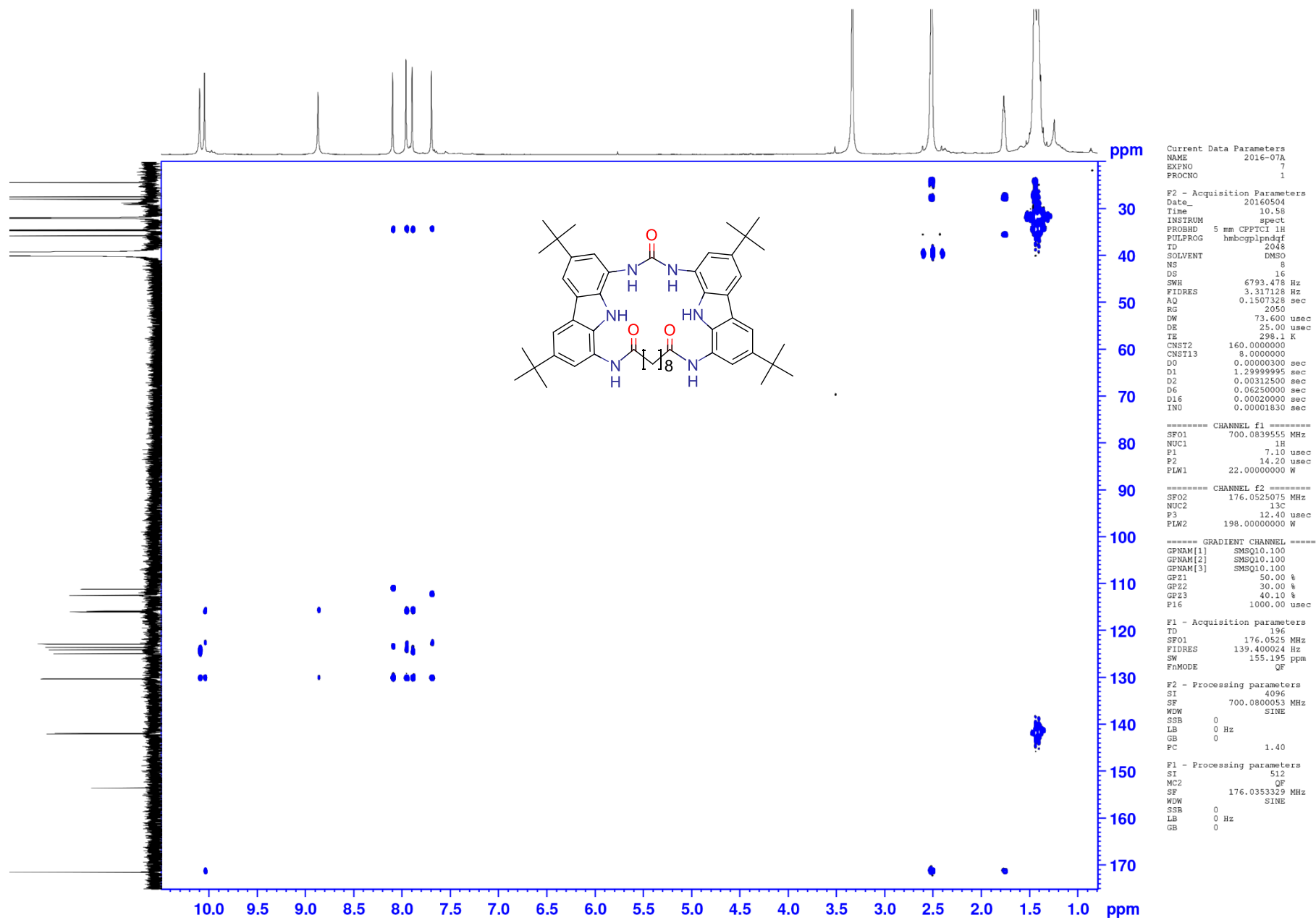
¹³C NMR spectrum (700.1 MHz) of compound **MC008**



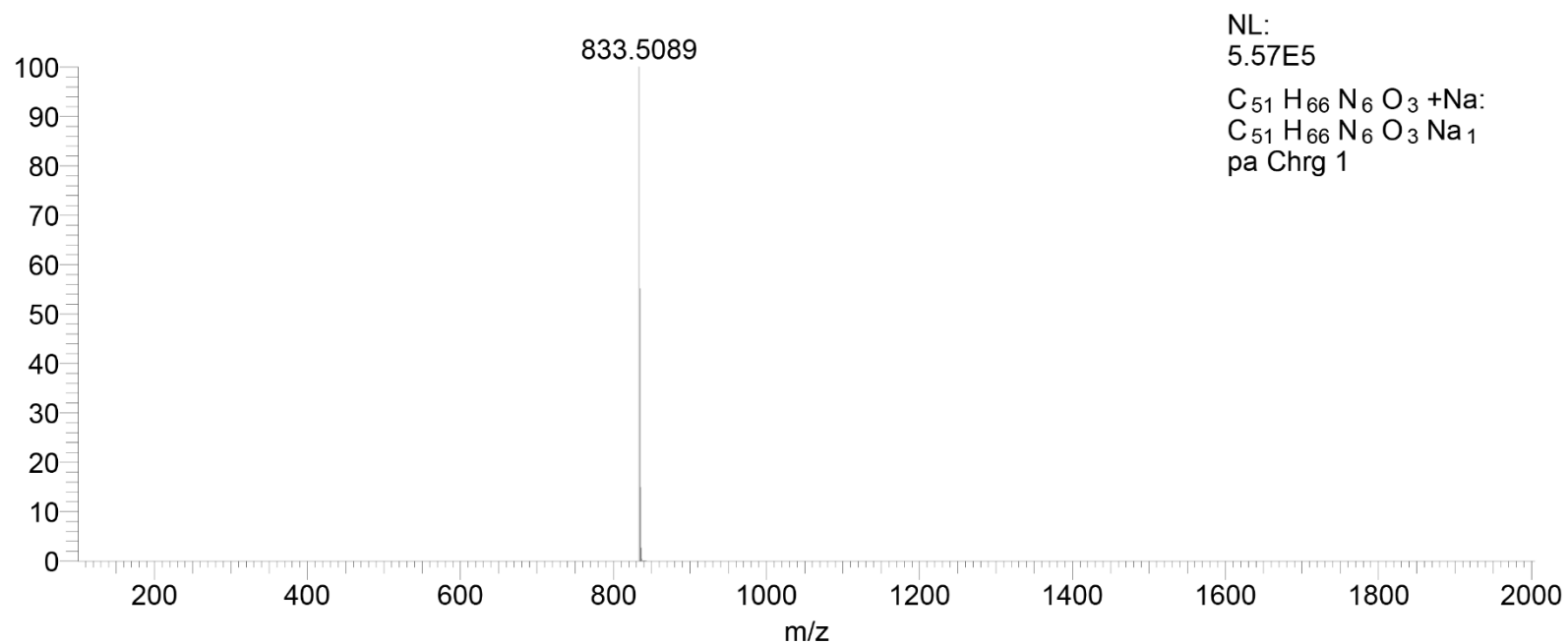
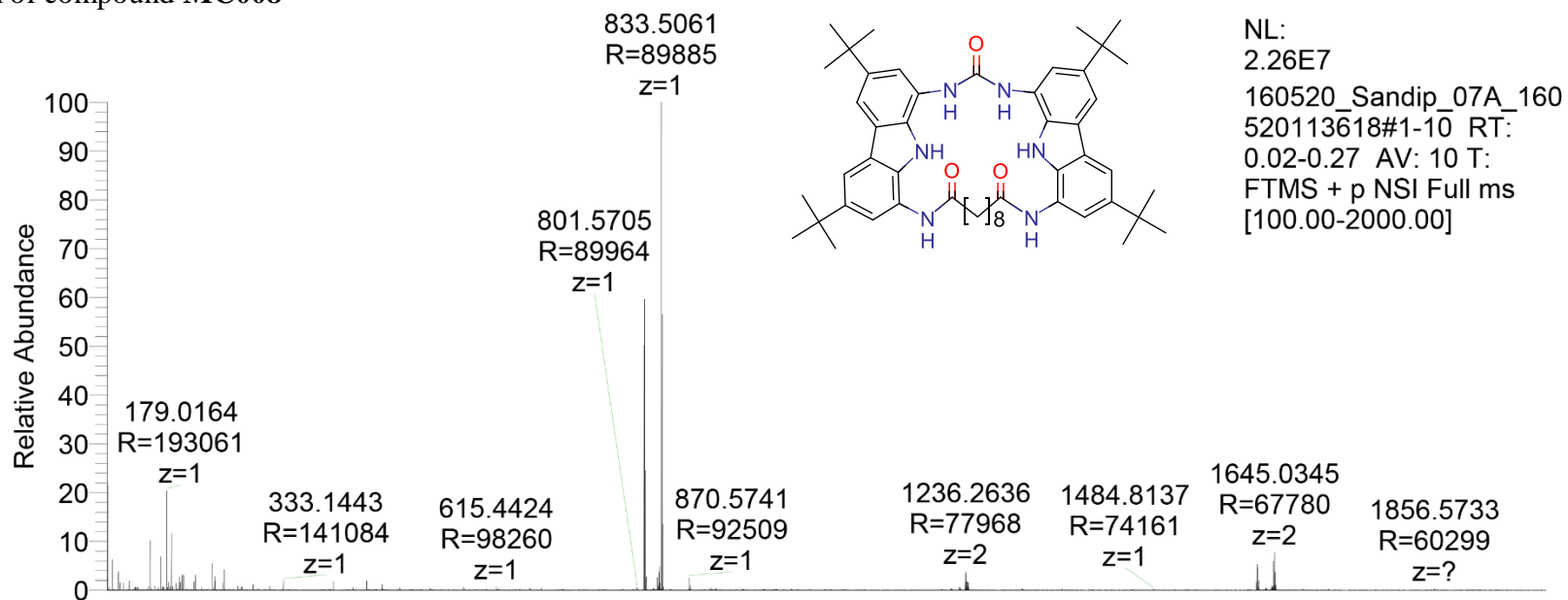
^1H - ^{13}C HSQC spectrum (700.1 MHz) of compound **MC008**



^1H - ^{13}C HMBC spectrum (700.1 MHz) of compound **MC008**



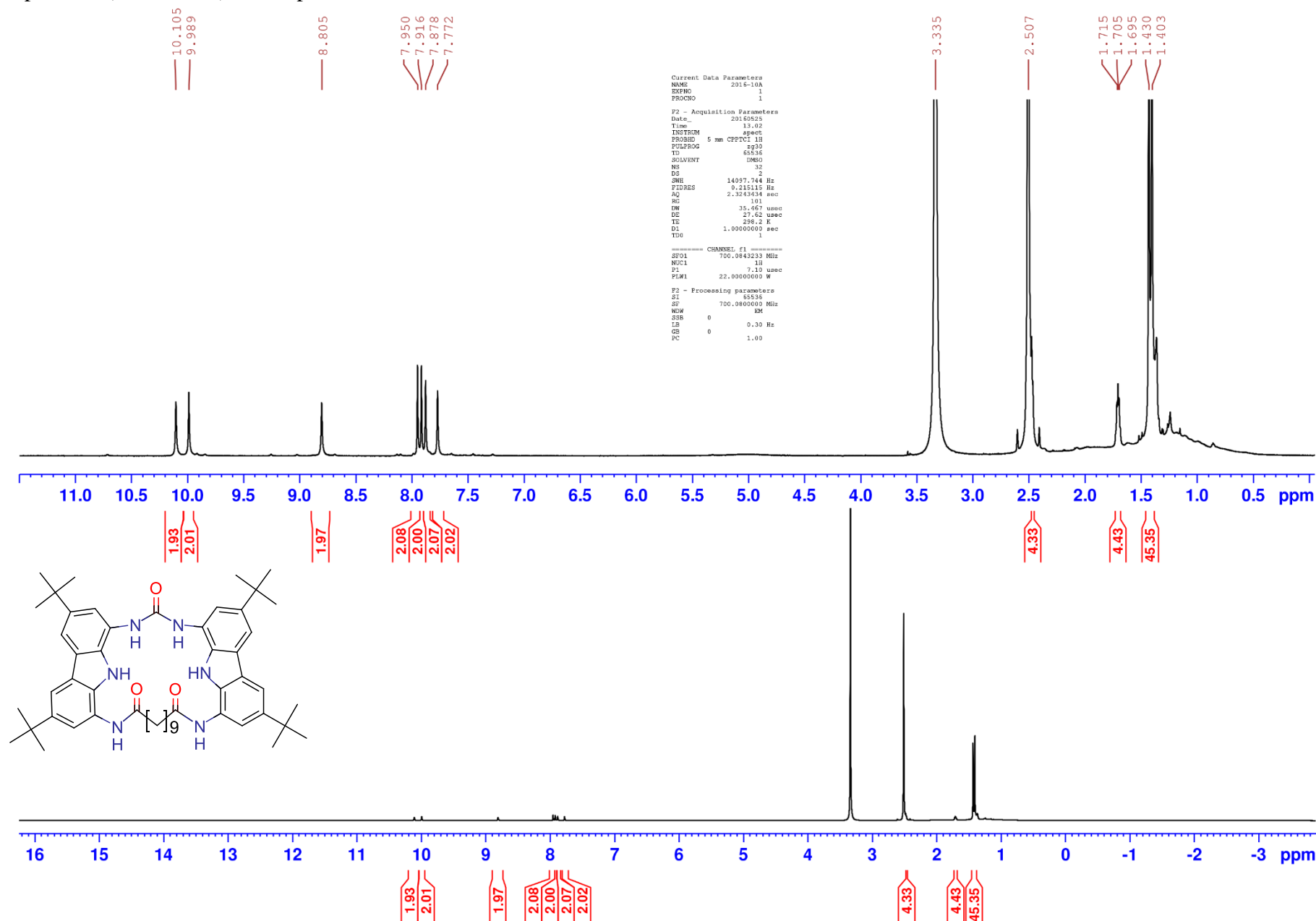
HRMS spectrum of compound **MC008**



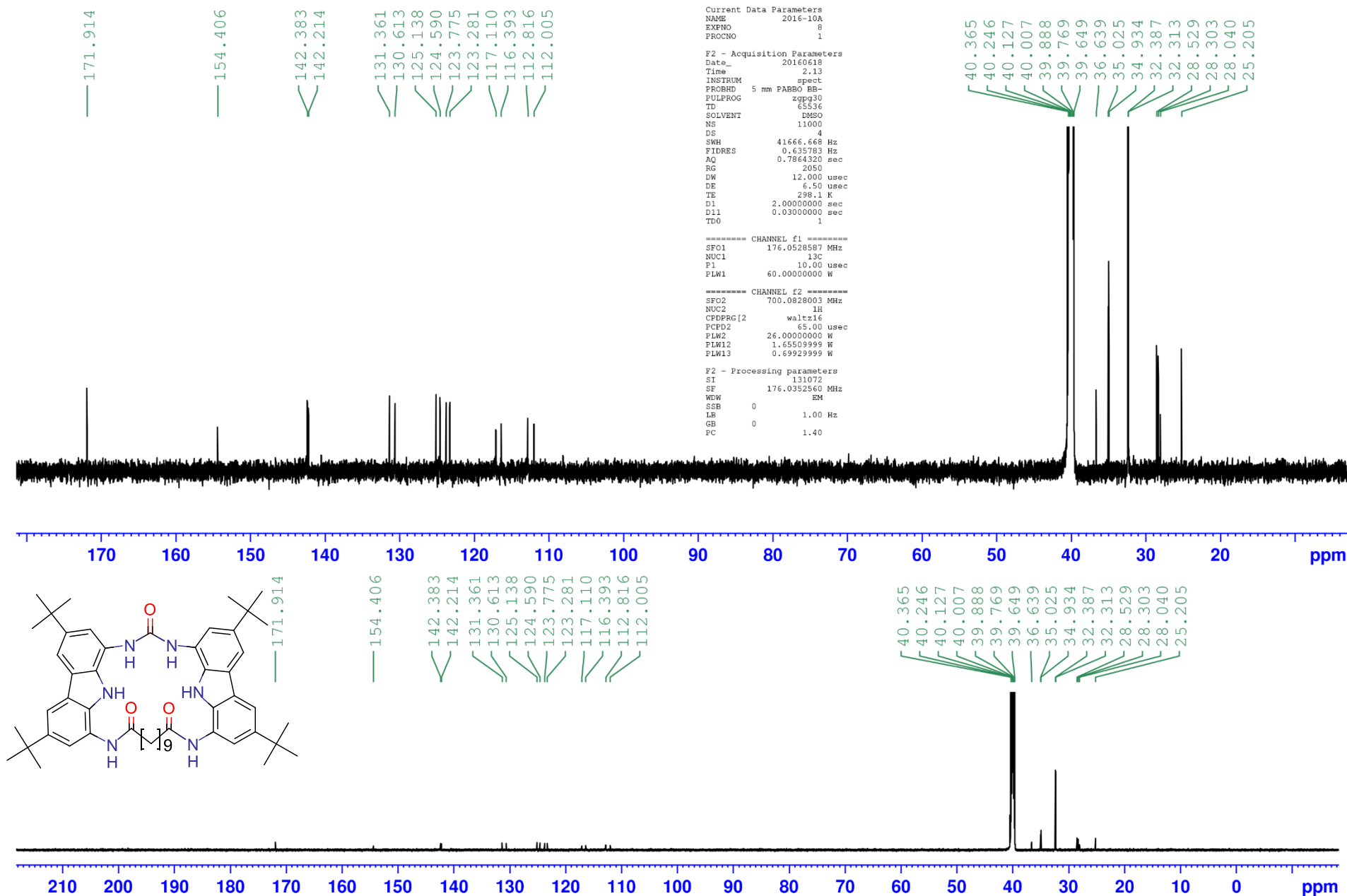
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0.02-0.27 AV: 10 T:
FTMS + p NSI Full ms
[100.00-2000.00]

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C₅₁ H₆₆ N₆ O₃ +Na:
C₅₁ H₆₆ N₆ O₃ Na₁
pa Chrg 1

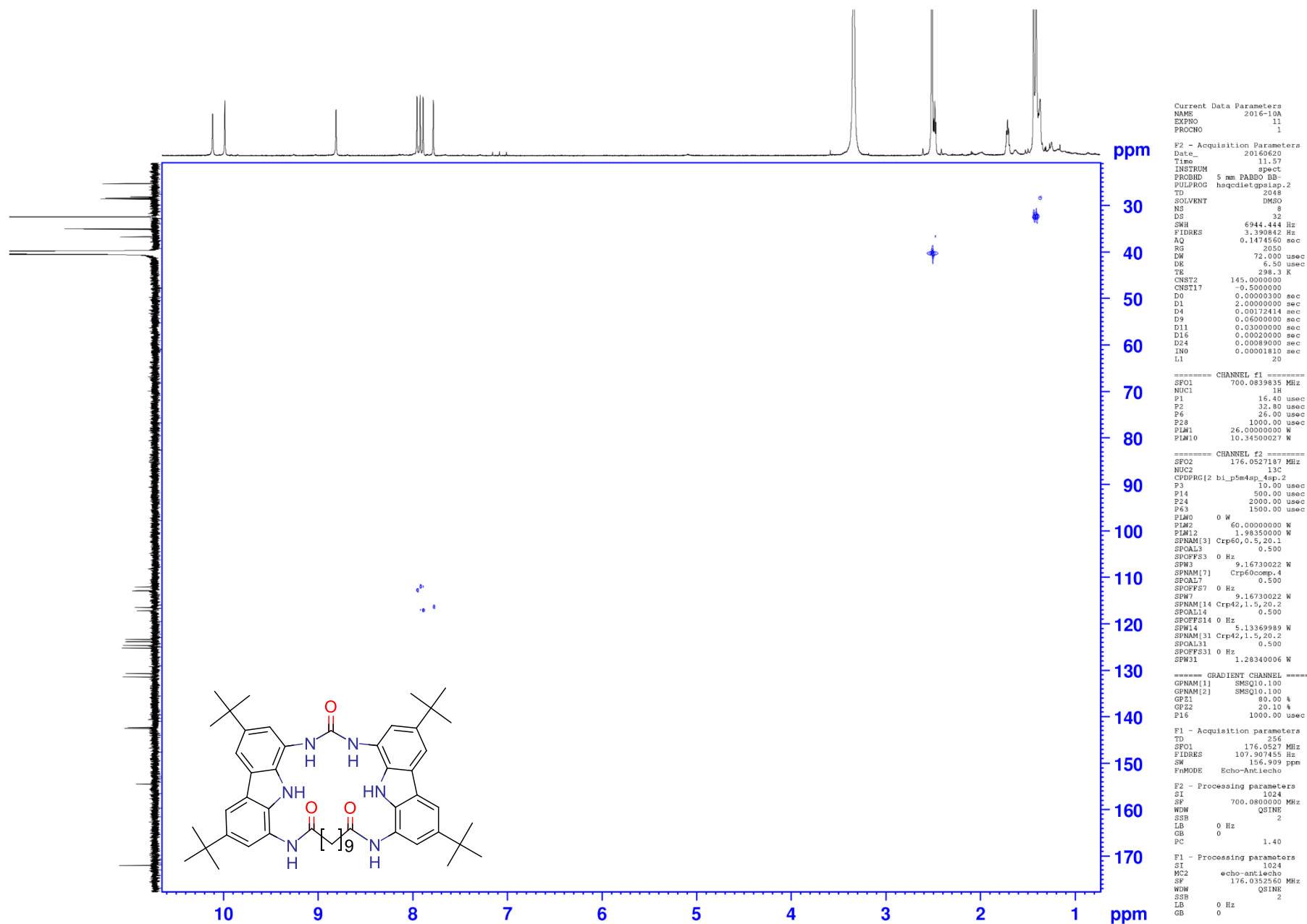
¹H NMR spectrum (700.1 MHz) of compound **MC009**



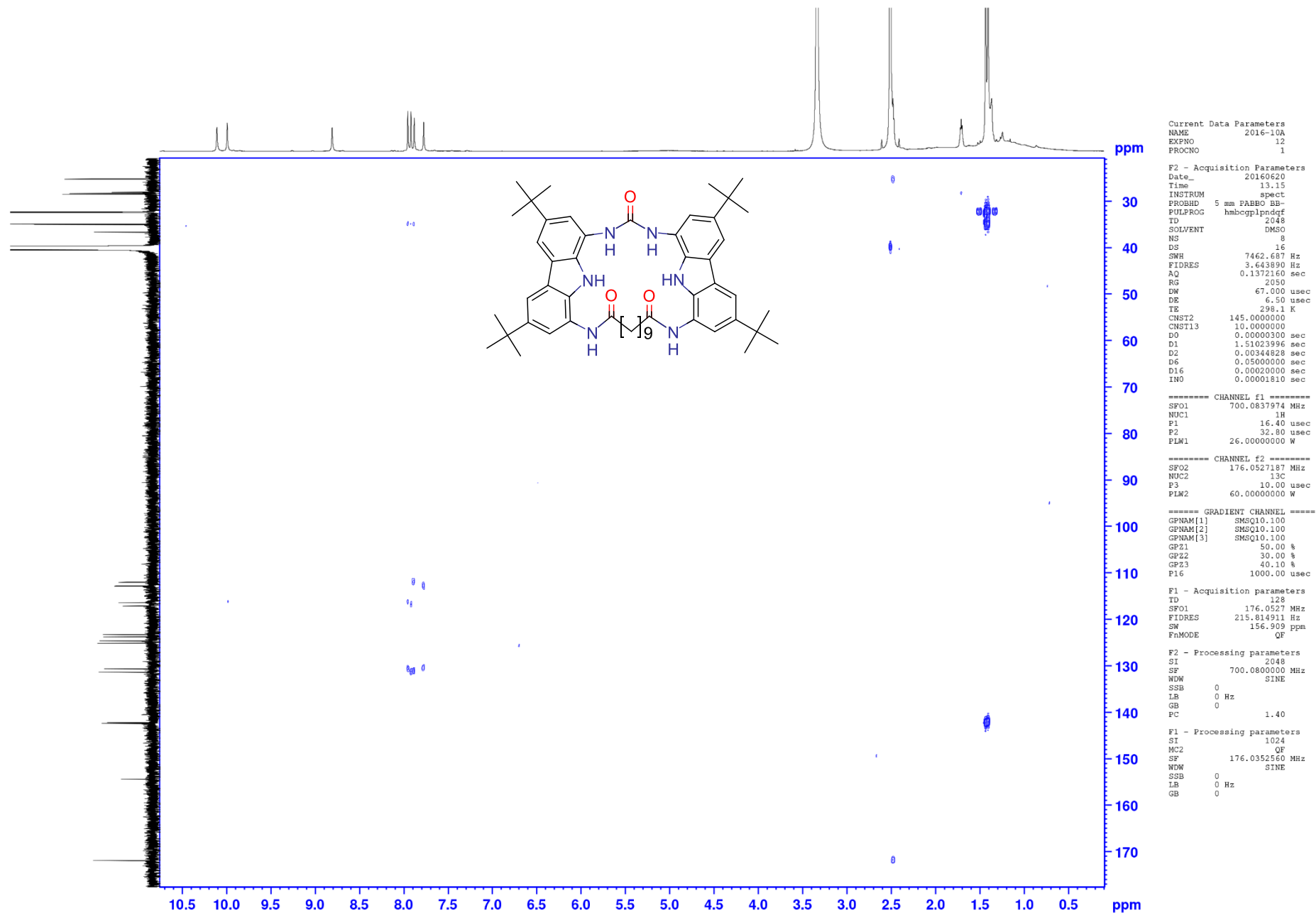
¹³C NMR spectrum (700.1 MHz) of compound **MC009**



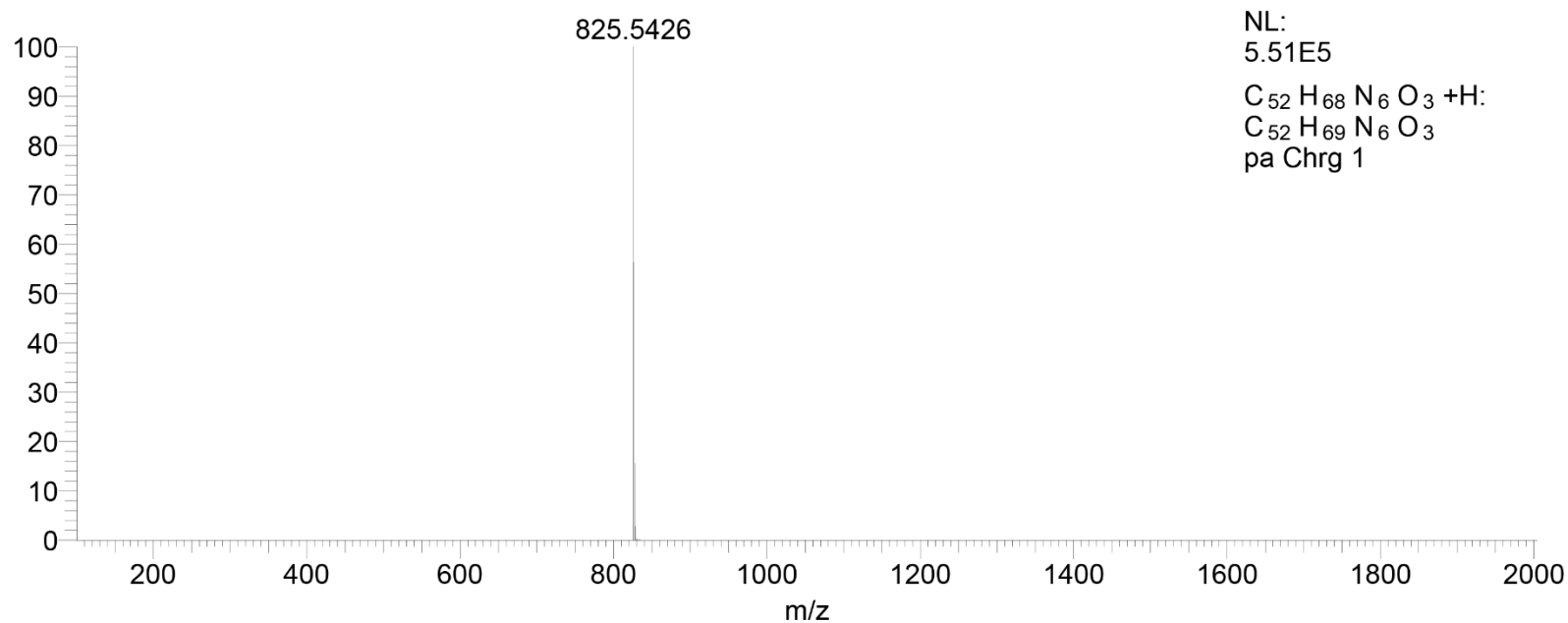
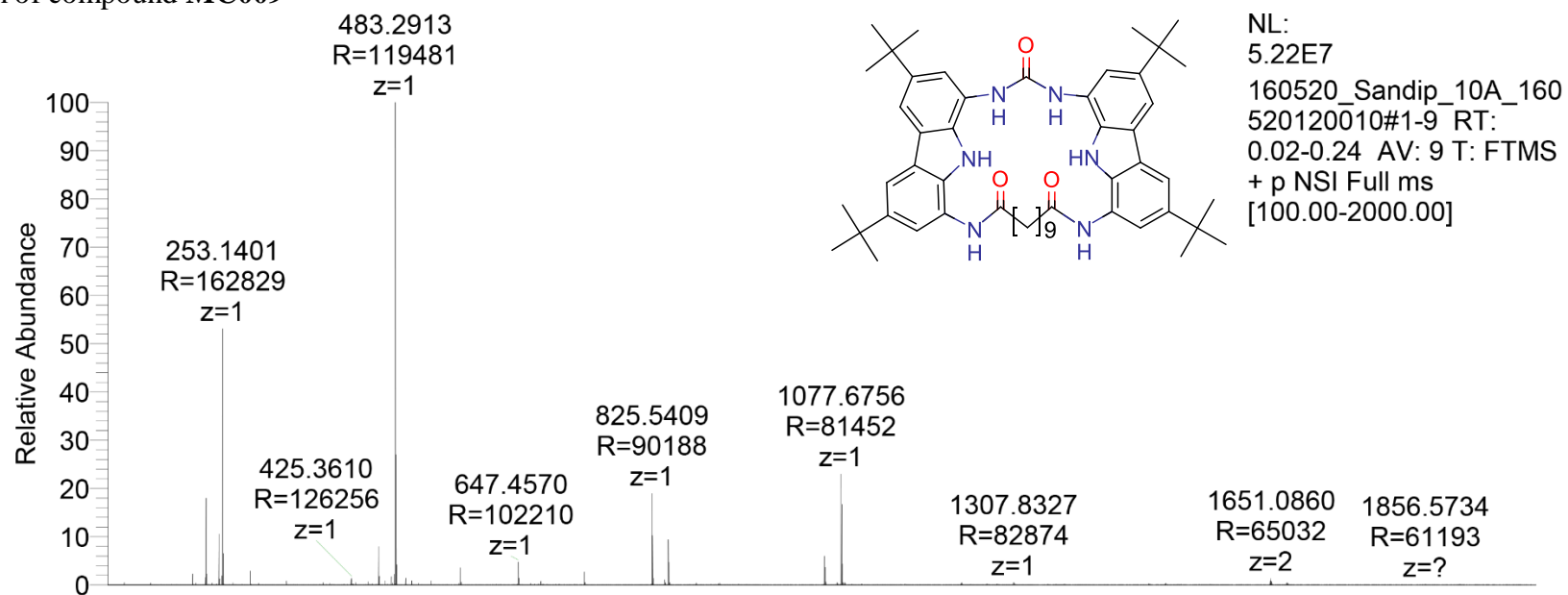
^1H - ^{13}C HSQC spectrum (700.1 MHz) of compound **MC009**



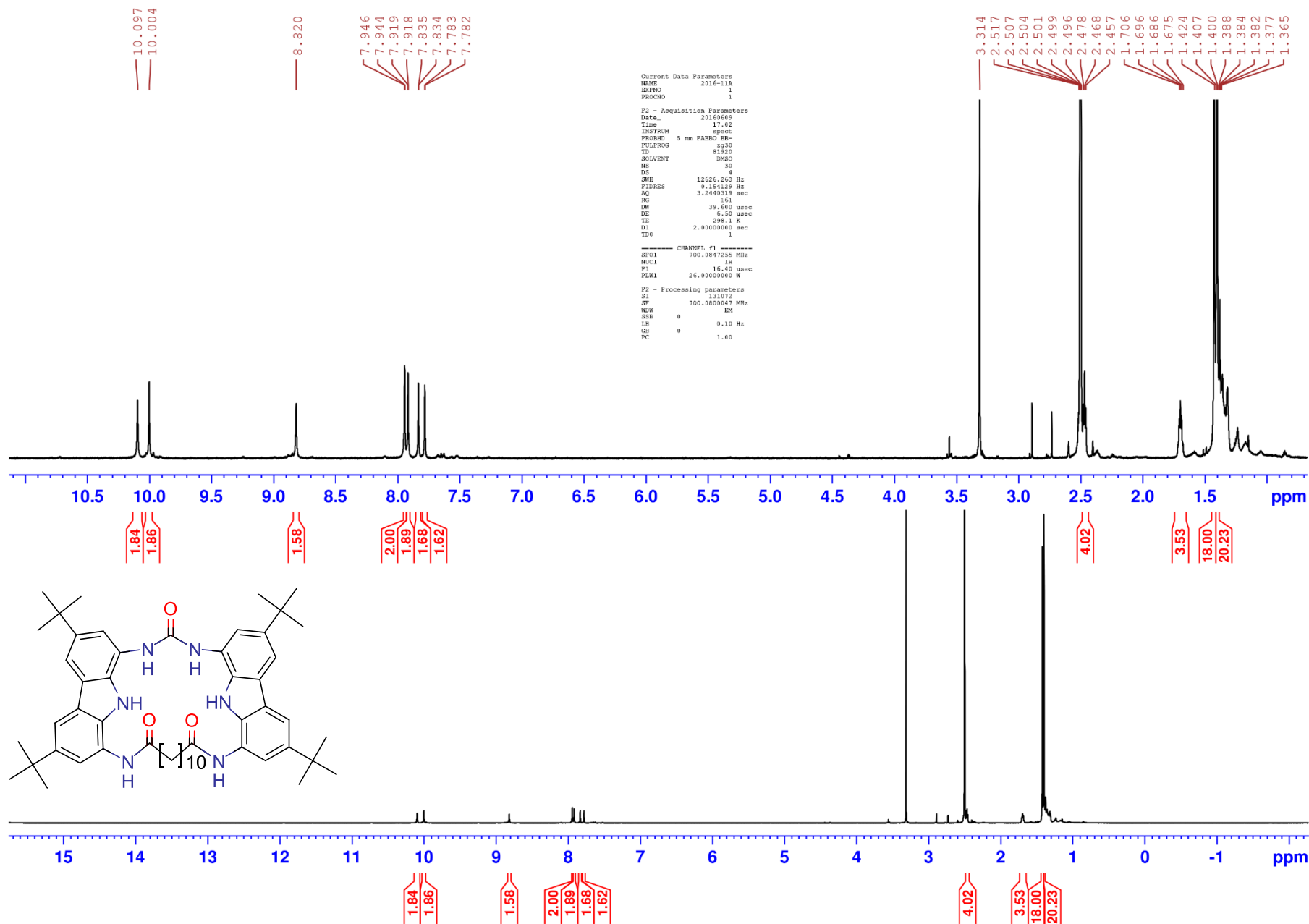
^1H - ^{13}C HMBC spectrum (700.1 MHz) of compound **MC009**



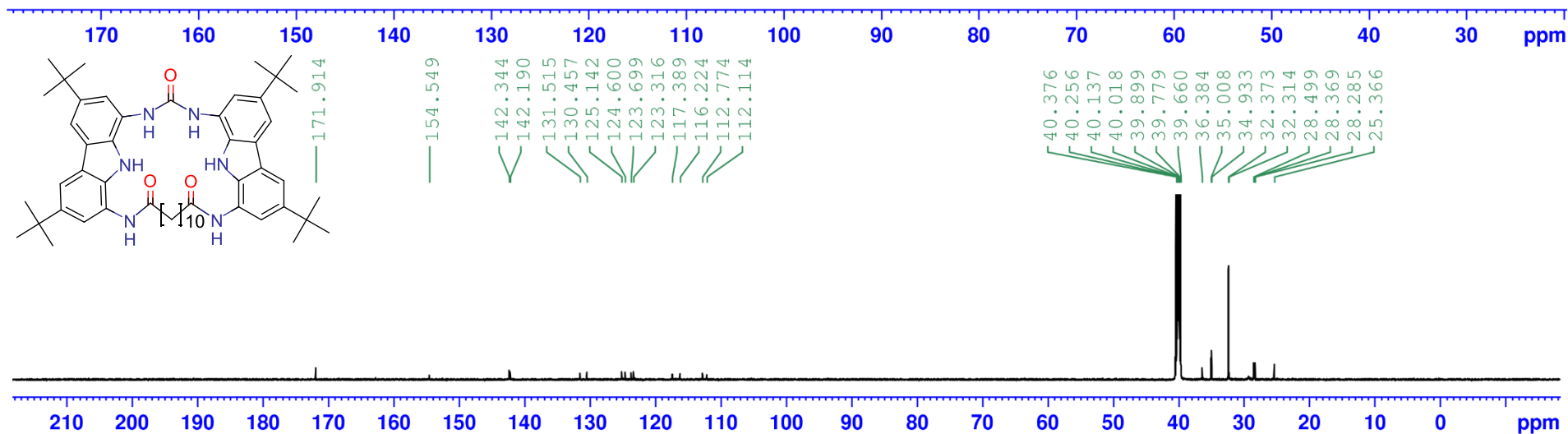
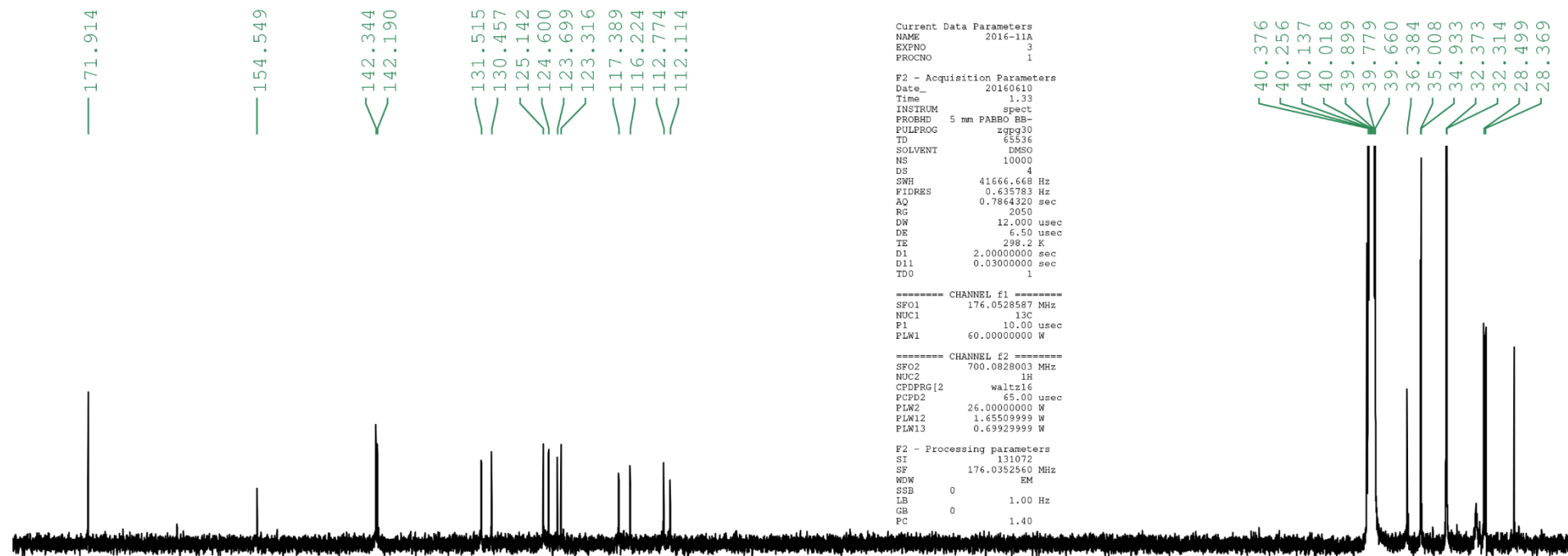
HRMS spectrum of compound **MC009**



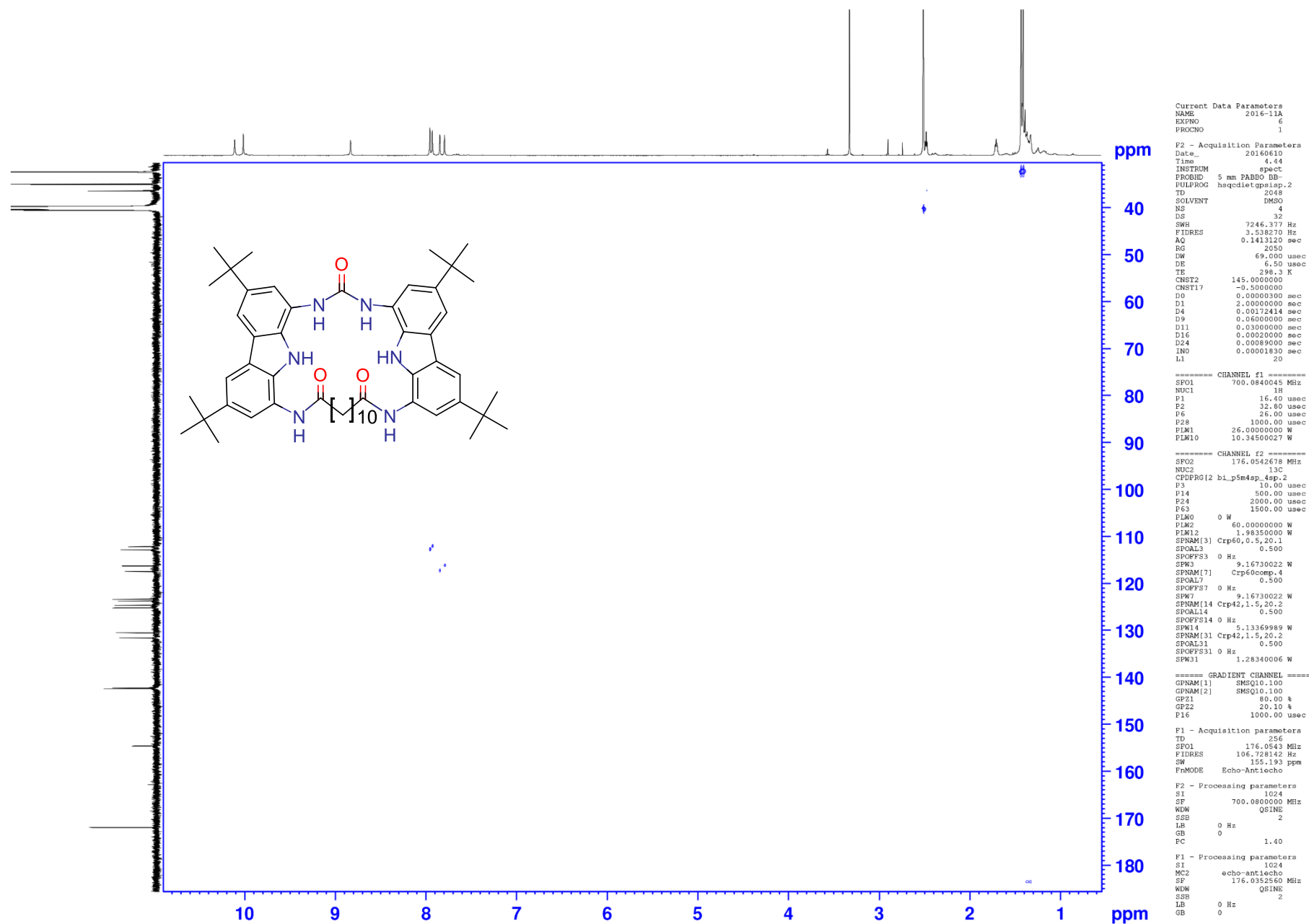
¹H NMR spectrum (700.1 MHz) of compound **MC010**



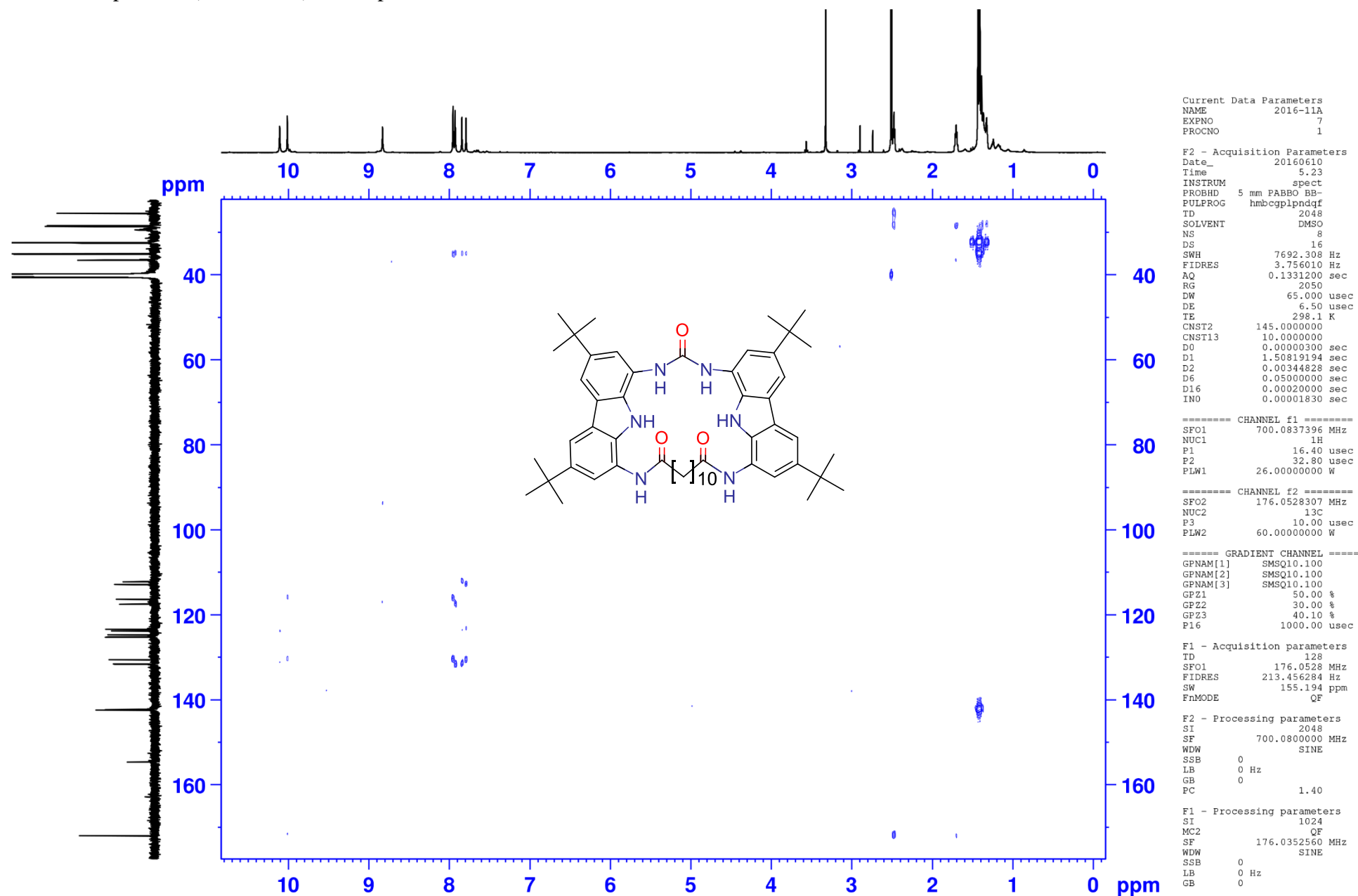
¹³C NMR spectrum (700.1 MHz) of compound **MC010**



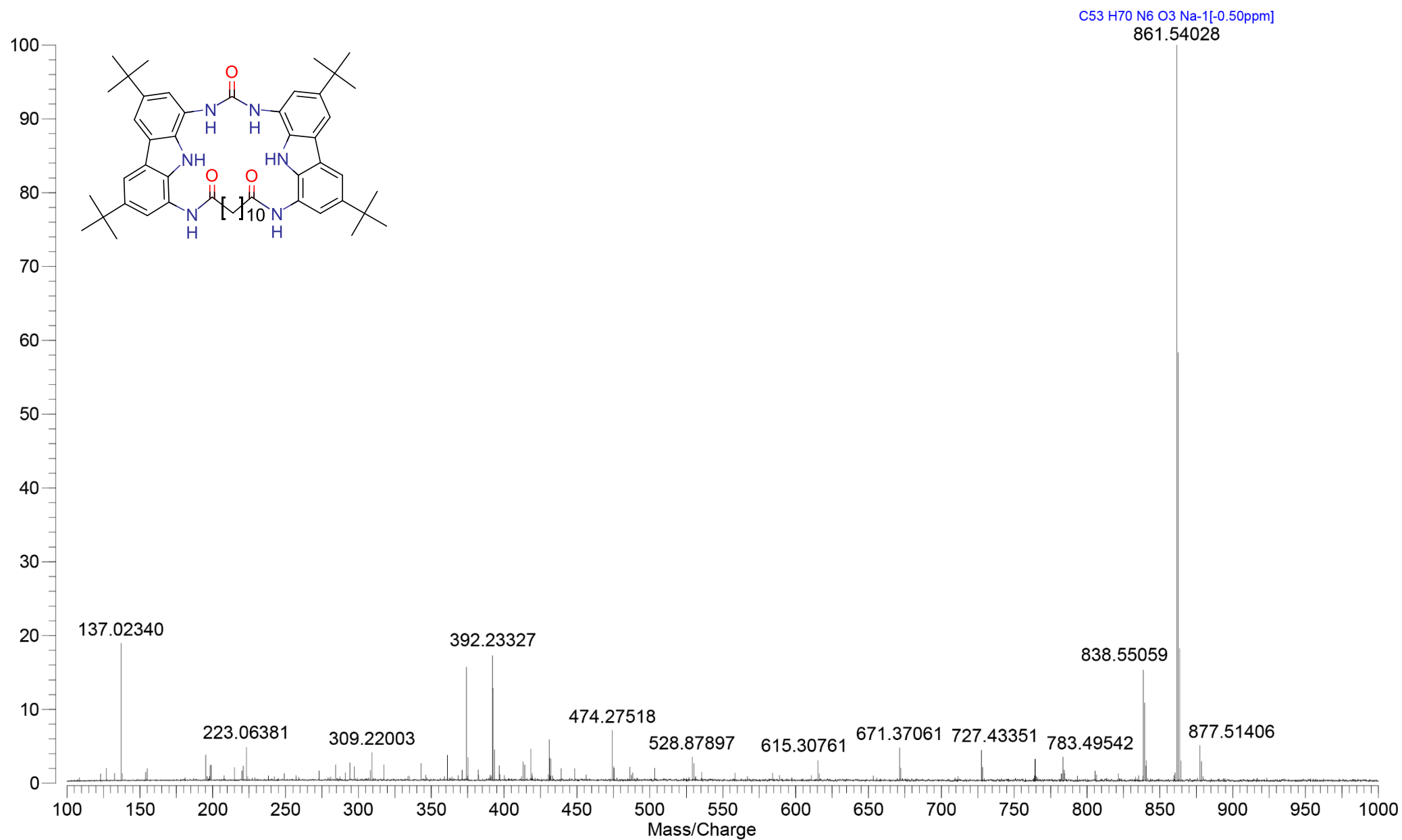
^1H - ^{13}C HSQC spectrum (700.1 MHz) of compound **MC010**



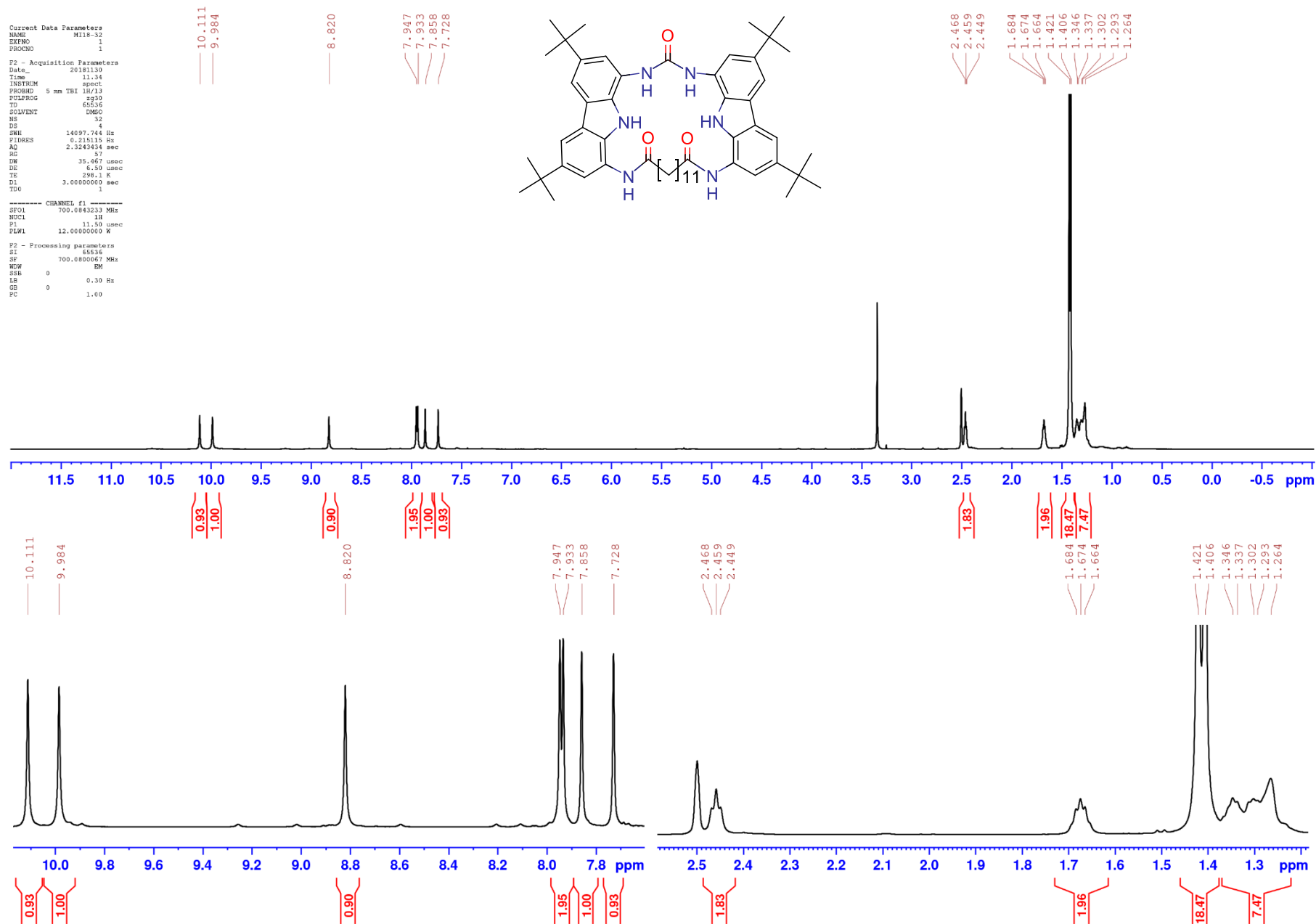
^1H - ^{13}C HMBC spectrum (700.1 MHz) of compound **MC010**



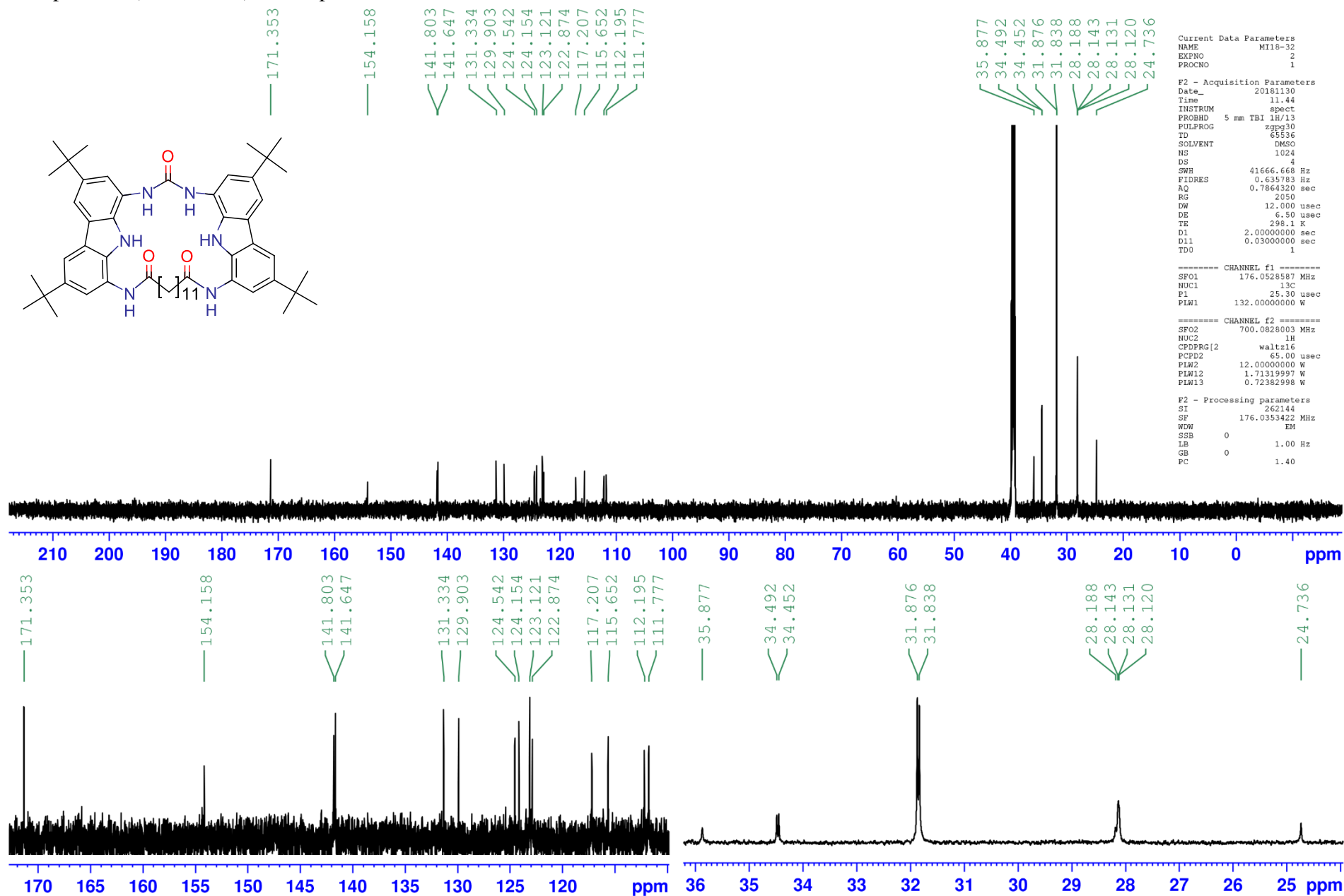
HRMS spectrum of compound **MC010**



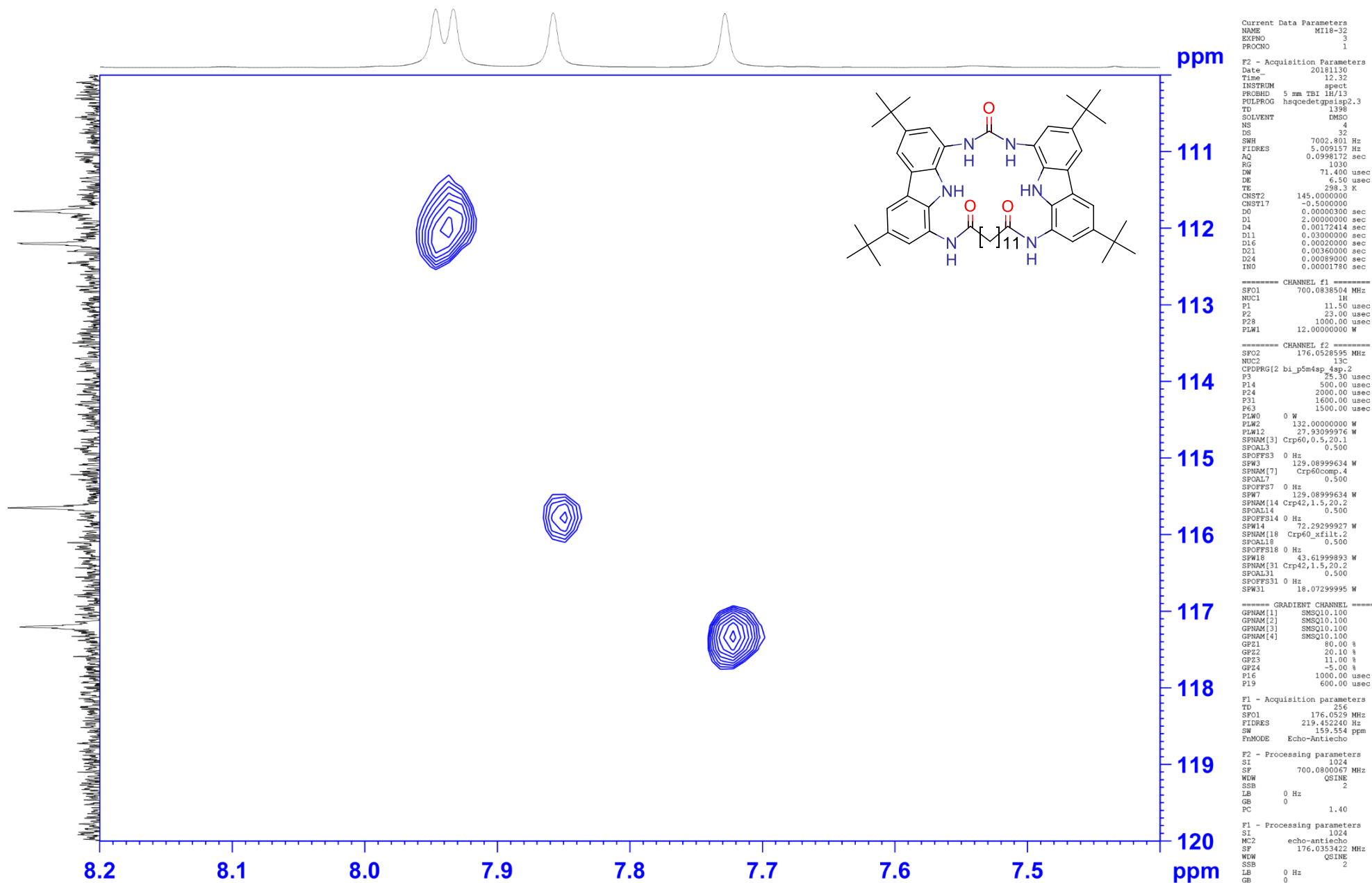
¹H NMR spectrum (700.1 MHz) of compound MC011



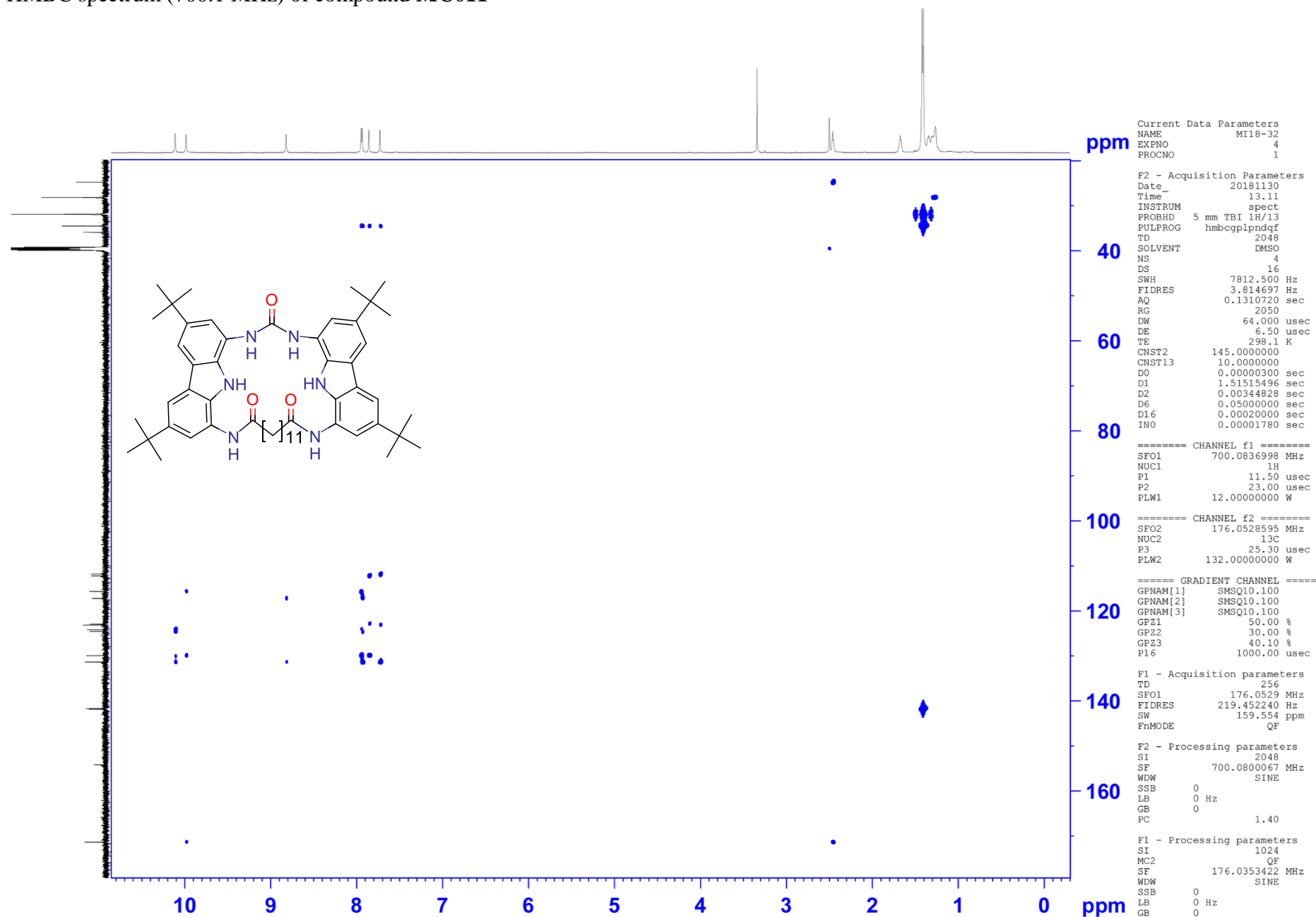
¹³C NMR spectrum (700.1 MHz) of compound **MC011**



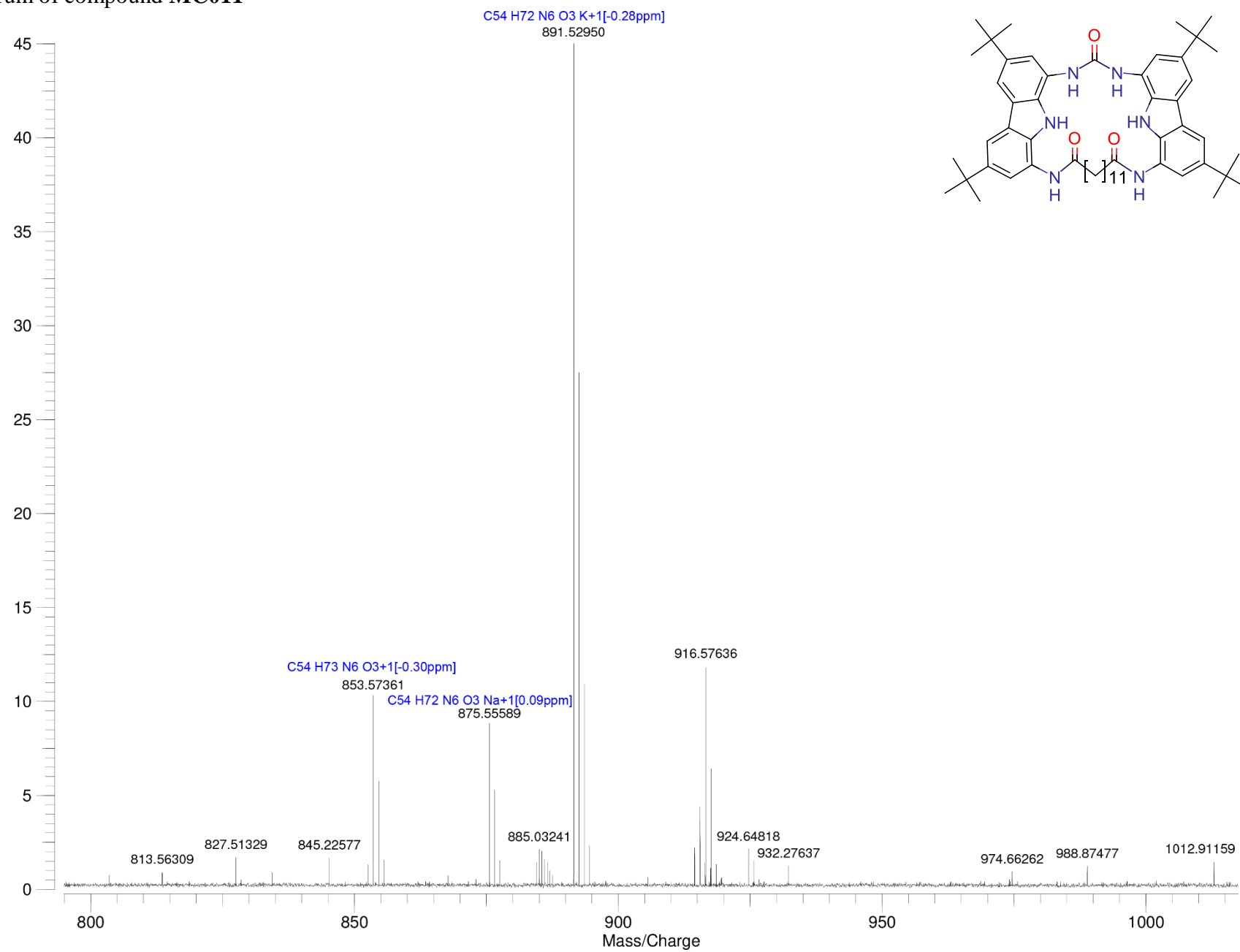
^1H - ^{13}C HSQC spectrum (700.1 MHz) of compound **MC011**



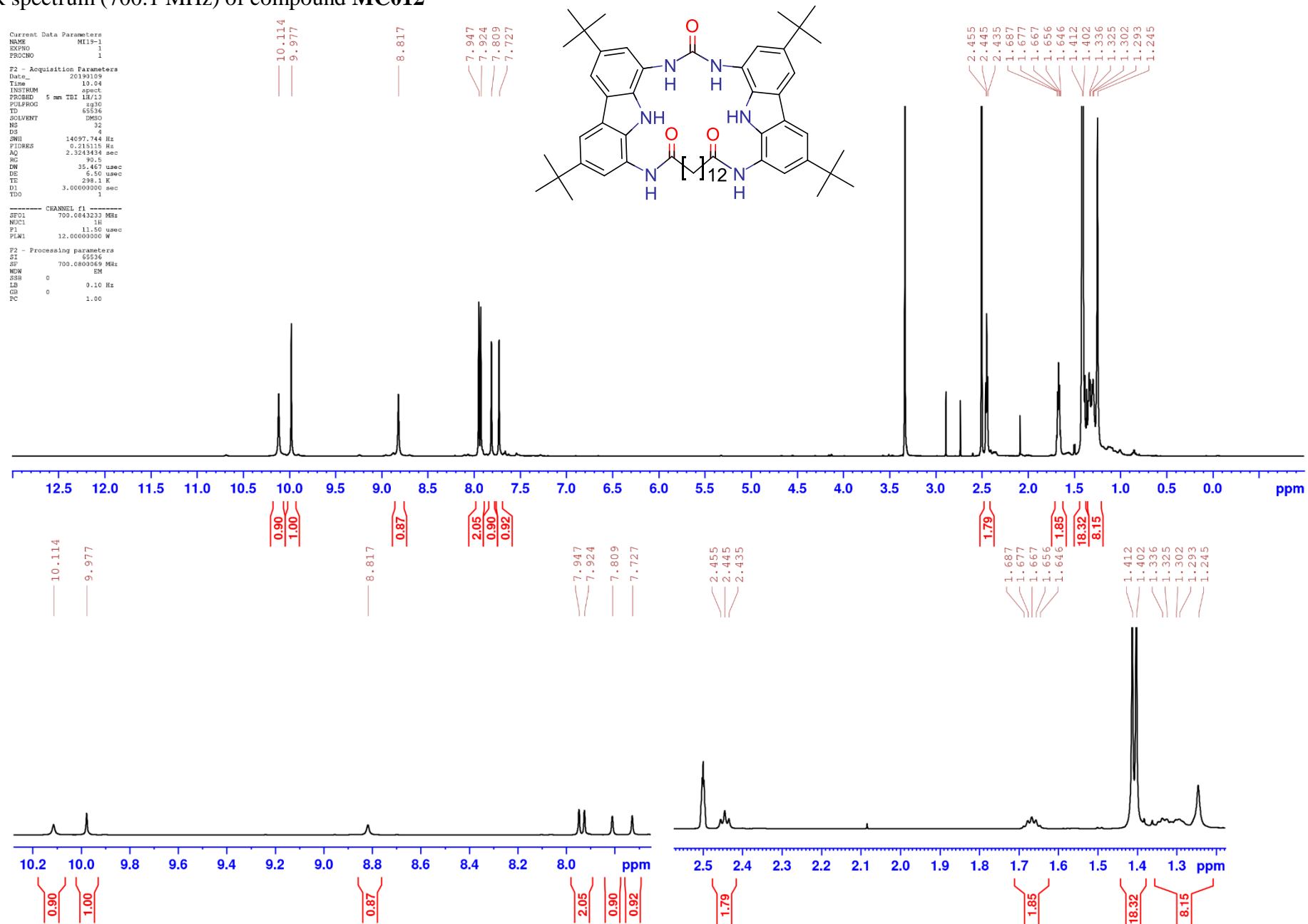
^1H - ^{13}C HMBC spectrum (700.1 MHz) of compound **MC011**



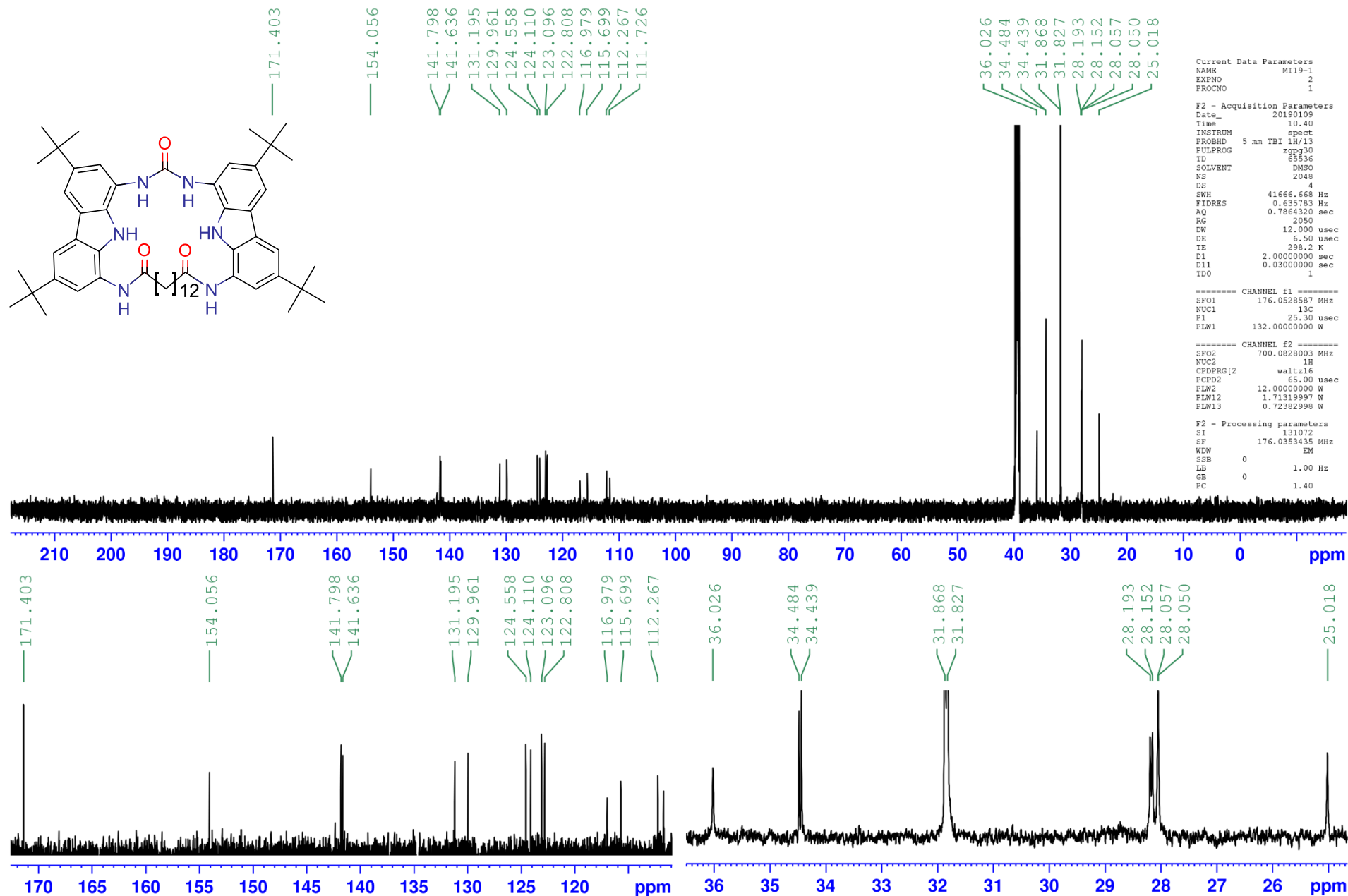
HRMS spectrum of compound MC011



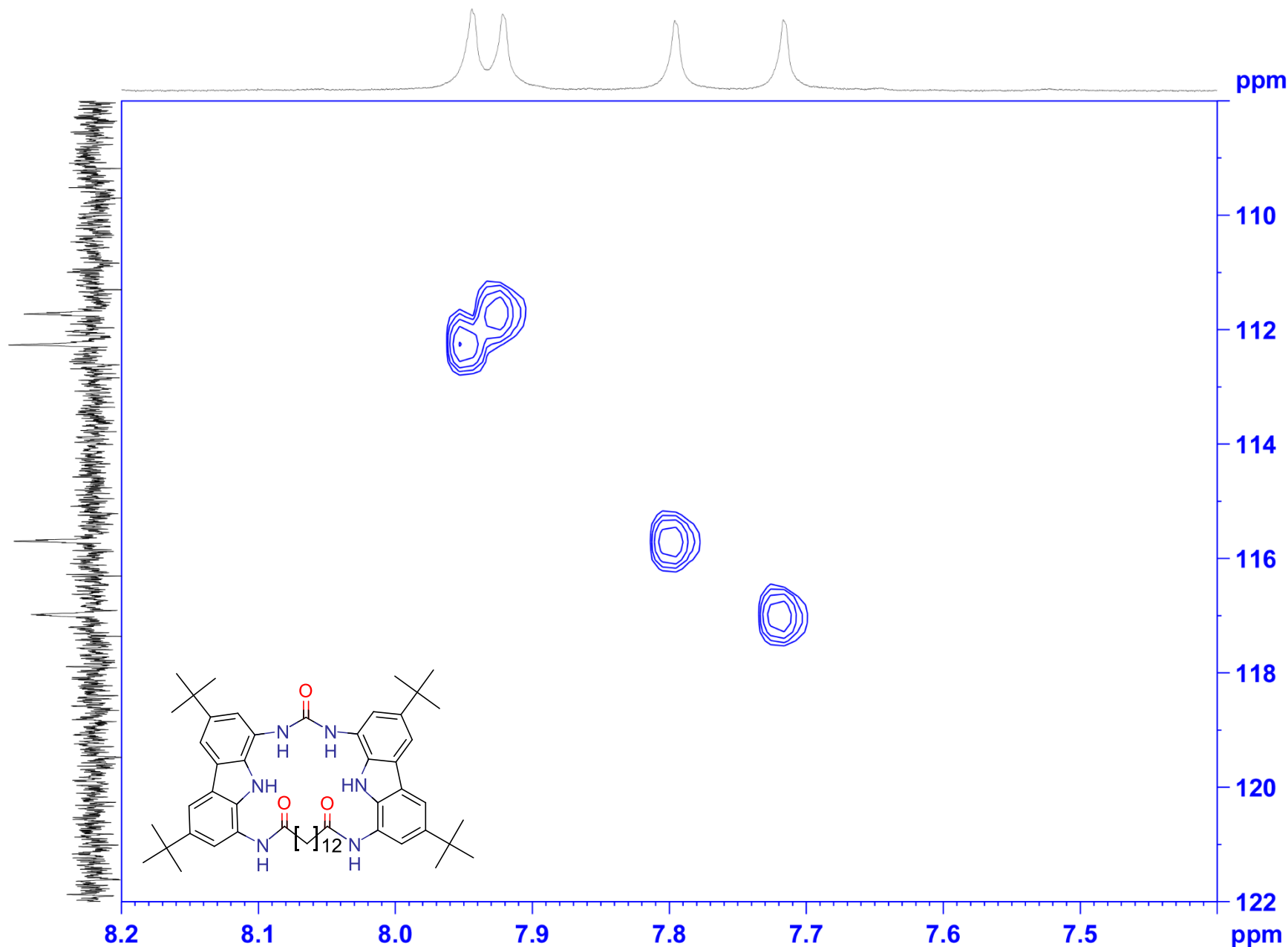
¹H NMR spectrum (700.1 MHz) of compound **MC012**



^{13}C NMR spectrum (700.1 MHz) of compound **MC012**



^1H - ^{13}C HSQC spectrum (700.1 MHz) of compound **MC012**



```

Current Data Parameters
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EXPNO         5
PROCNO        1

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Time          16.55
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SOLVENT       DMSO
NS            8
DS            32
SWH           7002.801 Hz
FIDRES       3.419337 Hz
AQ           0.1462272 sec
RG           2050
DW           71.400 usec
DE           6.50 usec
TE           299.2 K
CNST2        145.0000000
CNST17       -0.5000000
D0           0.00000300 sec
D1           2.00000000 sec
D4           0.00172414 sec
D11          0.03000000 sec
D16          0.00020000 sec
D21          0.00360000 sec
D24          0.00089000 sec
INO          0.00001780 sec

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SFO1         700.0838504 MHz
NUC1          1H
F1           15.06 usec
P2           30.12 usec
P28          1000.00 usec
PLW1         26.00000000 W

===== CHANNEL f2 =====
SFO2         176.0528595 MHz
NUC2          13C
CPDPRG2       bi_p5m4sp_4sp.2
P3           9.57 usec
P14          500.00 usec
P24          2000.00 usec
P31          1600.00 usec
P63          1500.00 usec
PLW0          0 W
PLW2         60.00000000 W
ELW12        1.81659997 W
SPNAM[3]     Crp60,0.5,20.1
SFOAL3        0 Hz
SPOFFS3       8.39589977 W
SPW3         8.39589977 W
SPNAM[7]     Crp60comp.4
SFOAL7        0 Hz
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SPW7         8.39589977 W
SPNAM[14]    Crp42,1.5,20.2
SFOAL14       0 Hz
SPOFFS14      4.70170021 W
SPW14        4.70170021 W
SPNAM[18]    Crp60_xfilt.2
SFOAL18       0 Hz
SPOFFS18      0.500
SPW18        2.83690000 W
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SFOAL31       0 Hz
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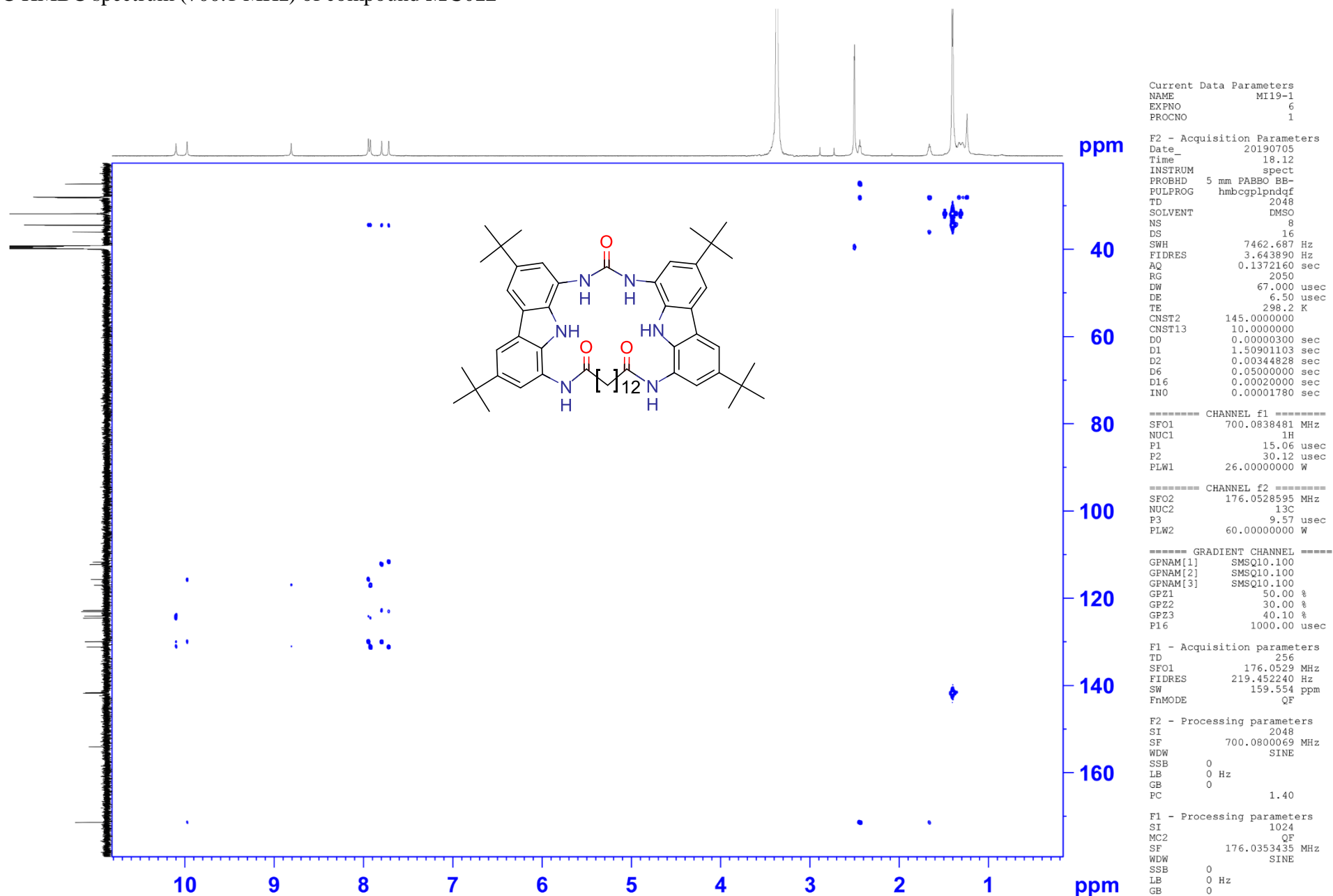
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GPZ3         11.00 %
GPZ4         -5.00 %
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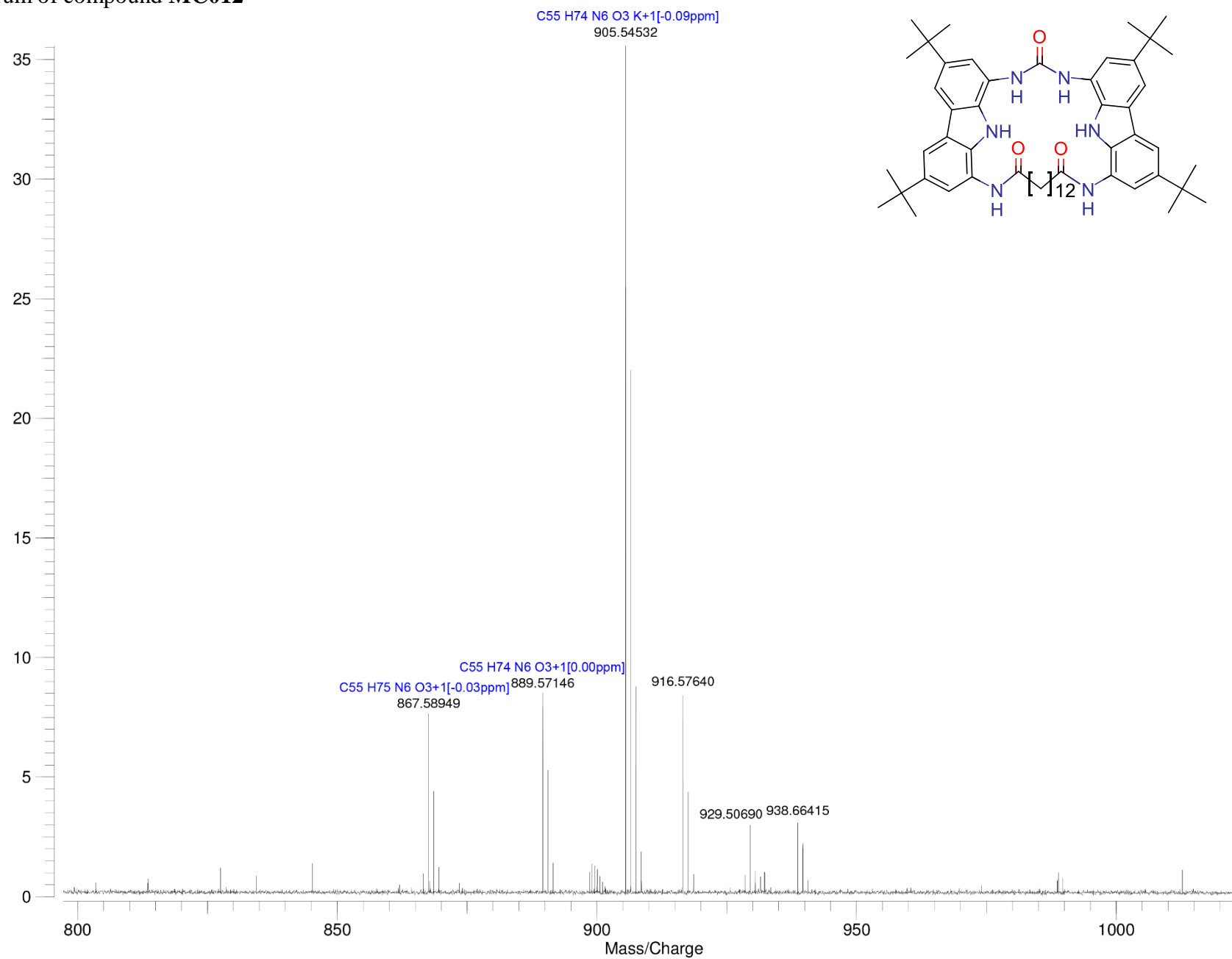
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SSB          2
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GB           0
PC           1.40

F1 - Processing parameters
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WC2          echo-antiecho
SF           176.0353435 MHz
WDW          QSINE
SSB          2
LB           0 Hz
GB           0
  
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^1H - ^{13}C HMBC spectrum (700.1 MHz) of compound **MC012**

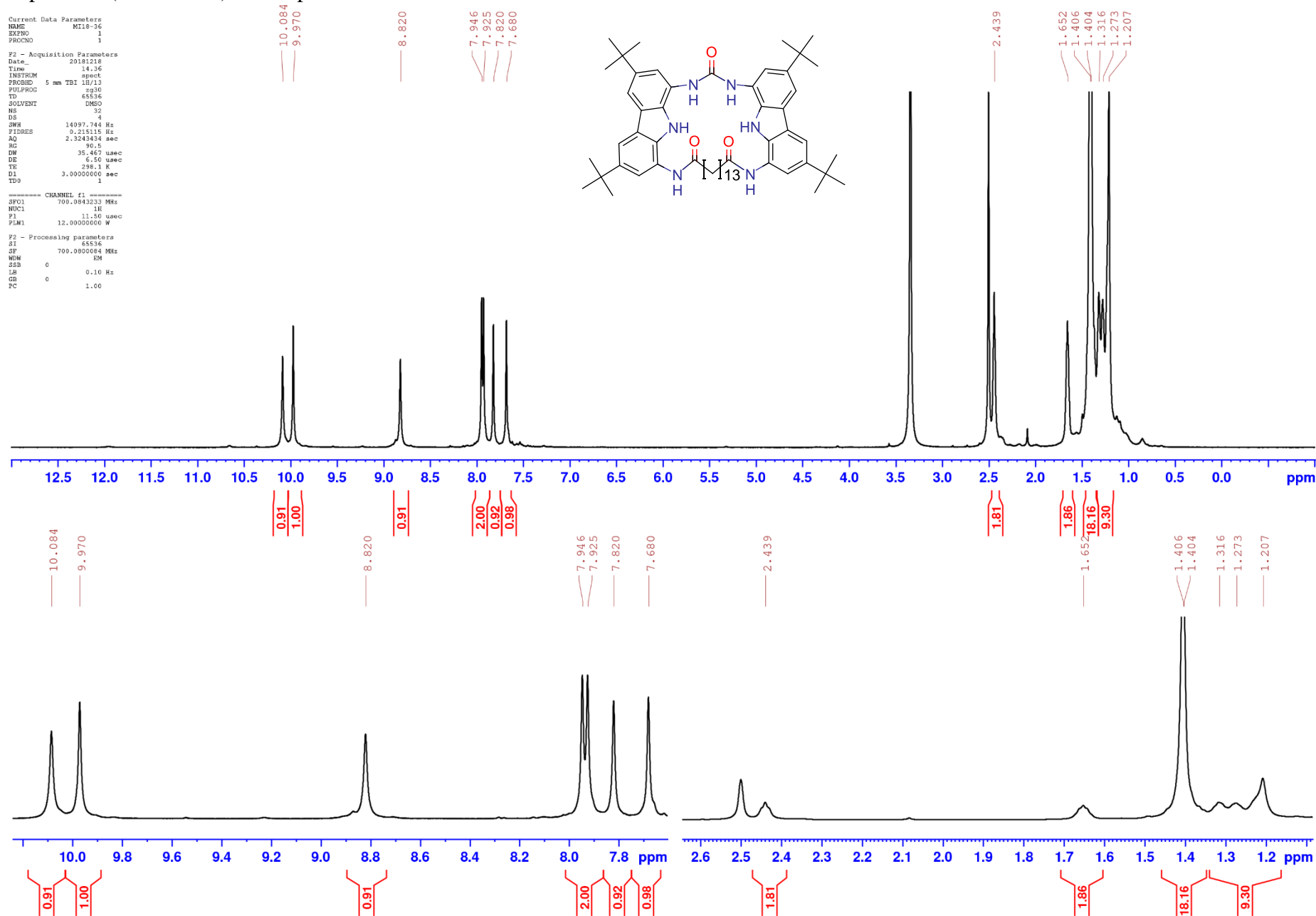
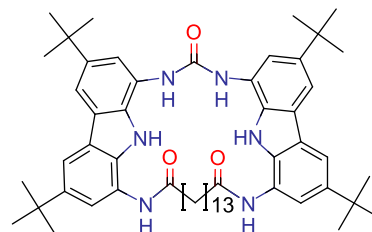


HRMS spectrum of compound MC012

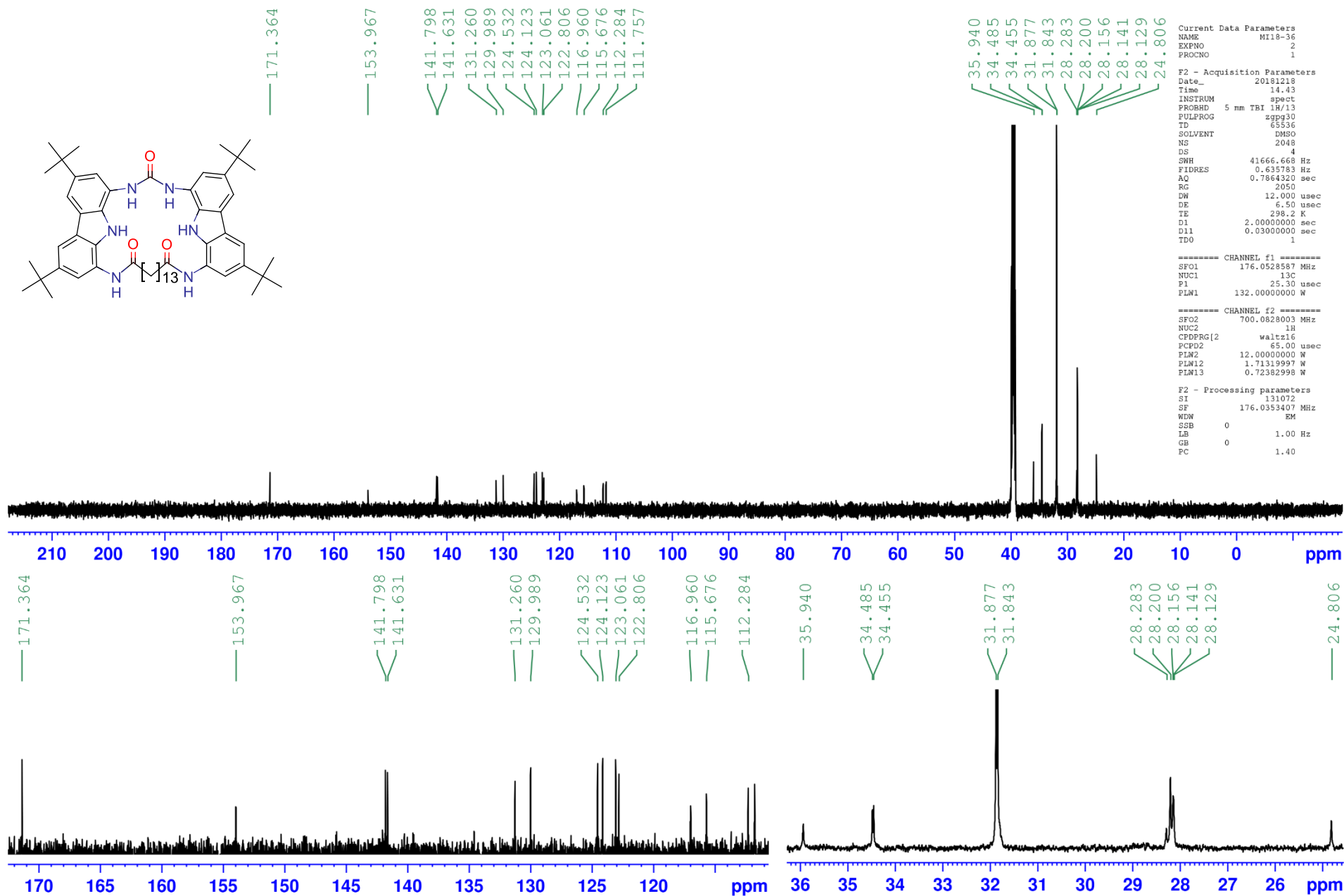


¹H NMR spectrum (700.1 MHz) of compound MC013

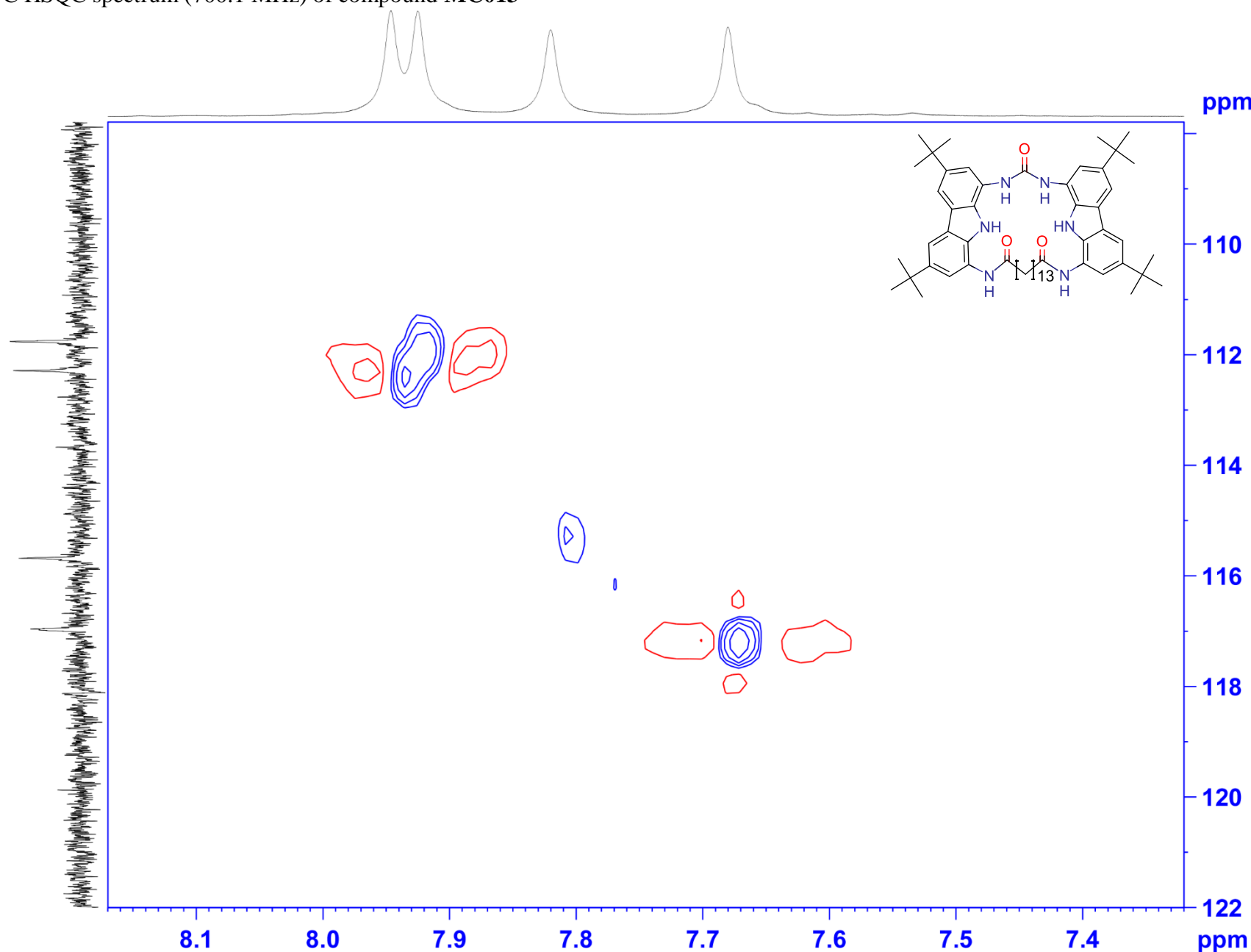
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PROCNO 1
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DS 4
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FIDRES 0.215115 Hz
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RG 90.5
DW 35.467 usec
DE 6.50 usec
TE 298.1 K
D1 3.0000000 sec
TD0 1
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NUC1 1H
P1 11.50 usec
PL1 12.00000000 W
F2 - Processing parameters
SI 65536
SF 700.0800084 MHz
WDW DM
SSB 0
LB 0.10 Hz
GB 0
PC 1.00



^{13}C NMR spectrum (700.1 MHz) of compound **MC013**



^1H - ^{13}C HSQC spectrum (700.1 MHz) of compound **MC013**



Current Data Parameters
NAME MI18-36
EXPNO 3
PROCNO 1

F2 - Acquisition Parameters
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Time 16.22
INSTRUM spect
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PULPROG haqcddetgpgasp2.3
TD 1398
SOLVENT DMSO
NS 4
DS 32
SWH 7002.801 Hz
FIDRES 5.009157 Hz
AQ 0.0998172 sec
RG 2050
DW 71.400 usec
DE 6.50 usec
TE 298.4 K
CNST2 145.0000000
CNST17 -0.5000000
DO 0.00000300 sec
D1 2.00000000 sec
D4 0.00172414 sec
D11 0.03000000 sec
D16 0.00020000 sec
D21 0.00360000 sec
D24 0.00089000 sec
IN0 0.00001780 sec

===== CHANNEL f1 =====
SFO1 700.0838504 MHz
NUC1 1H
P1 11.50 usec
P2 23.00 usec
P28 1000.00 usec
PLW1 12.00000000 W

===== CHANNEL f2 =====
SFO2 176.0525075 MHz
NUC2 13C
CPDPRG2 bi_p5m4sp_4sp.2
P3 25.30 usec
P14 500.00 usec
P24 2000.00 usec
P31 1600.00 usec
P63 1500.00 usec
PLW0 0 W
PLW2 132.00000000 W
PLW12 27.93099976 W
SFOAM[3] Crp60,0.5,20.1
SFOAL3 0.500
SPOFFS3 0 Hz
SPW3 129.08999634 W
SFOAM[7] Crp60comp.4
SFOAL7 0.500
SPOFFS7 0 Hz
SPW7 129.08999634 W
SFOAM[14] Crp42,1.5,20.2
SFOAL14 0.500
SPOFFS14 0 Hz
SPW14 72.29299927 W
SFOAM[18] Crp60_xfilt.2
SFOAL18 0.500
SPOFFS18 0 Hz
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SFOAM[31] Crp42,1.5,20.2
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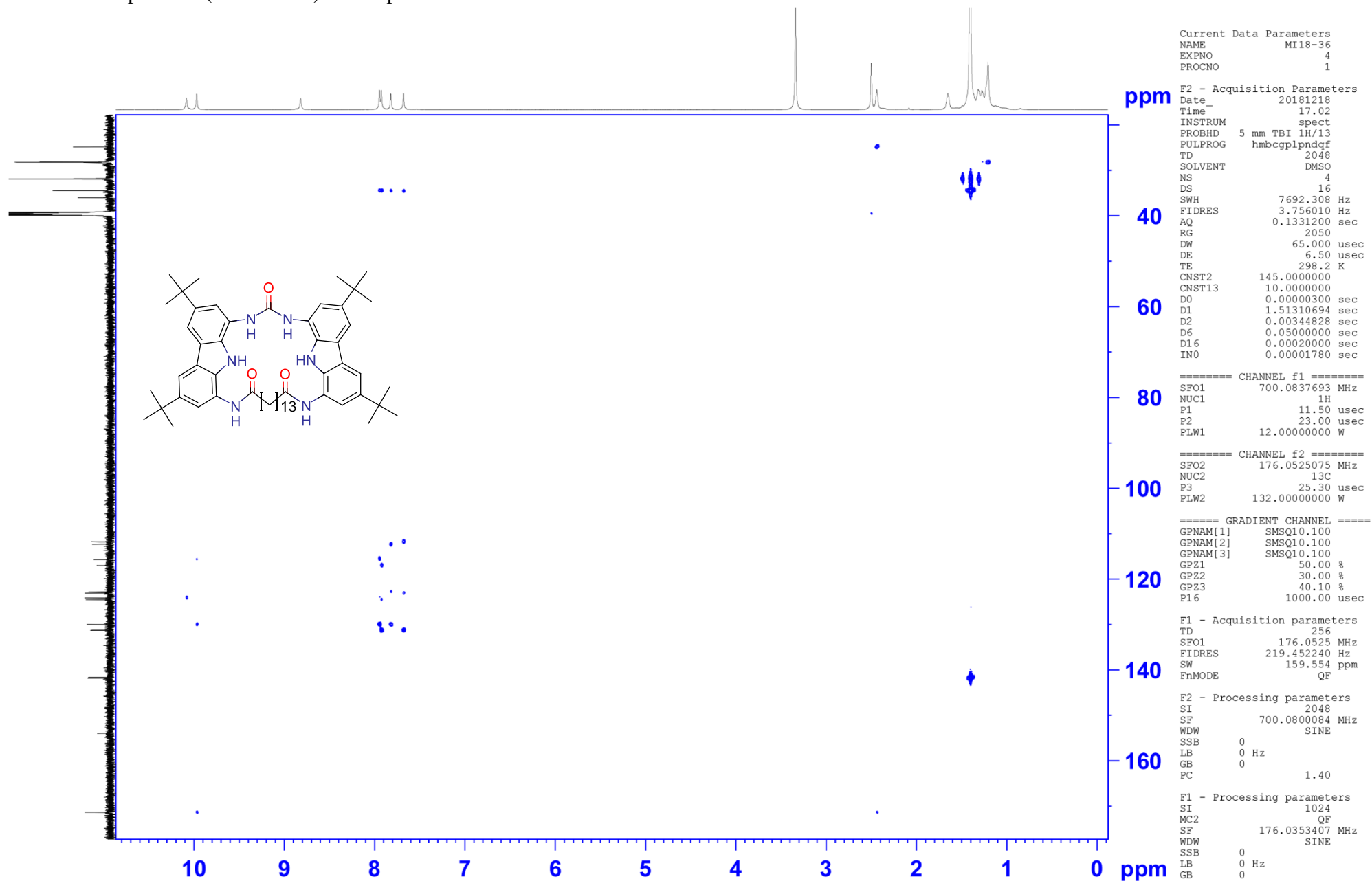
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GFZ2 20.10 %
GFZ3 11.00 %
GFZ4 -5.00 %
P16 1000.00 usec
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F1 - Acquisition parameters
TD 256
SFO1 176.0525 MHz
FIDRES 219.452240 Hz
SW 159.554 ppm
FMODE Echo-Antiecho

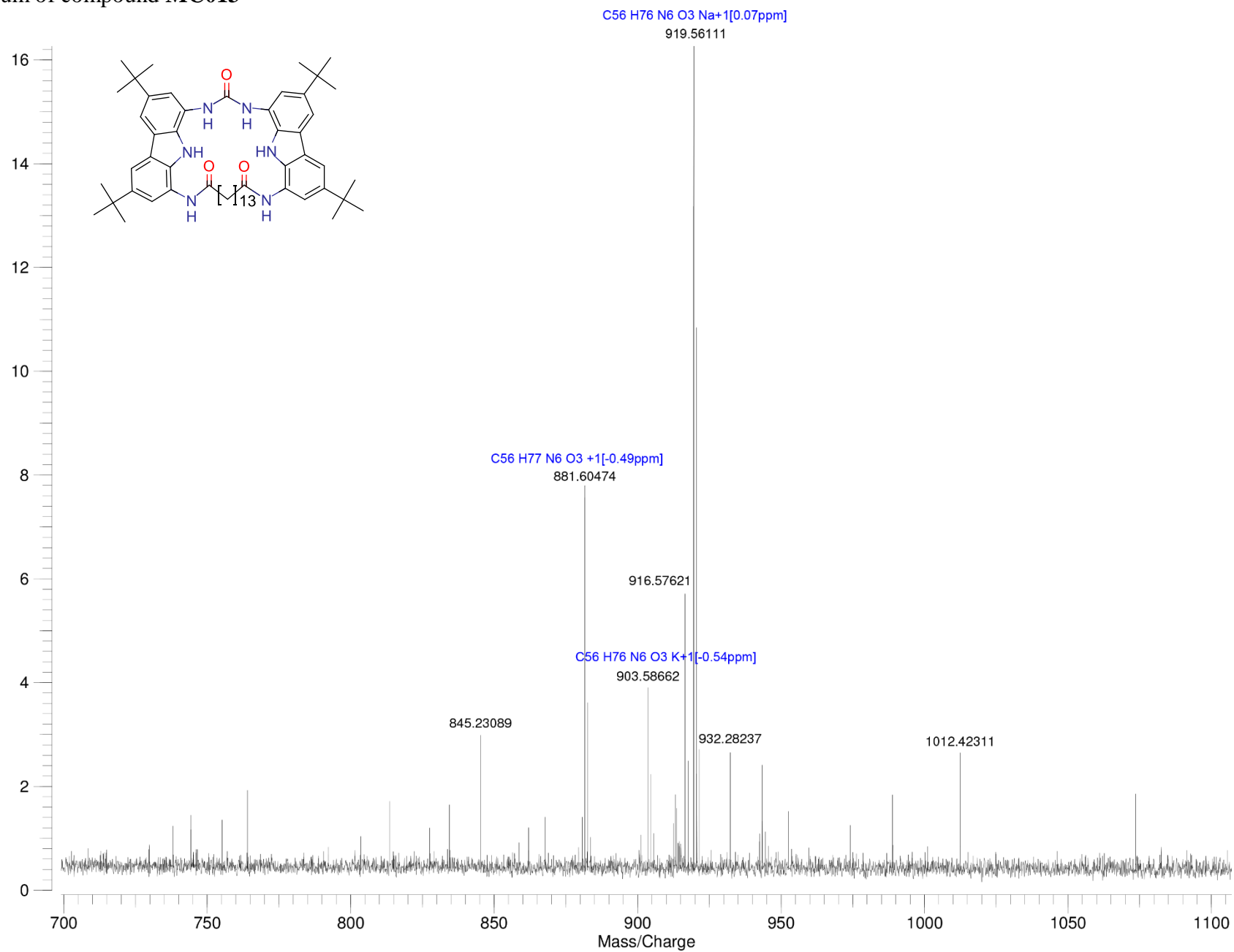
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SSB 2
LB 0 Hz
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F1 - Processing parameters
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WDW QSINE
SSB 2
LB 0 Hz
GB 0

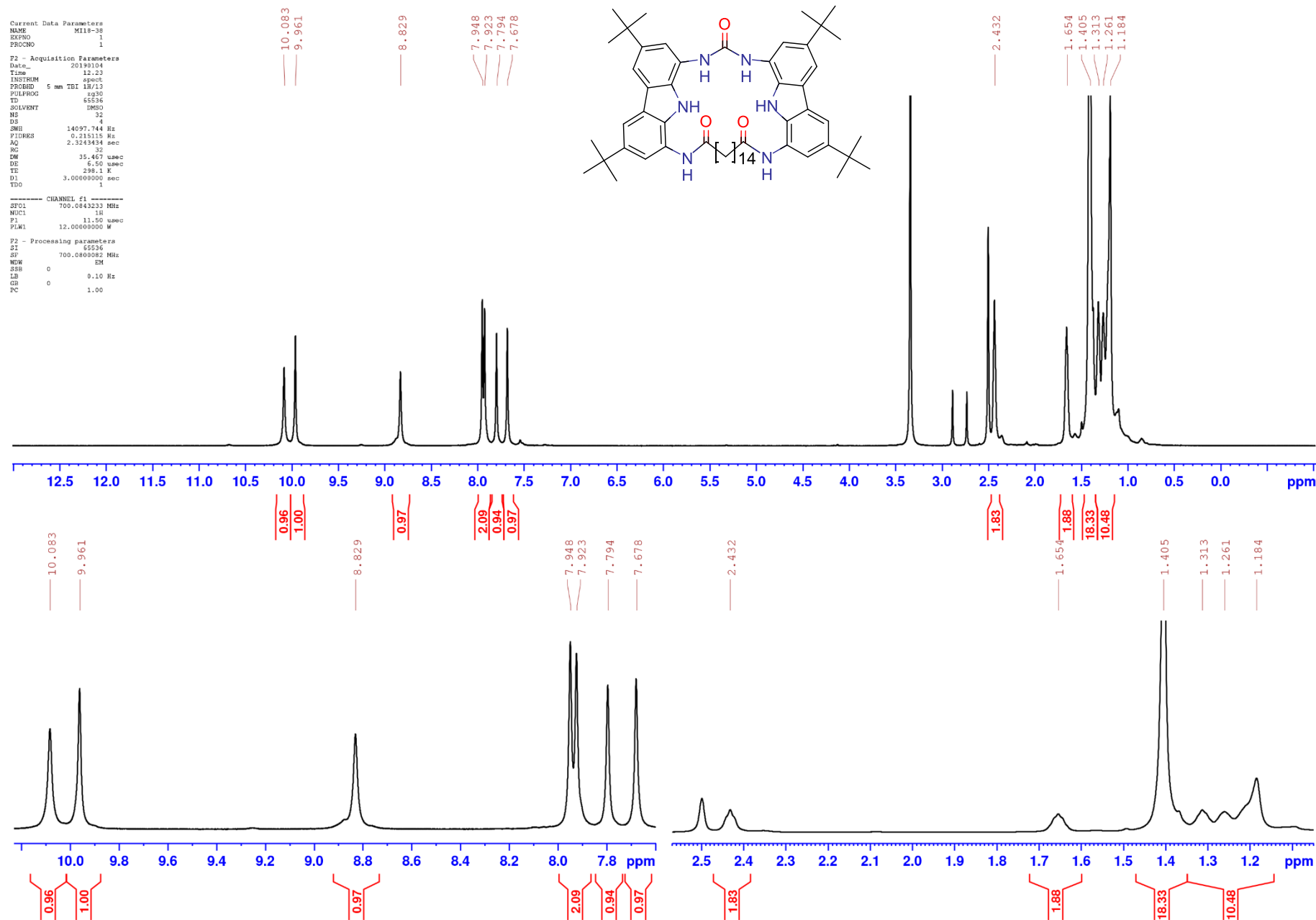
^1H - ^{13}C HMBC spectrum (700.1 MHz) of compound **MC013**



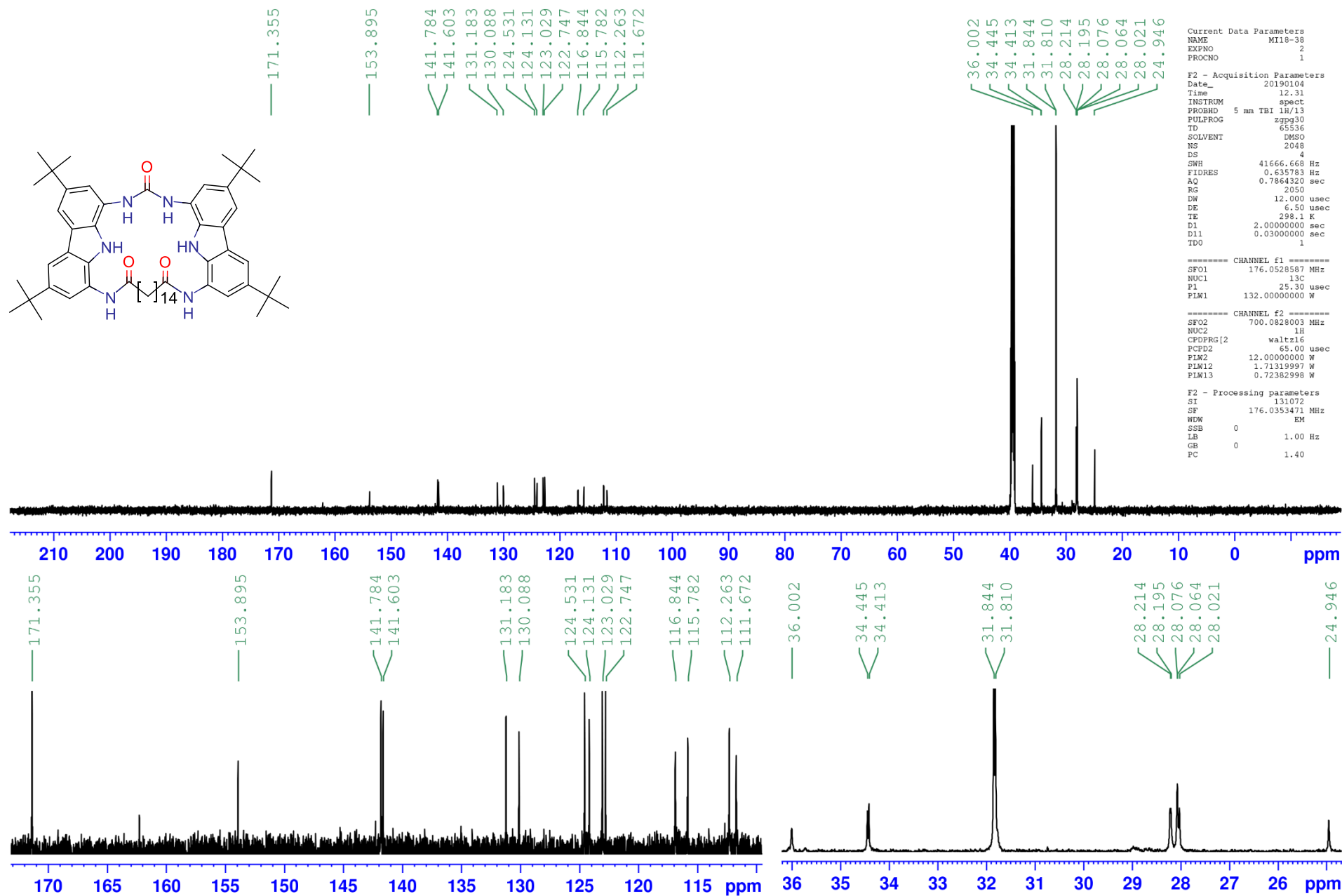
HRMS spectrum of compound **MC013**



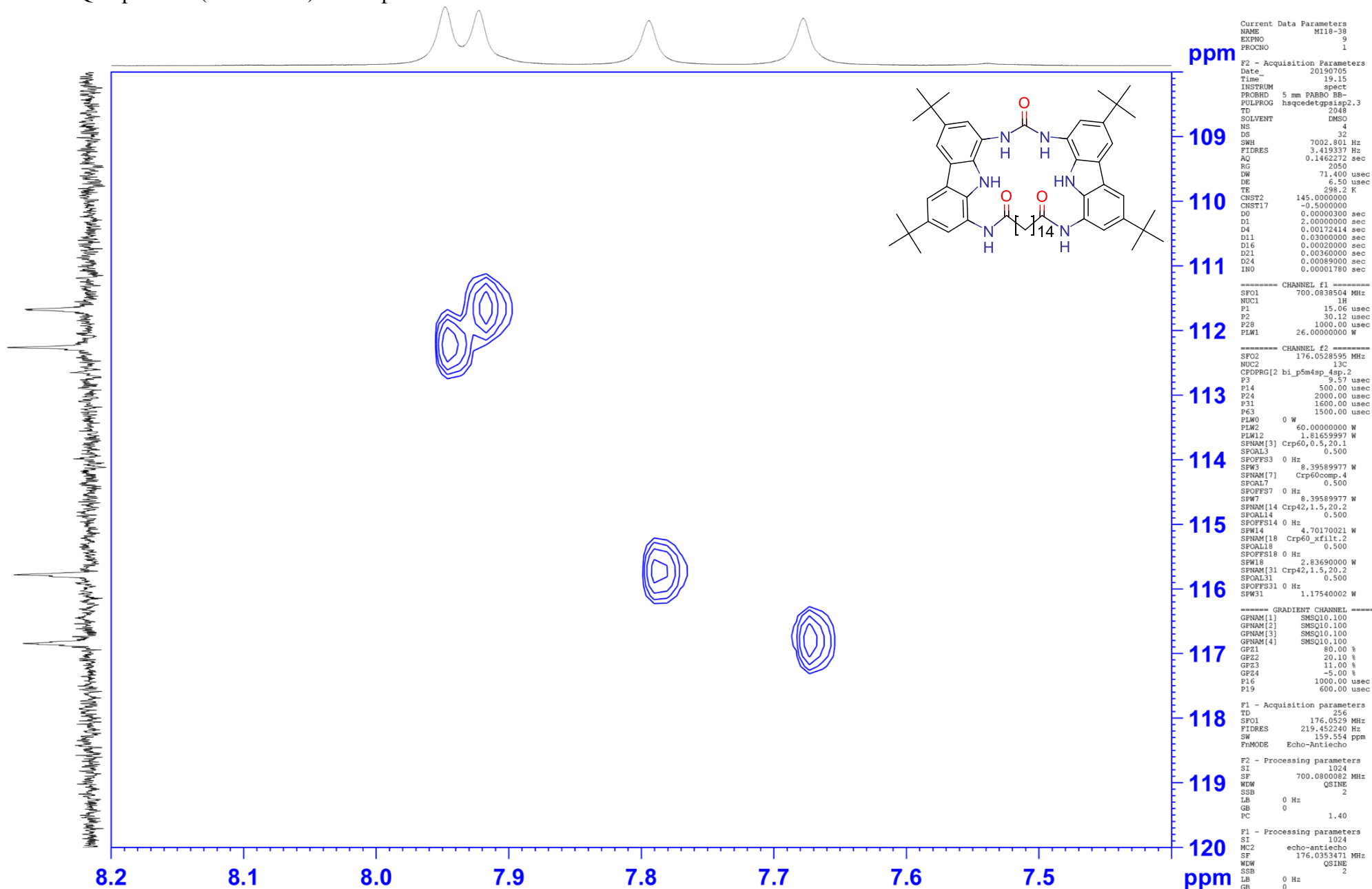
¹H NMR spectrum (700.1 MHz) of compound **MC014**



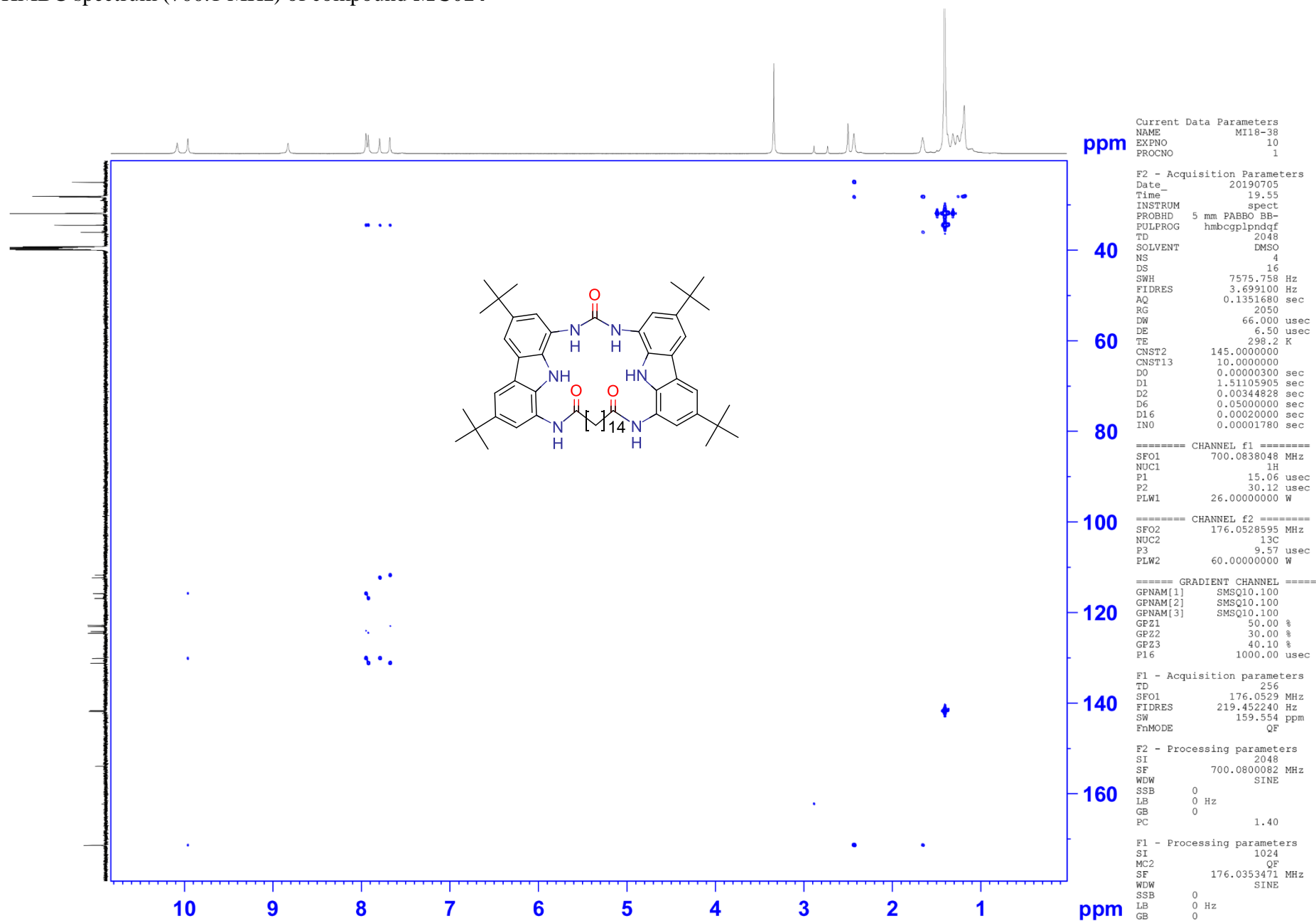
^{13}C NMR spectrum (700.1 MHz) of compound **MC014**



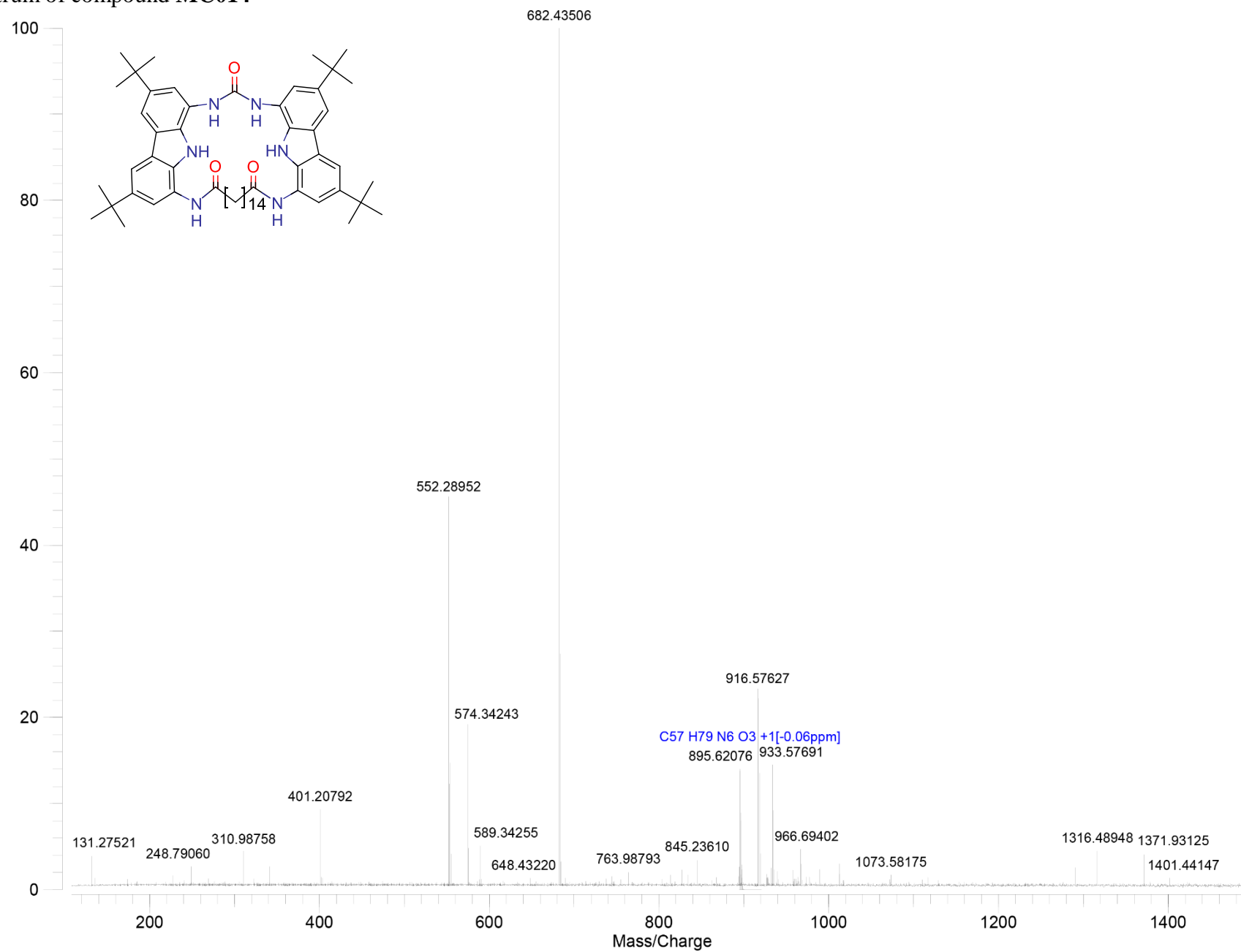
^1H - ^{13}C HSQC spectrum (700.1 MHz) of compound **MC014**



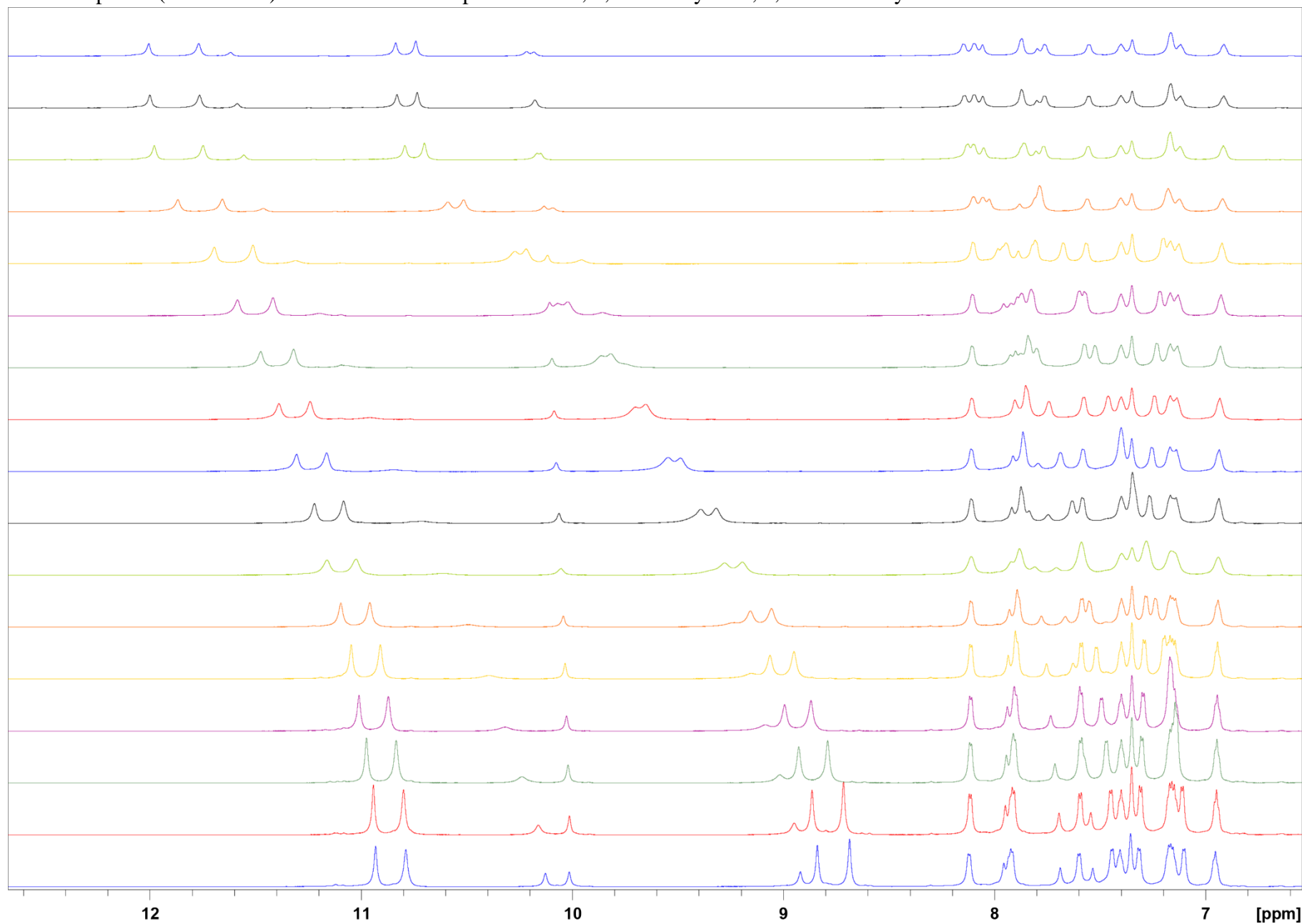
^1H - ^{13}C HMBC spectrum (700.1 MHz) of compound **MC014**



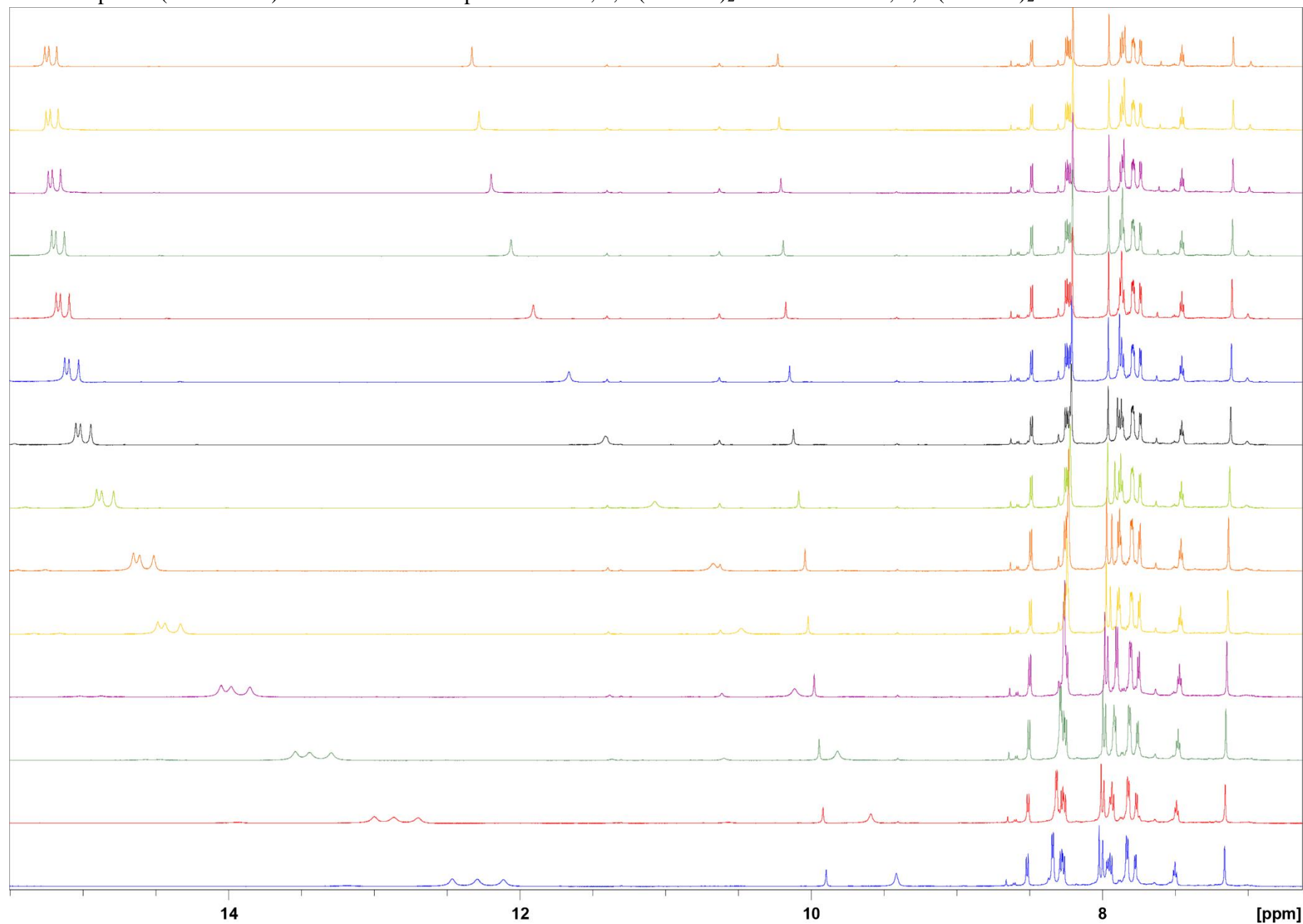
HRMS spectrum of compound MC014



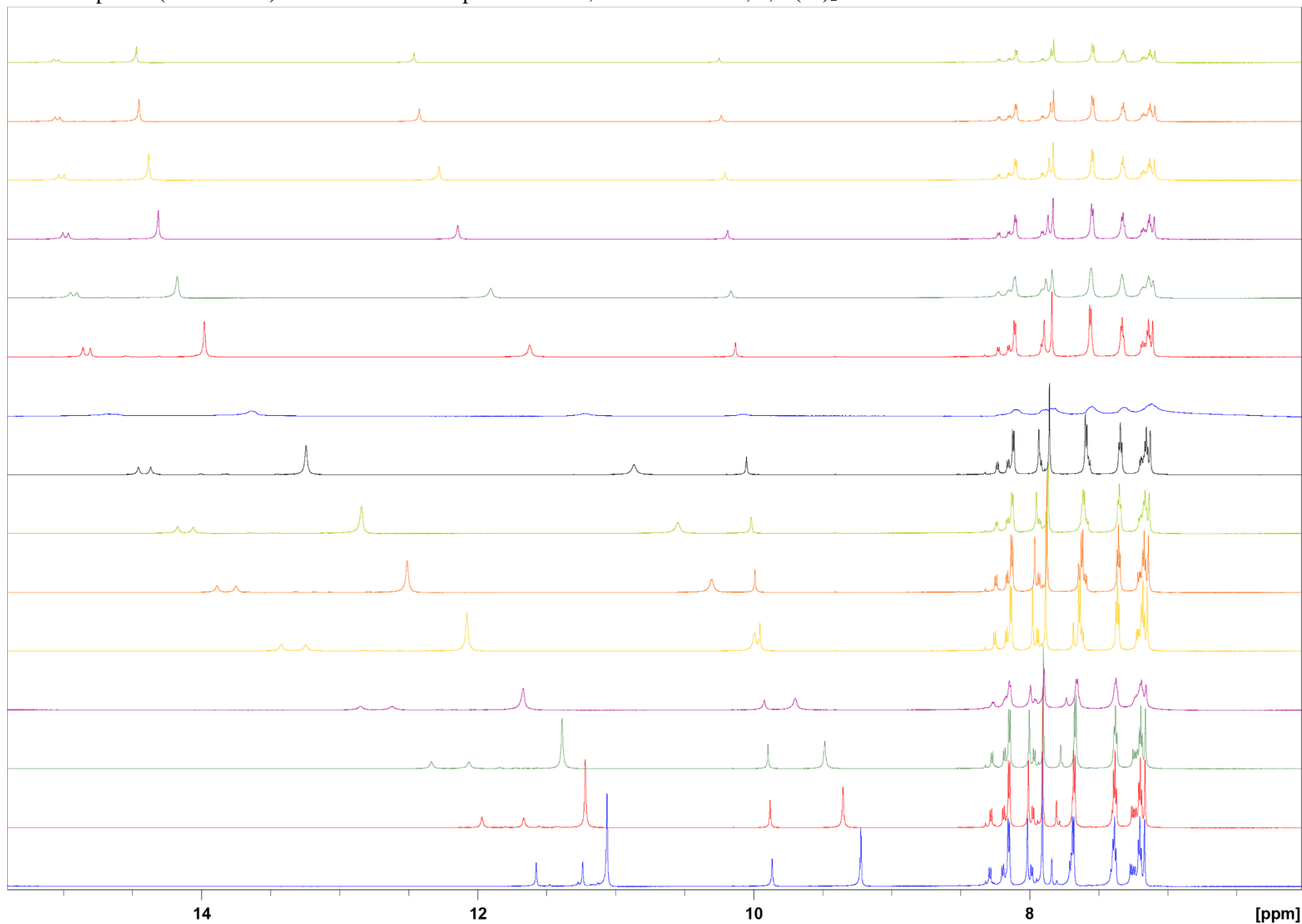
Stacked ^1H NMR spectra (700.1 MHz) of a mixture of receptors **CZ016**; 1,3-diindolylurea; 1,3-dicarbazolylurea + TBA-acetate



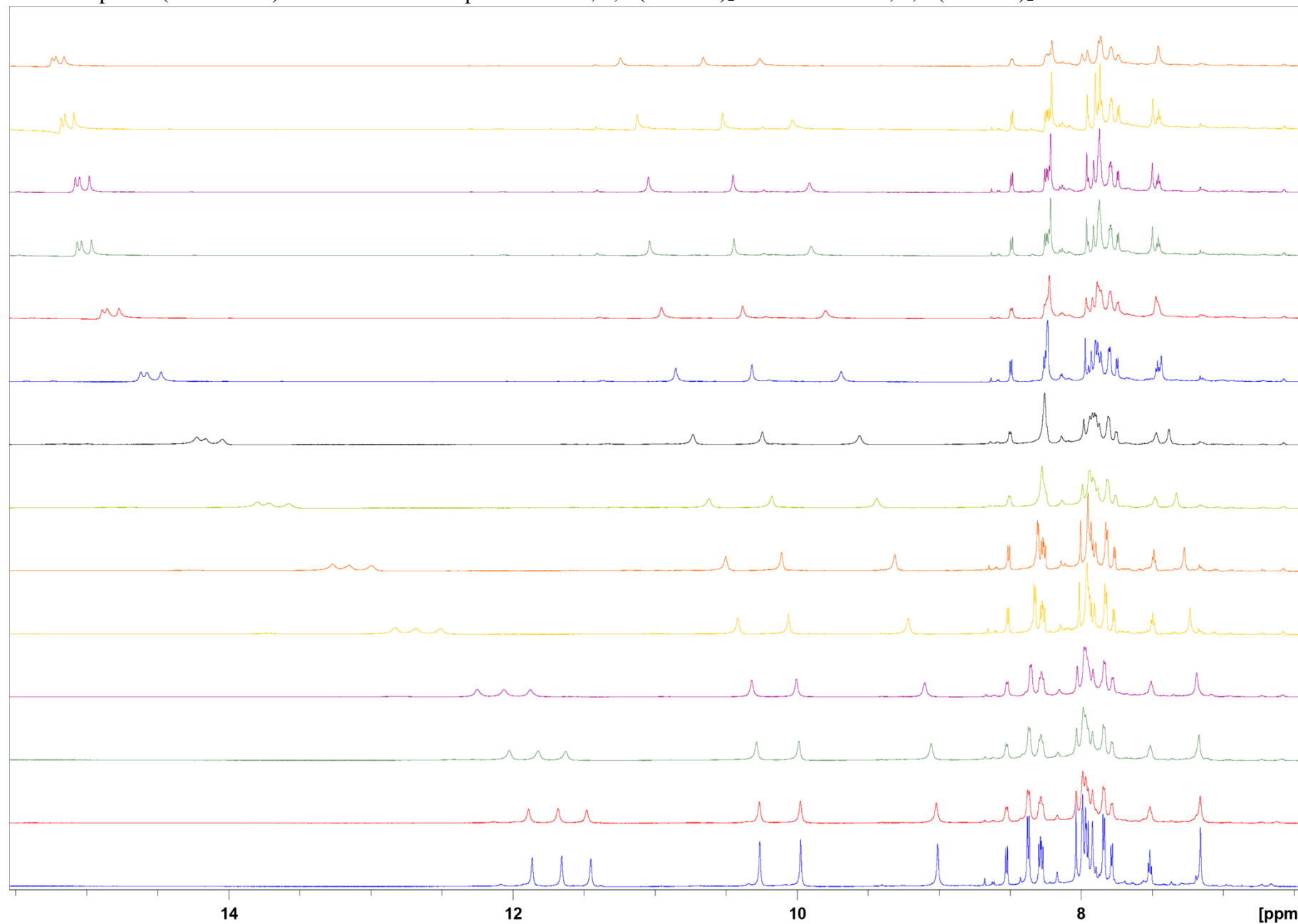
Stacked ^1H NMR spectra (700.1 MHz) of a mixture of receptors **MC001**; 2,7-(COOBu) $_2$ -indolocarbazole; 2,9-(COOBu) $_2$ -indolocarbazole + TBA-acetate



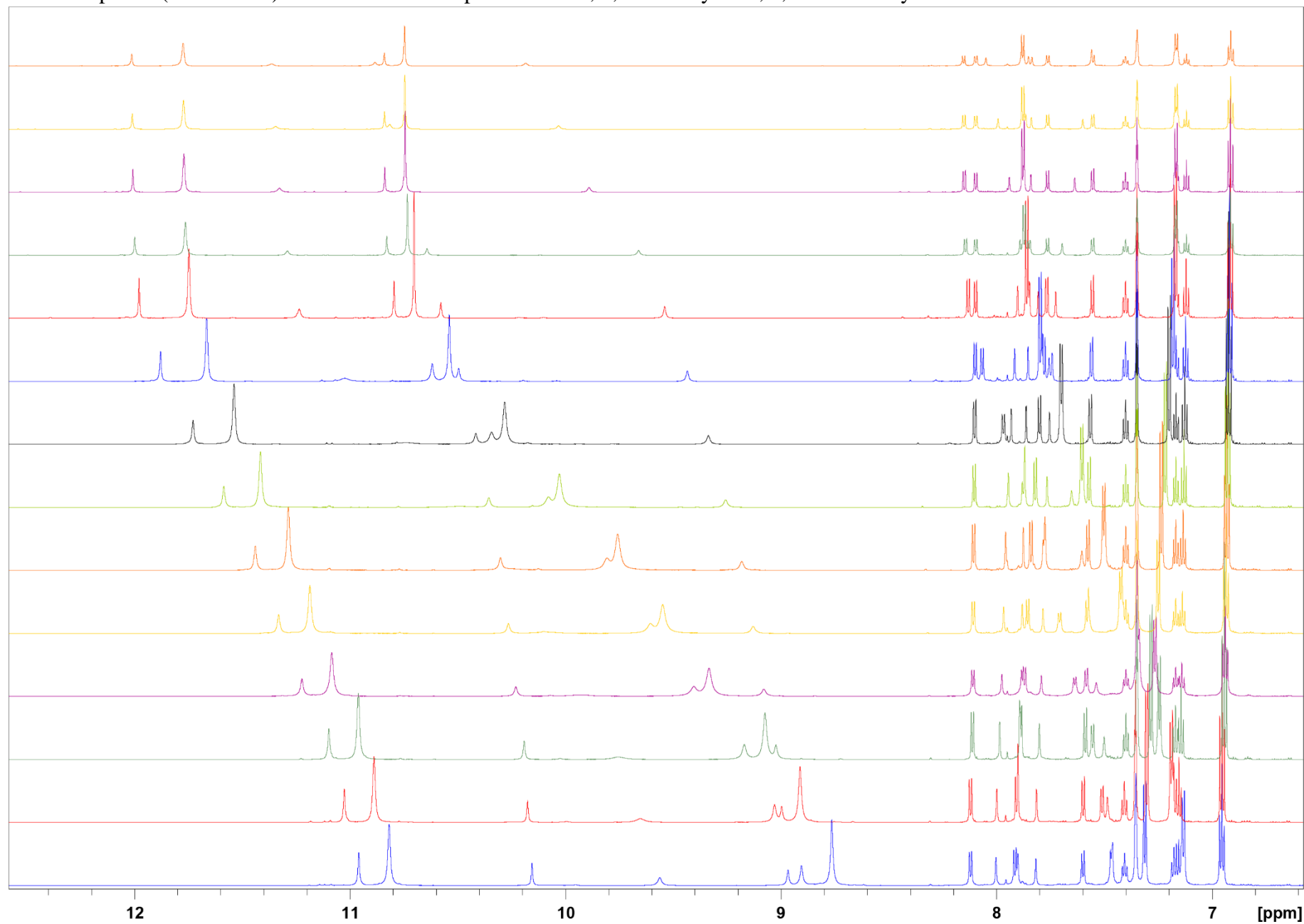
Stacked ^1H NMR spectra (700.1 MHz) of a mixture of receptors **MC002**; indolocarbazole; 2,7-(Cl)₂-indolocarbazole + TBA-acetate



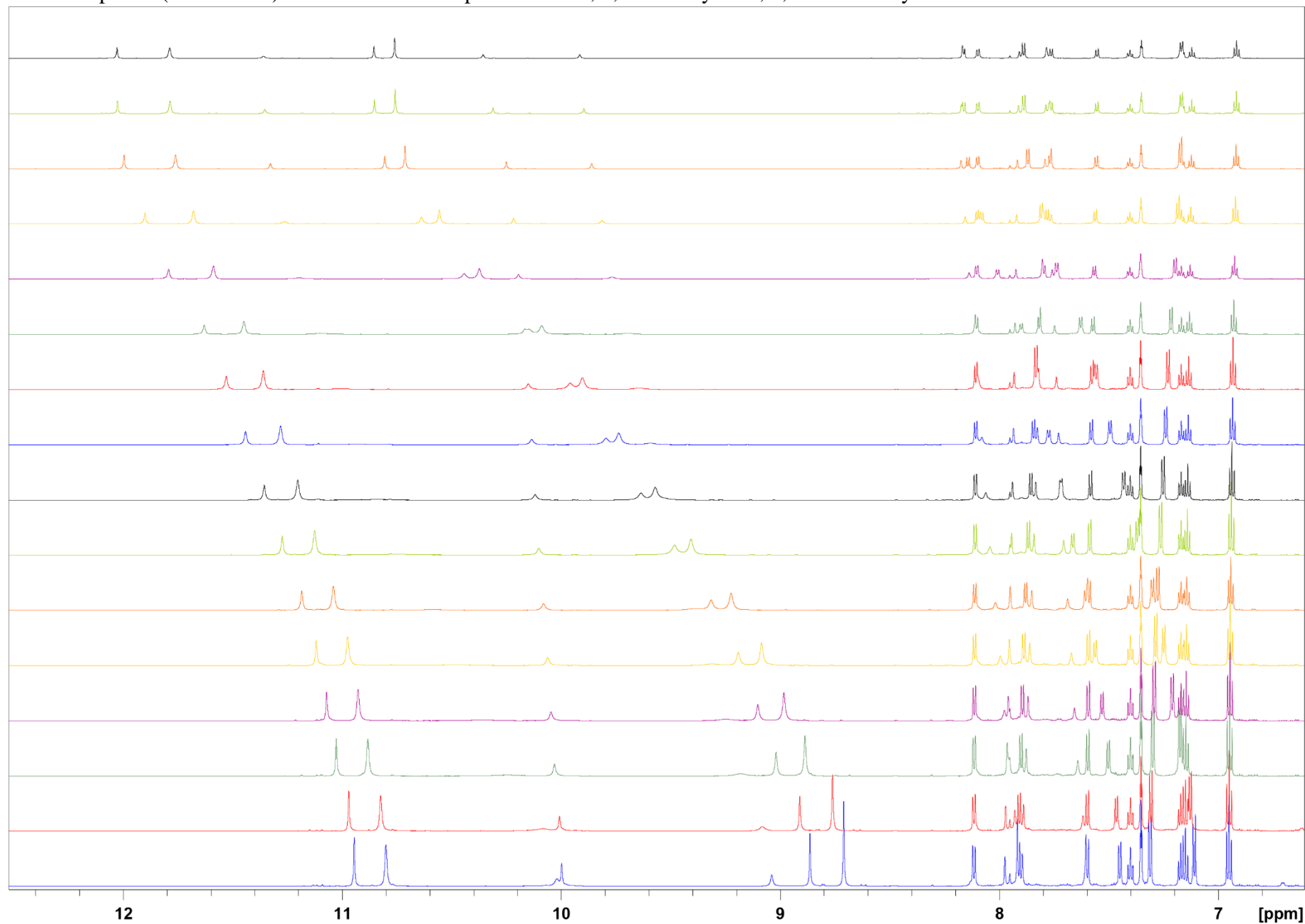
Stacked ^1H NMR spectra (700.1 MHz) of a mixture of receptors **MC003**; 2,7-(COOBu) $_2$ -indolocarbazole; 2,9-(COOBu) $_2$ -indolocarbazole + TBA-acetate



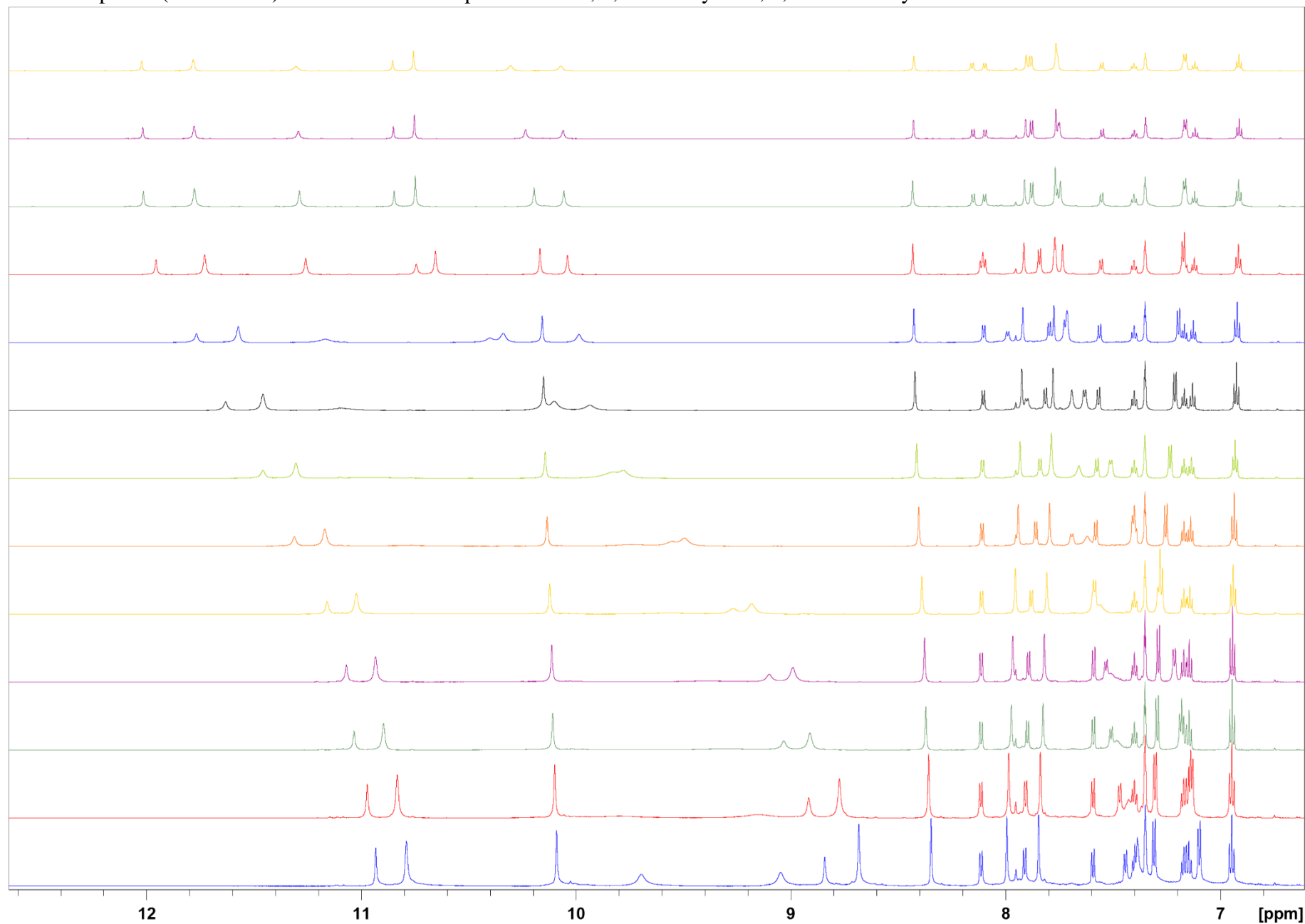
Stacked ^1H NMR spectra (700.1 MHz) of a mixture of receptors **MC004**; 1,3-diindolylurea; 1,3-dicarbazolylurea + TBA-acetate



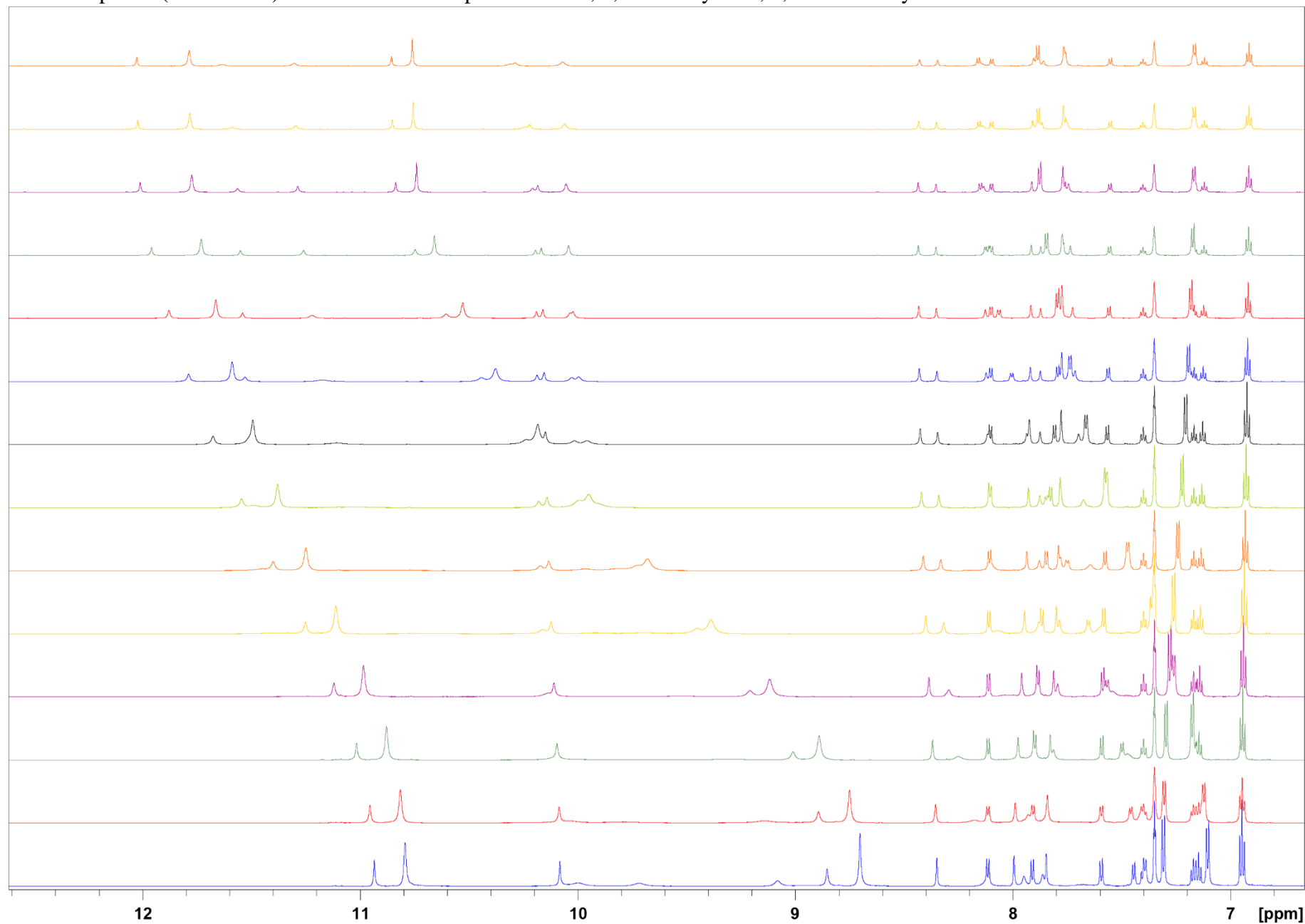
Stacked ^1H NMR spectra (700.1 MHz) of a mixture of receptors **MC005**; 1,3-diindolylurea; 1,3-dicarbazolylurea + TBA-acetate



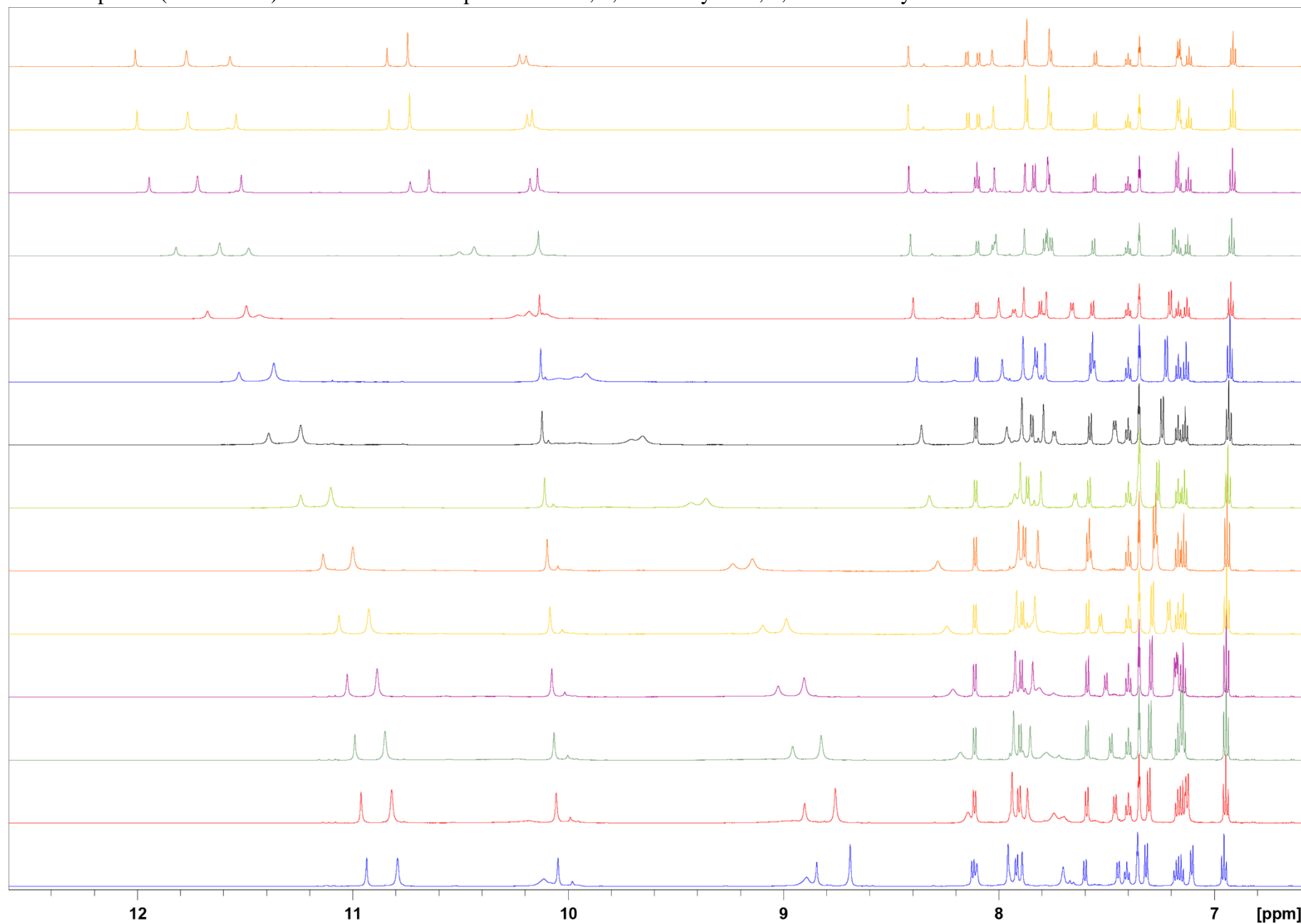
Stacked ^1H NMR spectra (700.1 MHz) of a mixture of receptors **MC006**; 1,3-diindolylurea; 1,3-dicarbazolylurea + TBA-acetate



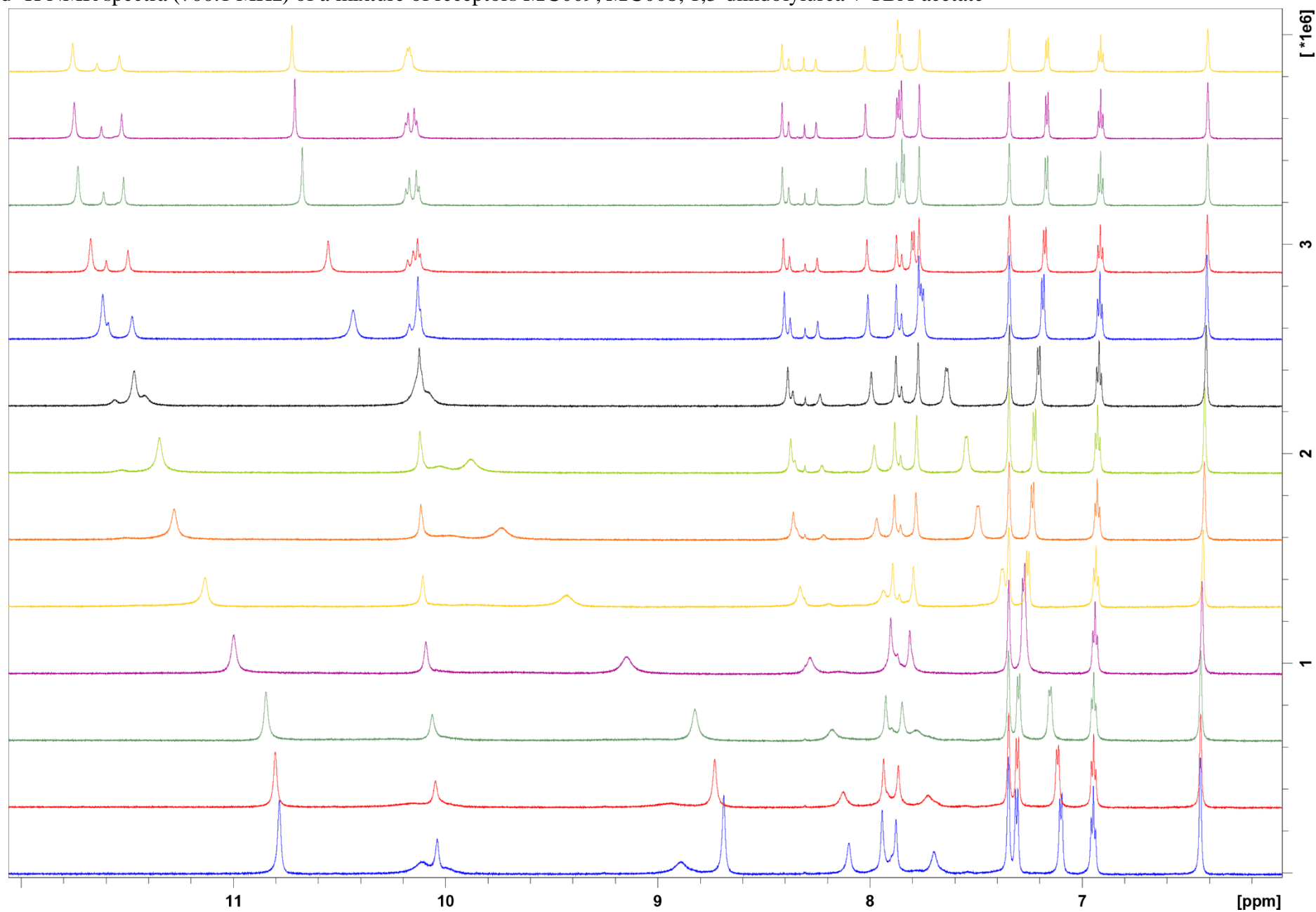
Stacked ^1H NMR spectra (700.1 MHz) of a mixture of receptors **MC007**; 1,3-diindolylurea; 1,3-dicarbazolylurea + TBA-acetate



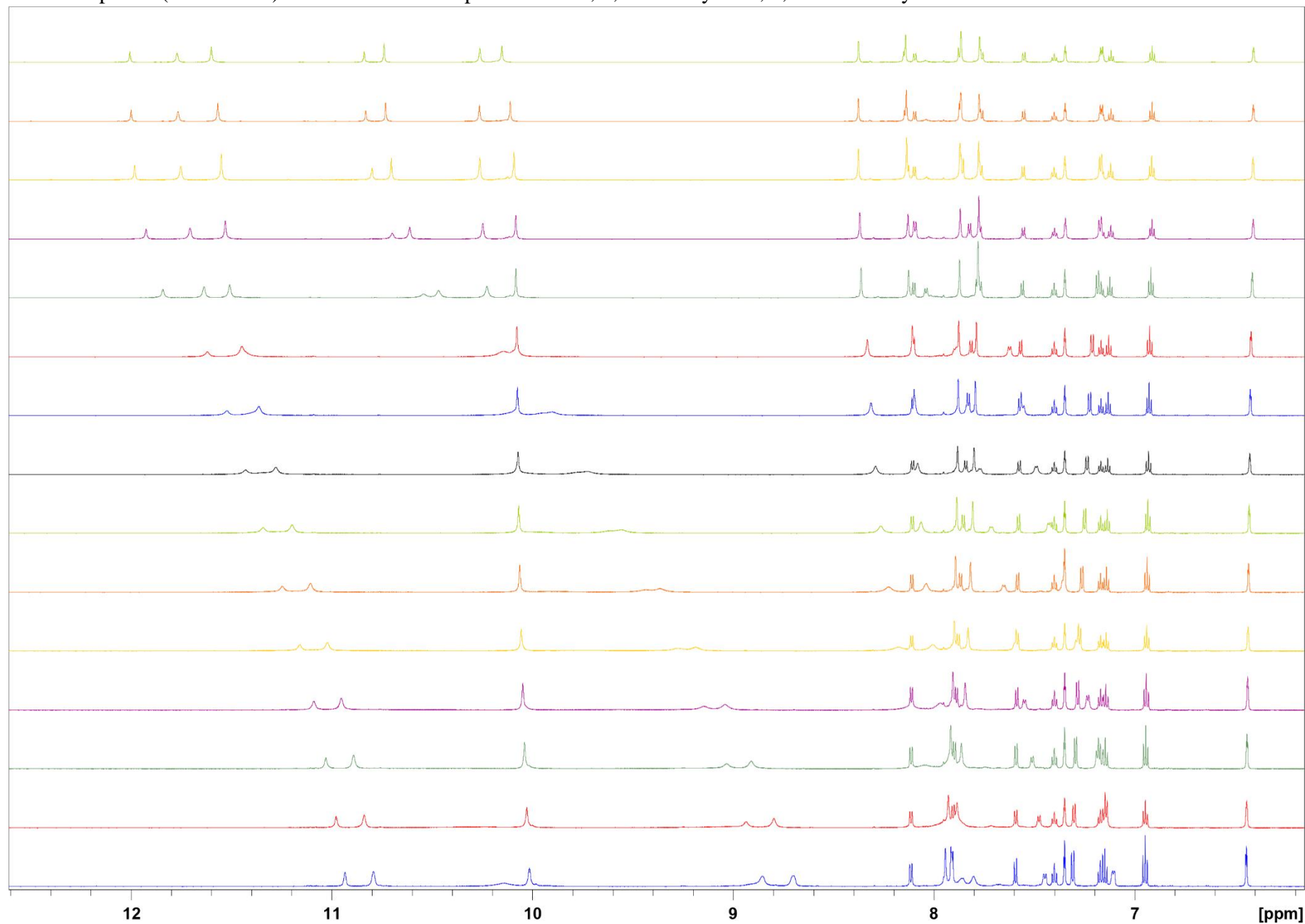
Stacked ^1H NMR spectra (700.1 MHz) of a mixture of receptors **MC008**; 1,3-diindolylurea; 1,3-dicarbazolylurea + TBA-acetate



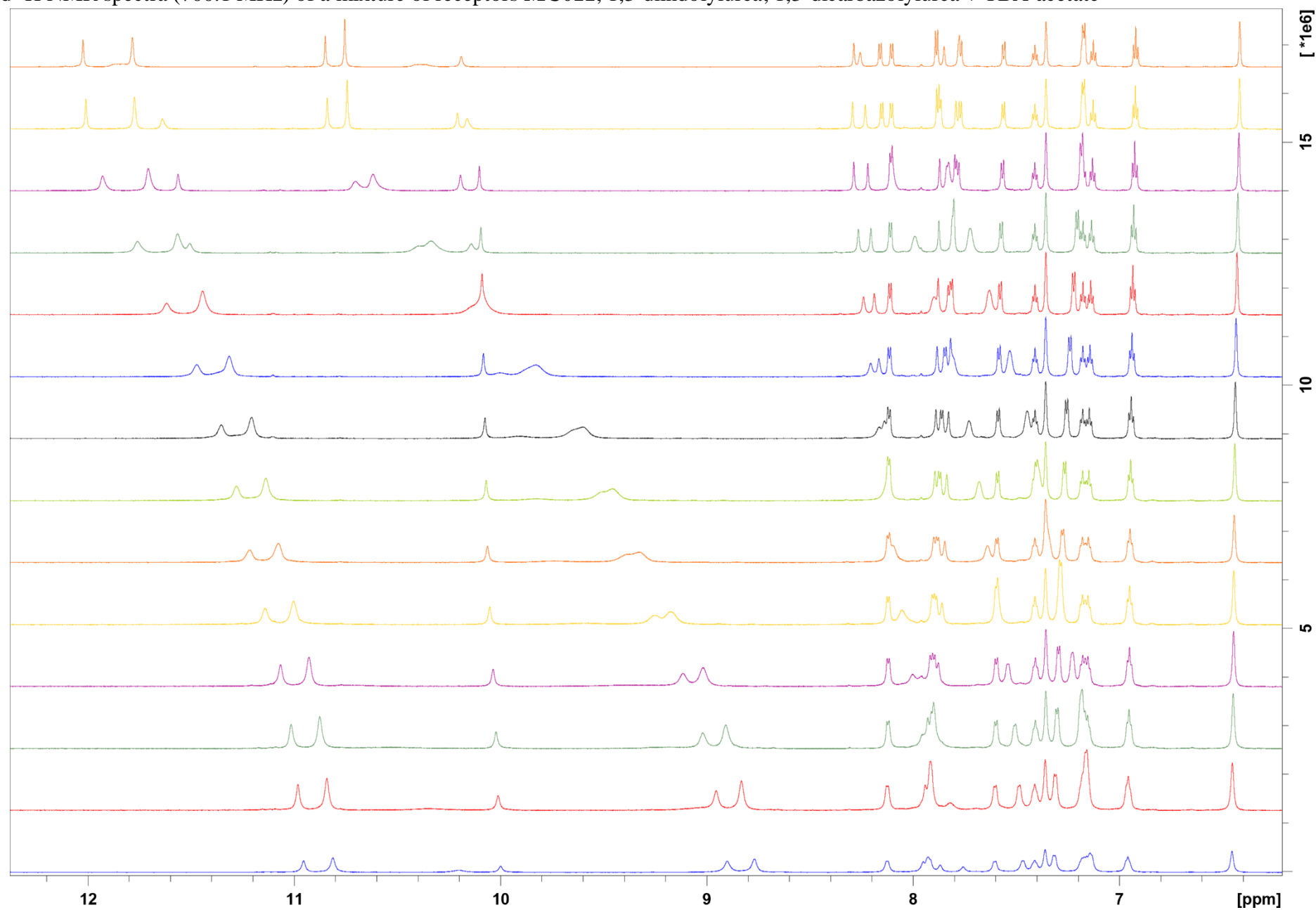
Stacked ^1H NMR spectra (700.1 MHz) of a mixture of receptors **MC009**; **MC008**; 1,3-diindolylurea + TBA-acetate



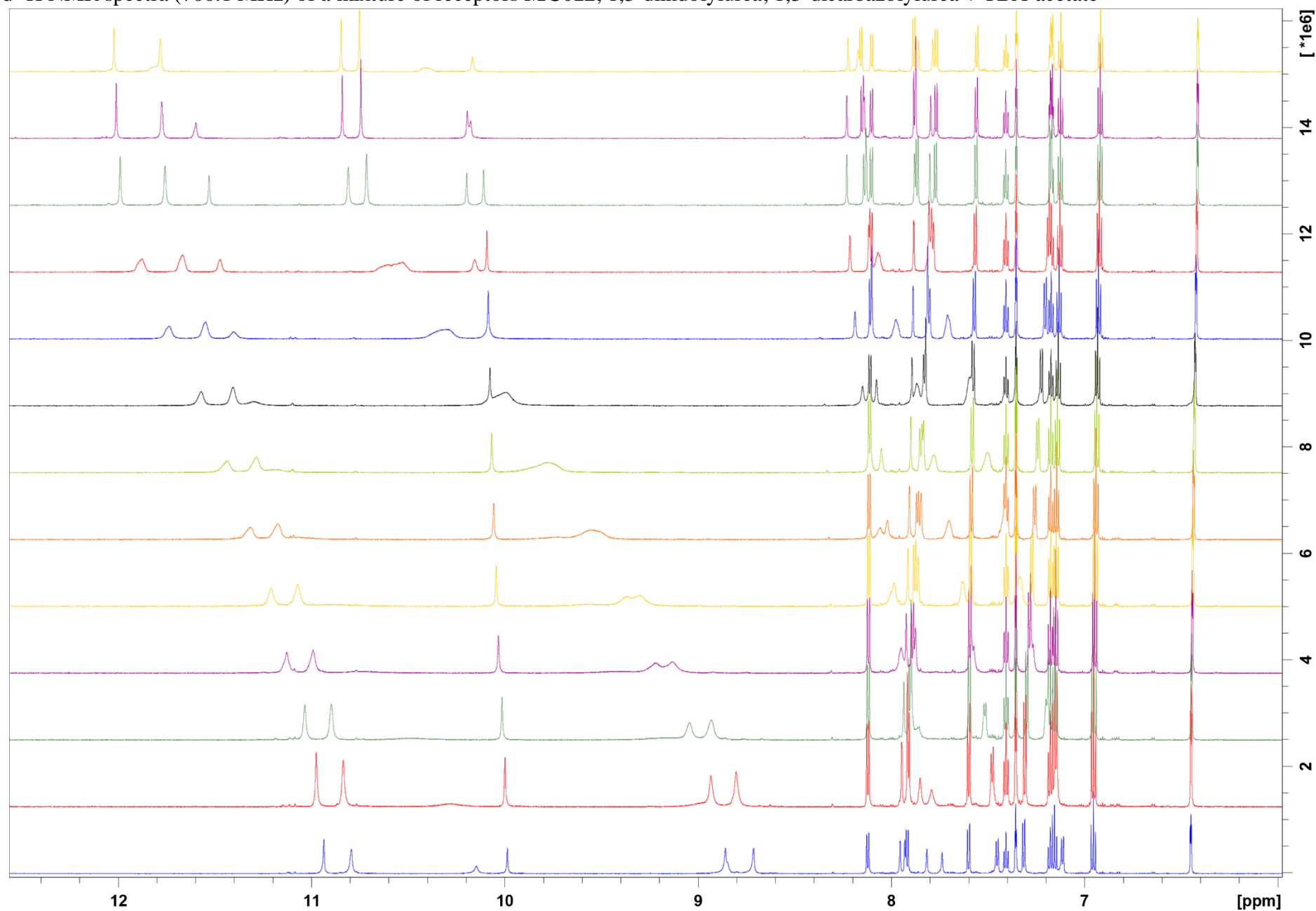
Stacked ^1H NMR spectra (700.1 MHz) of a mixture of receptors **MC010**; 1,3-diindolylurea; 1,3-dicarbazolylurea + TBA-acetate



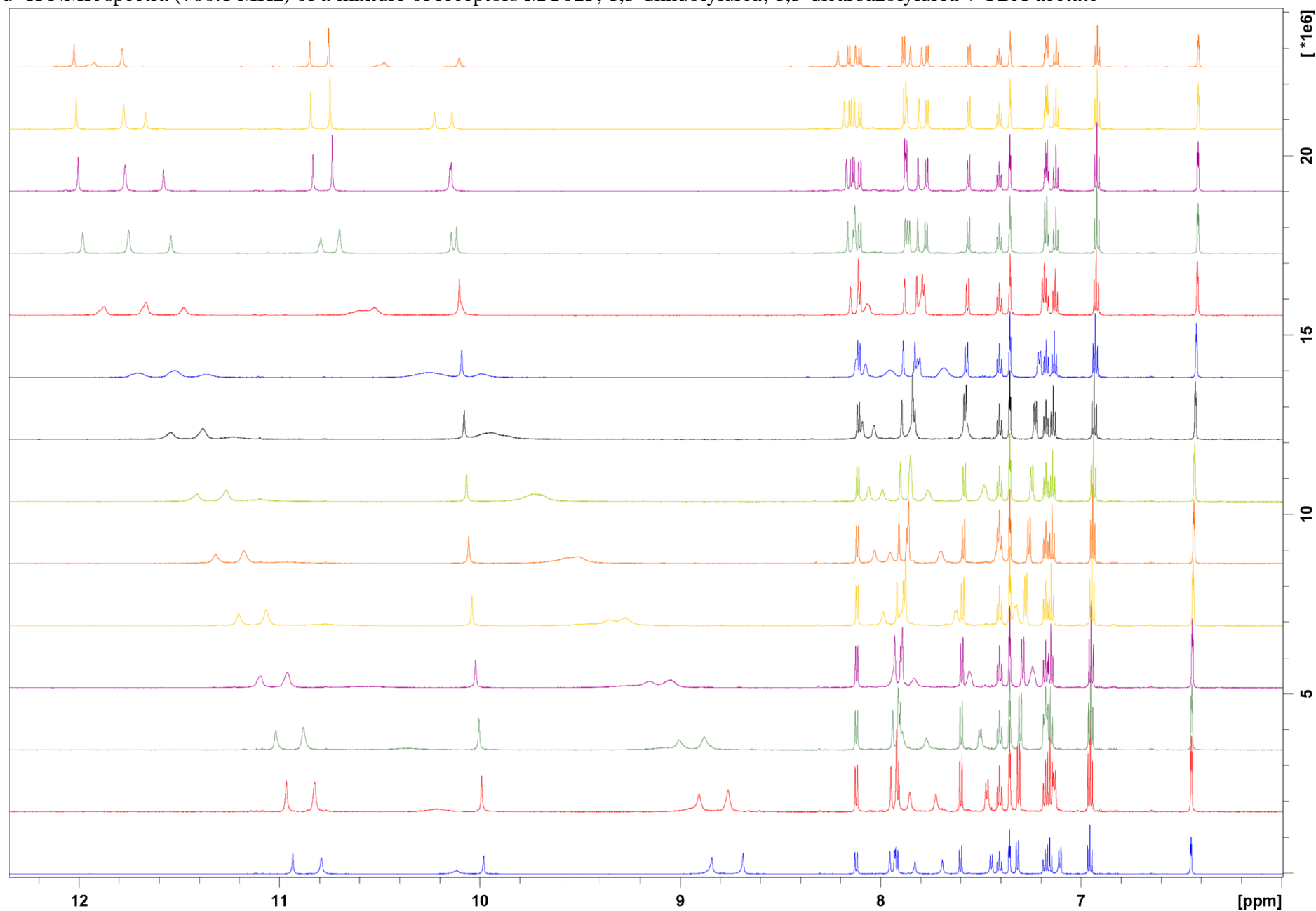
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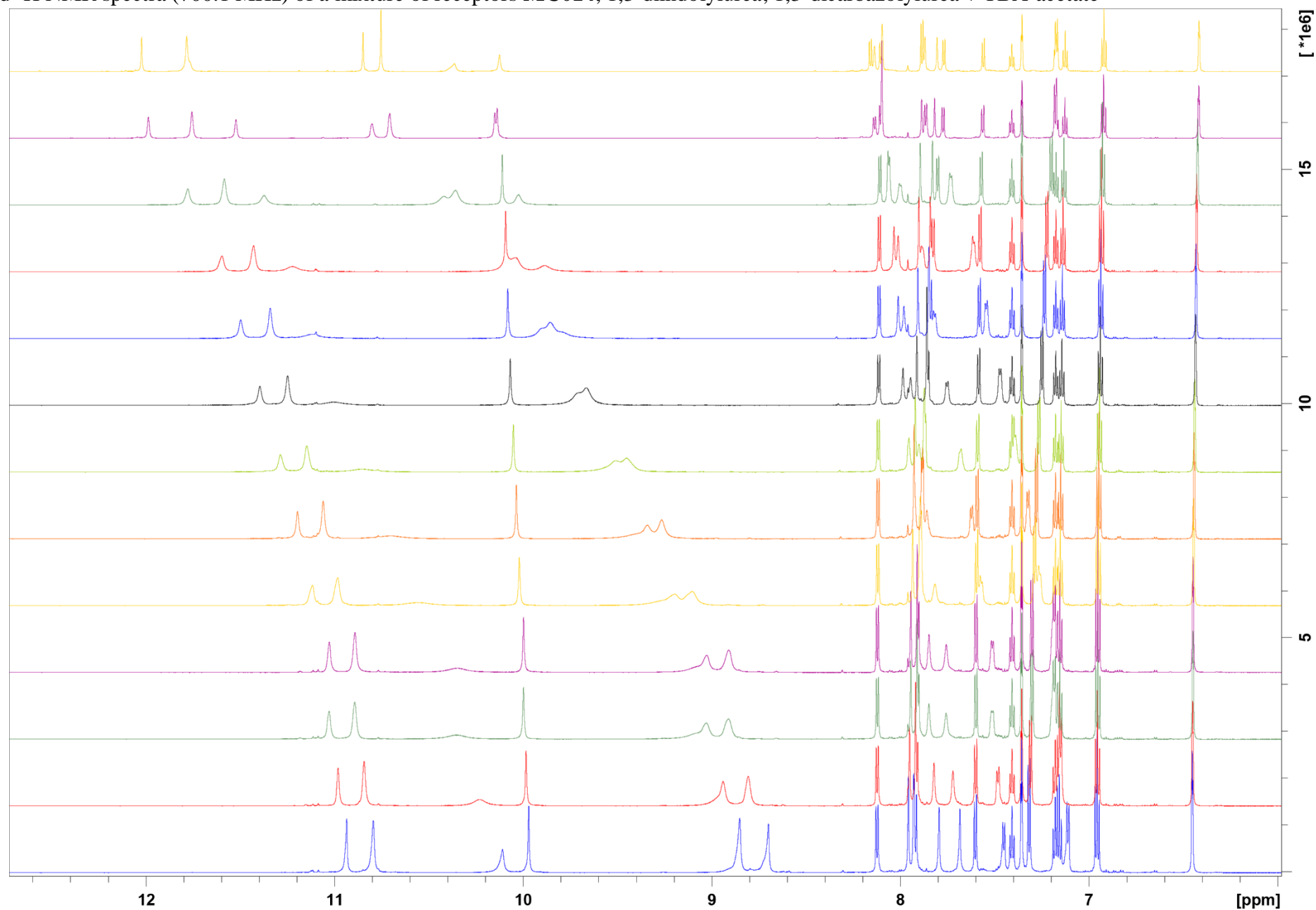
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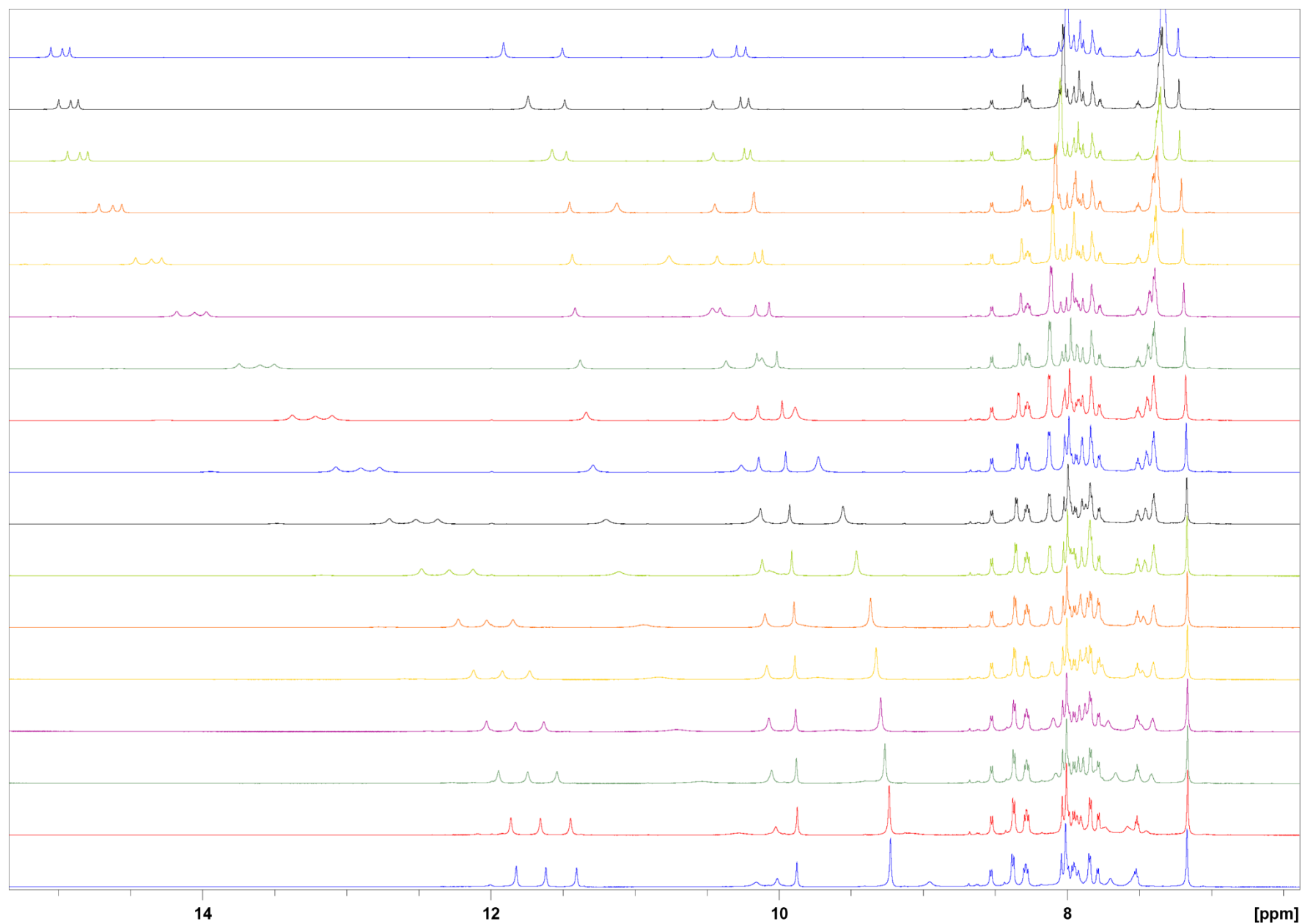
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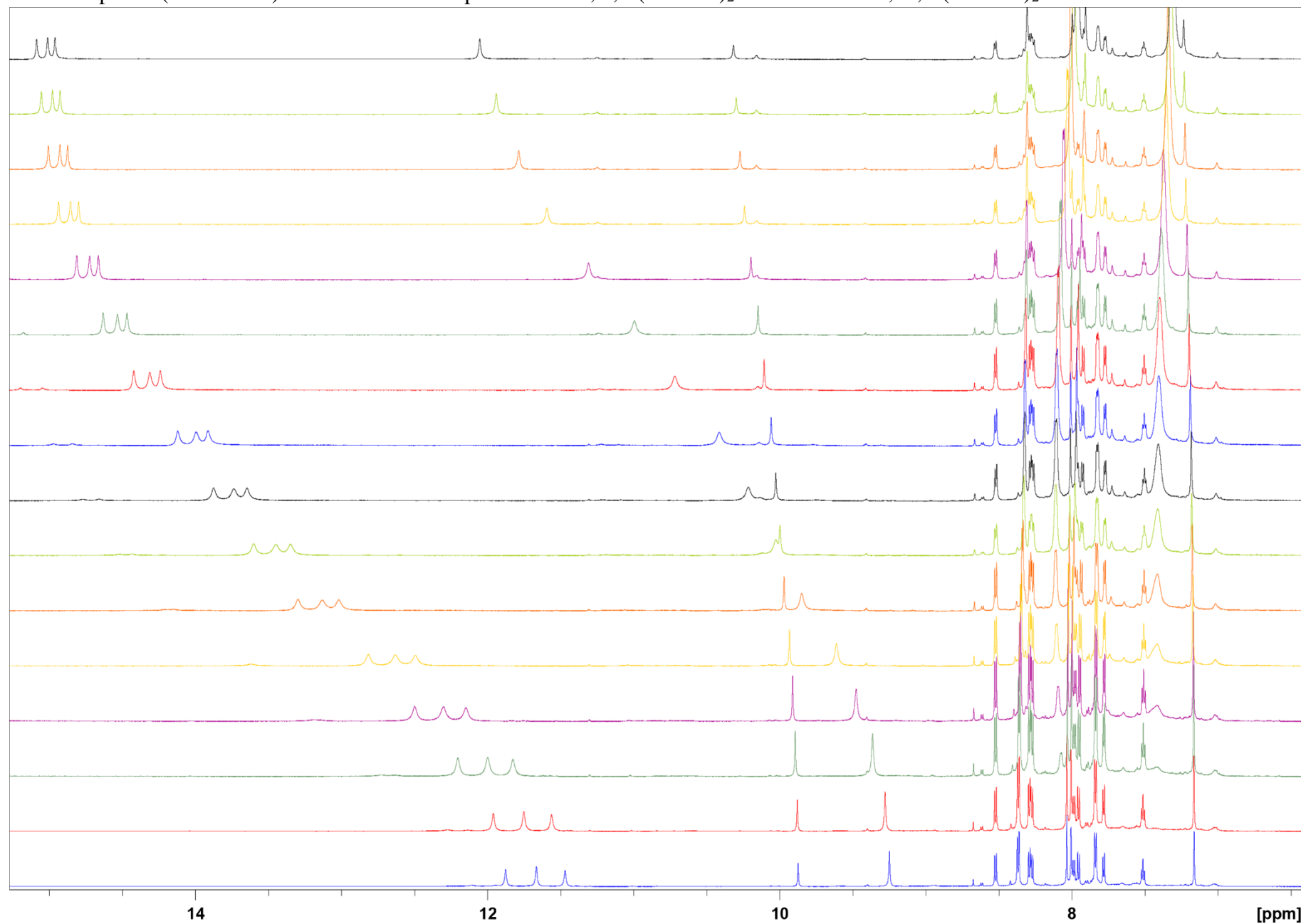
Stacked ^1H NMR spectra (700.1 MHz) of a mixture of receptors **MC014**; 1,3-diindolylurea; 1,3-dicarbazolylurea + TBA-acetate



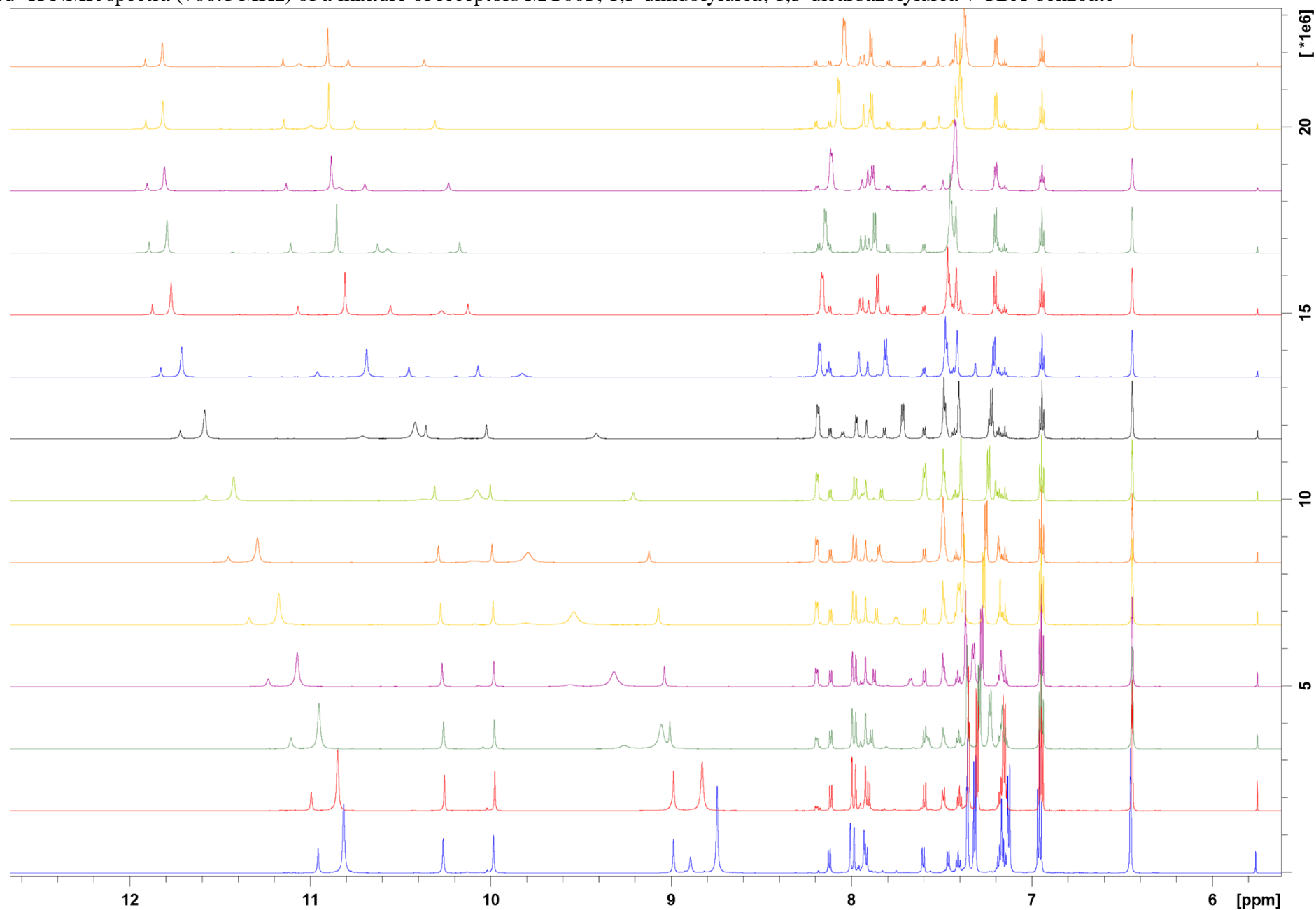
Stacked ^1H NMR spectra (700.1 MHz) of a mixture of receptors **CZ016**; **MC002**; 2,7-(COOBu) $_2$ -indolocarbazole; 2,9-(COOBu) $_2$ -indolocarbazole + TBA-benzoate



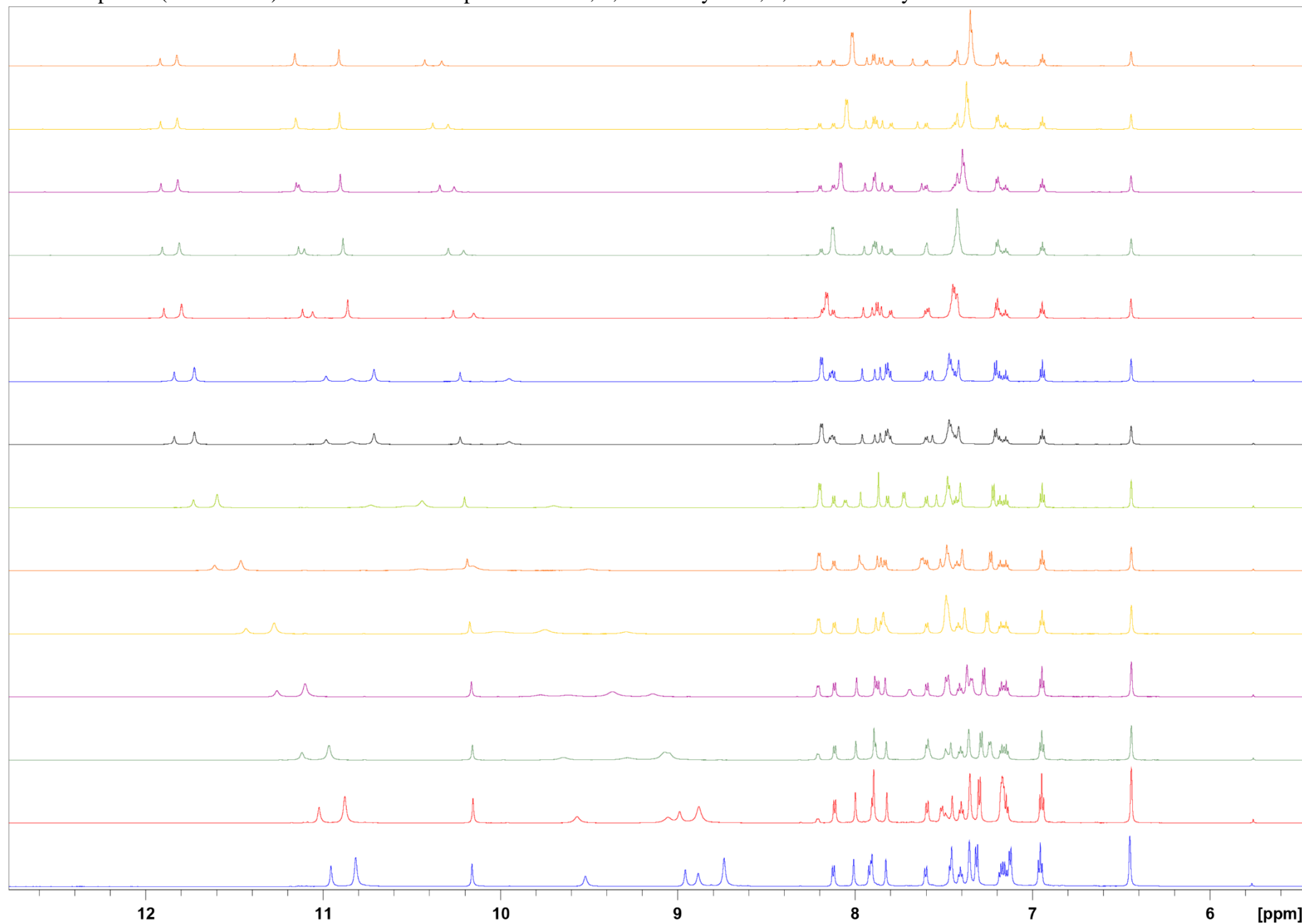
Stacked ^1H NMR spectra (700.1 MHz) of a mixture of receptors **MC001**; 2,7-(COOBu) $_2$ -indolocarbazole; 2,9-(COOBu) $_2$ -indolocarbazole + TBA-benzoate



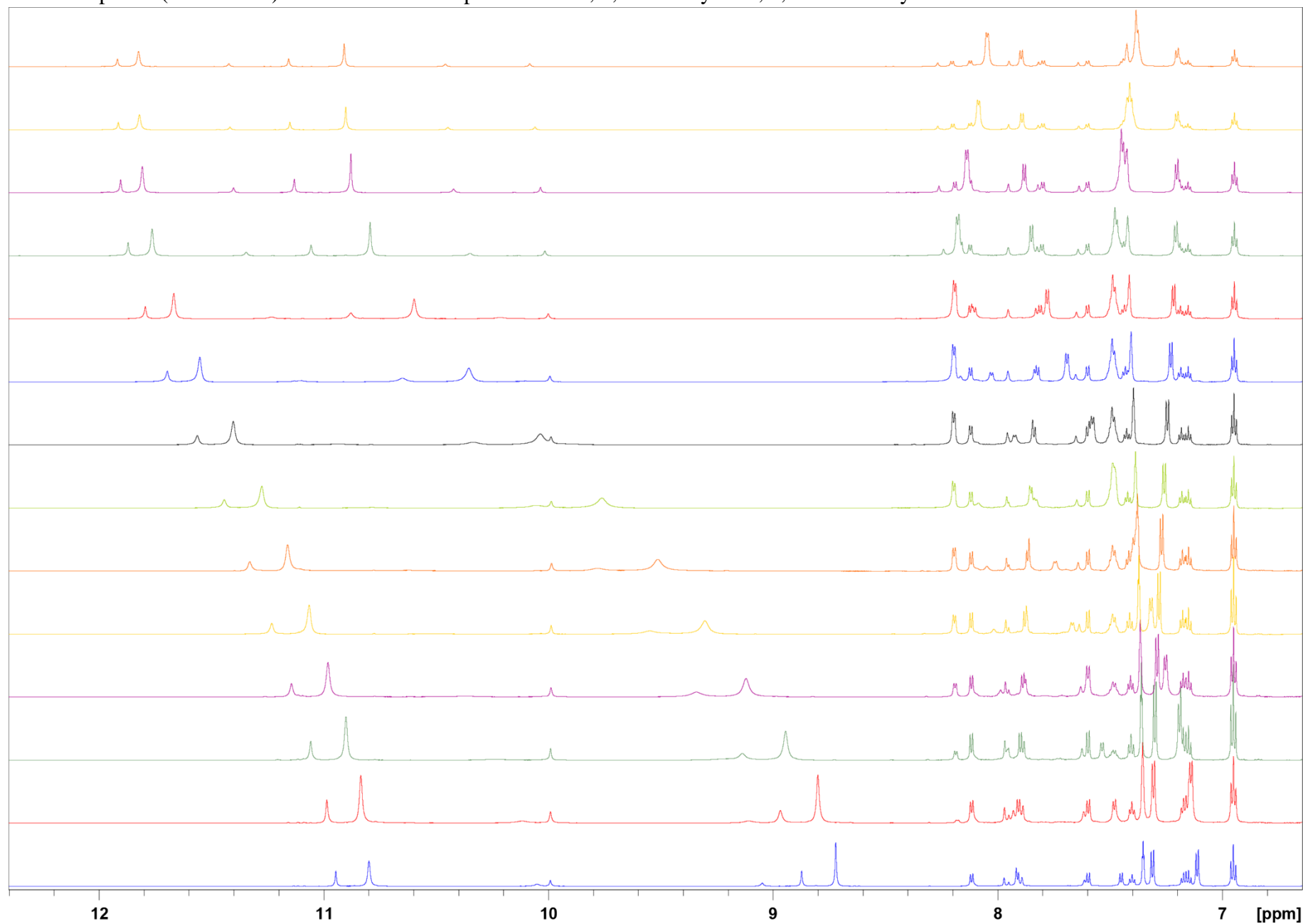
Stacked ^1H NMR spectra (700.1 MHz) of a mixture of receptors **MC003**; 1,3-diindolylurea; 1,3-dicarbazolylurea + TBA-benzoate



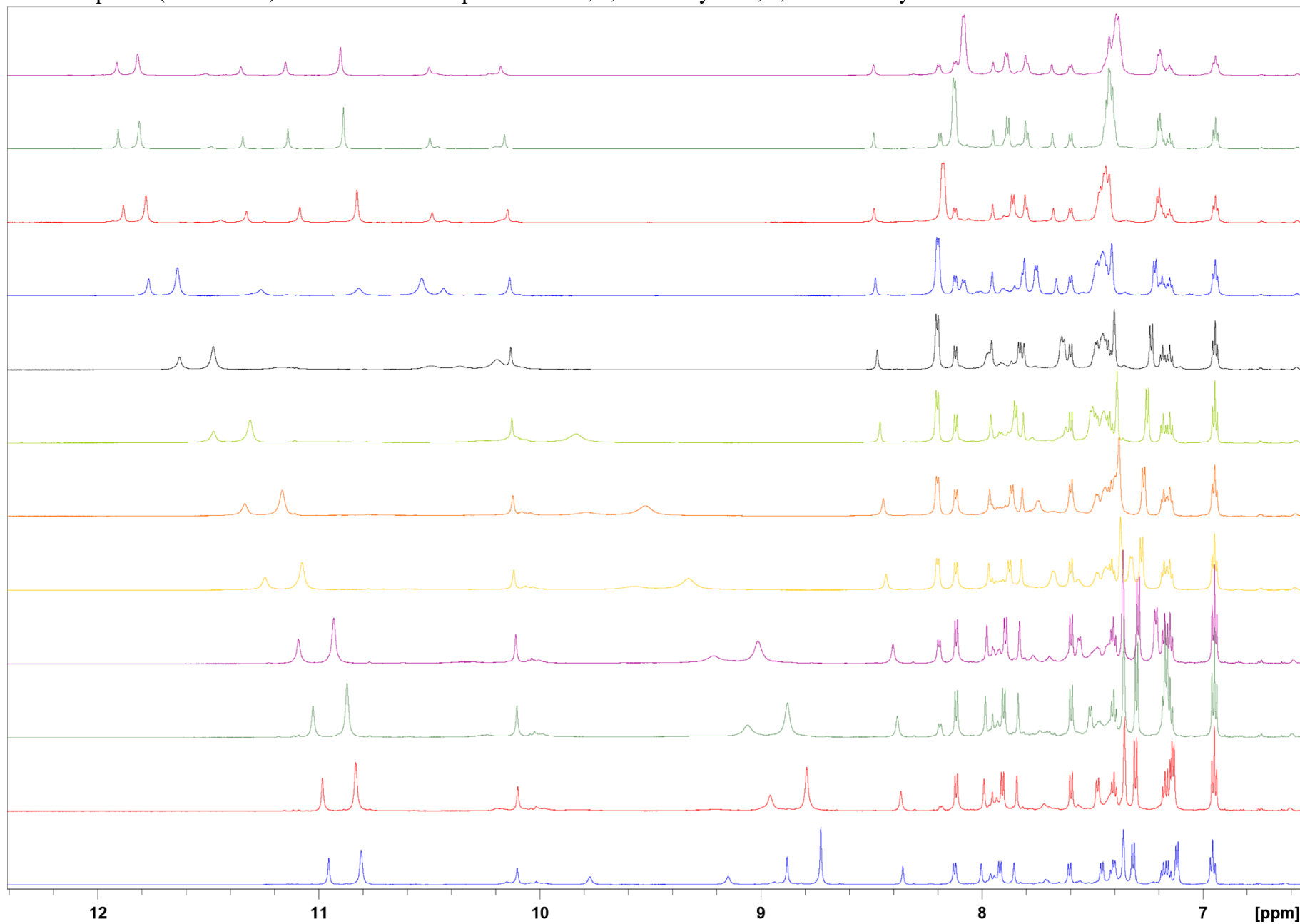
Stacked ^1H NMR spectra (700.1 MHz) of a mixture of receptors **MC004**; 1,3-diindolylurea; 1,3-dicarbazolylurea + TBA-benzoate



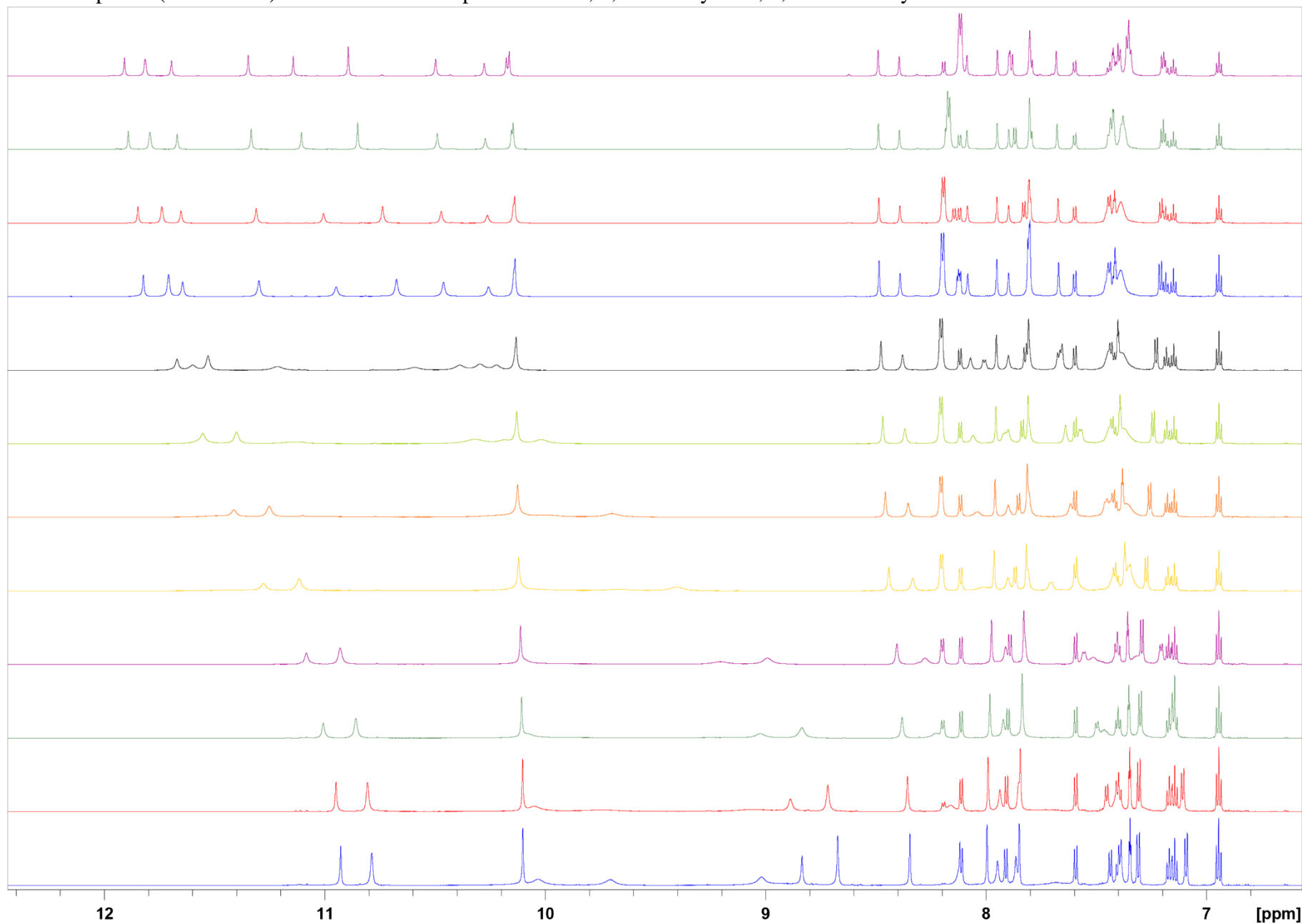
Stacked ^1H NMR spectra (700.1 MHz) of a mixture of receptors **MC005**; 1,3-diindolylurea; 1,3-dicarbazolylurea + TBA-benzoate



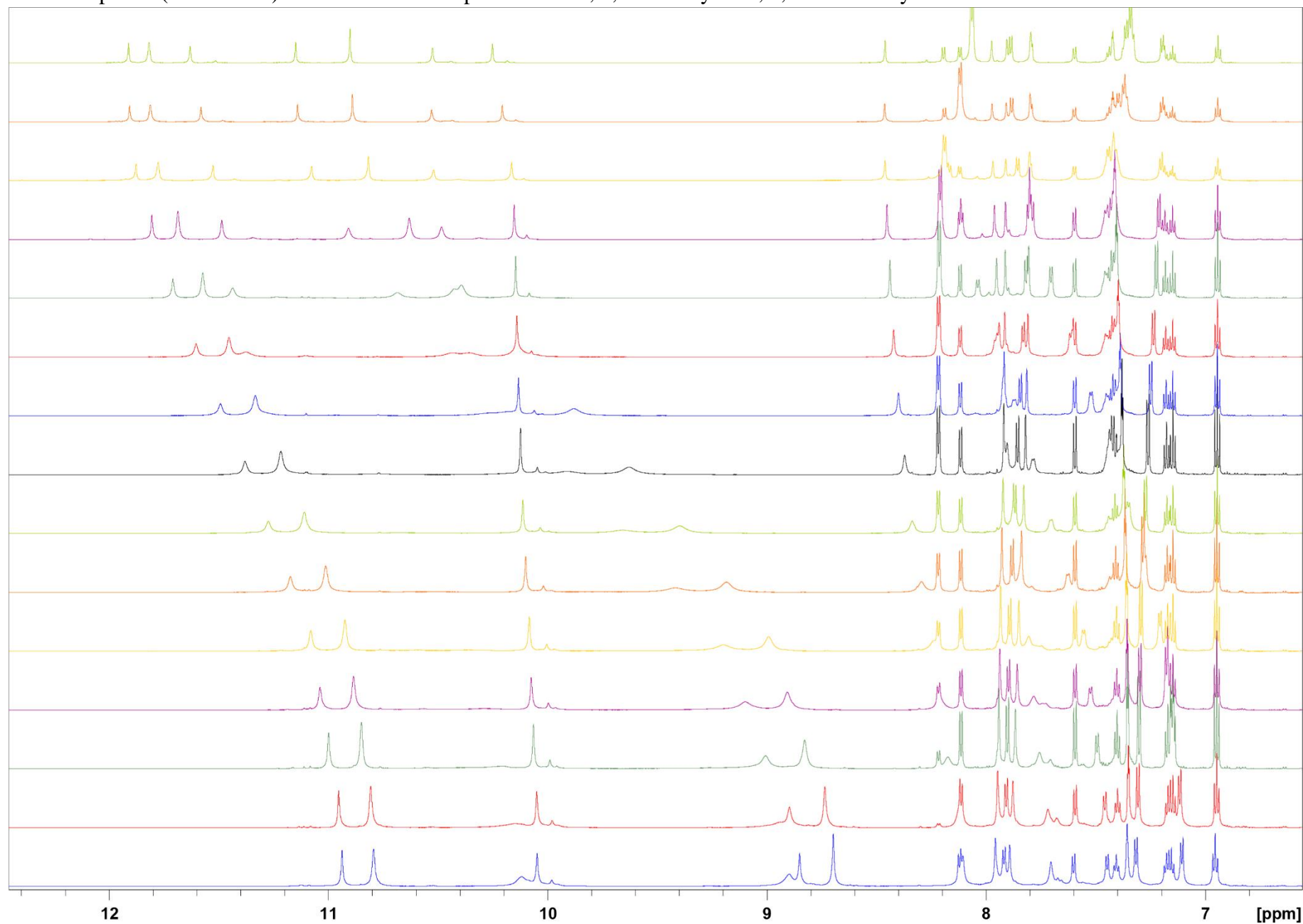
Stacked ^1H NMR spectra (700.1 MHz) of a mixture of receptors **MC006**; 1,3-diindolylurea; 1,3-dicarbazolylurea + TBA-benzoate



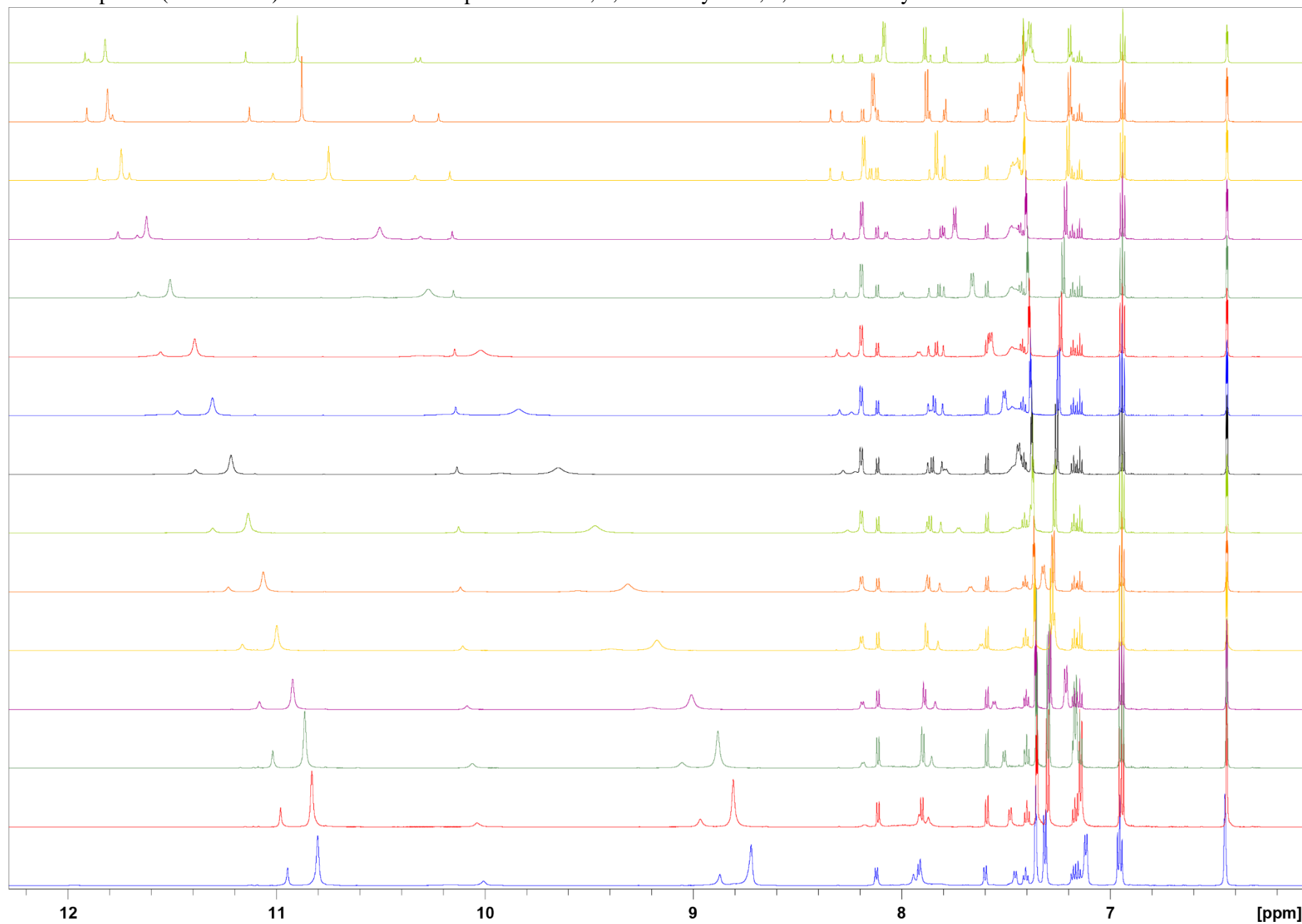
Stacked ^1H NMR spectra (700.1 MHz) of a mixture of receptors **MC007**; 1,3-diindolylurea; 1,3-dicarbazolylurea + TBA-benzoate



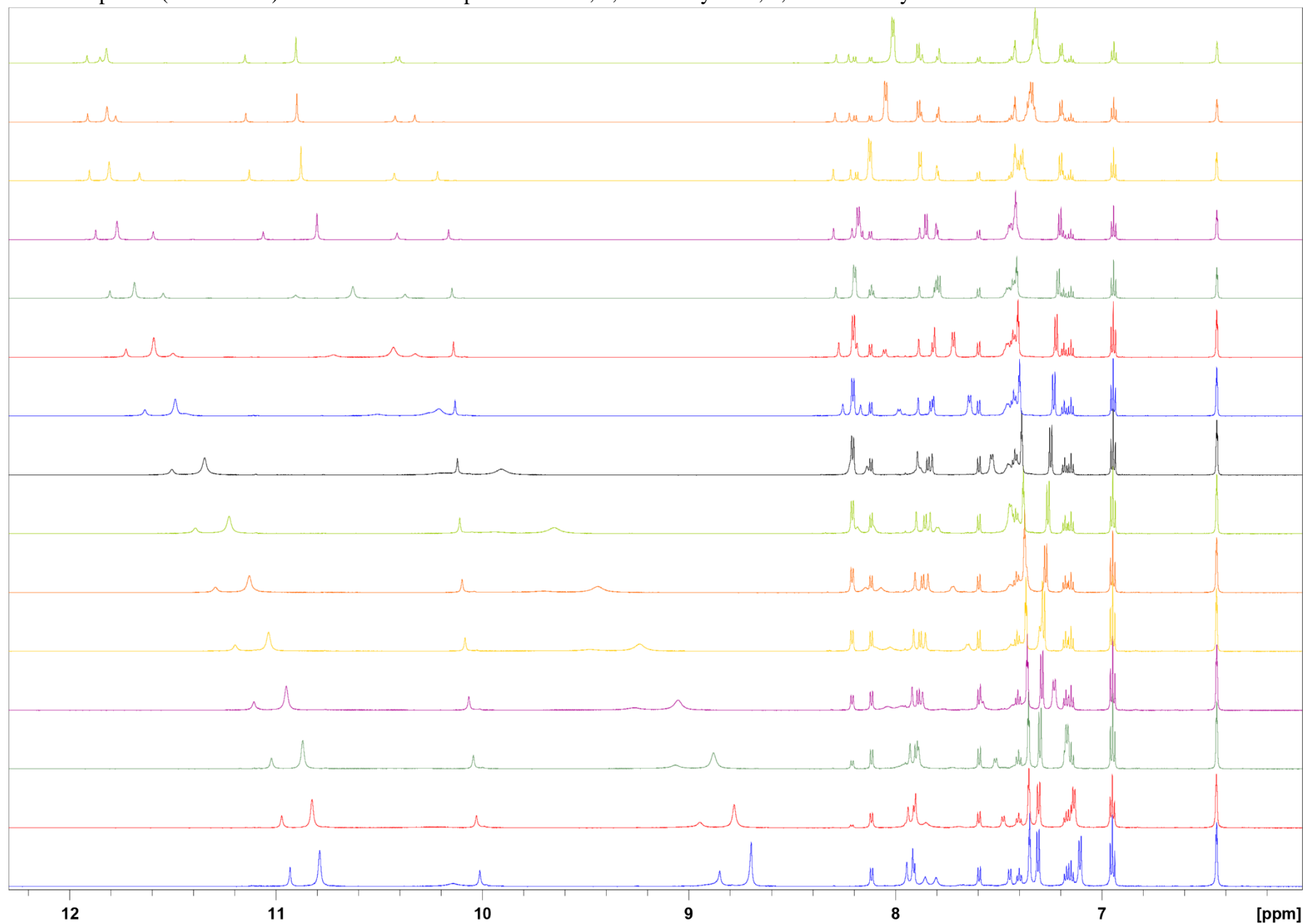
Stacked ^1H NMR spectra (700.1 MHz) of a mixture of receptors **MC008**; 1,3-diindolylurea; 1,3-dicarbazolylurea + TBA-benzoate



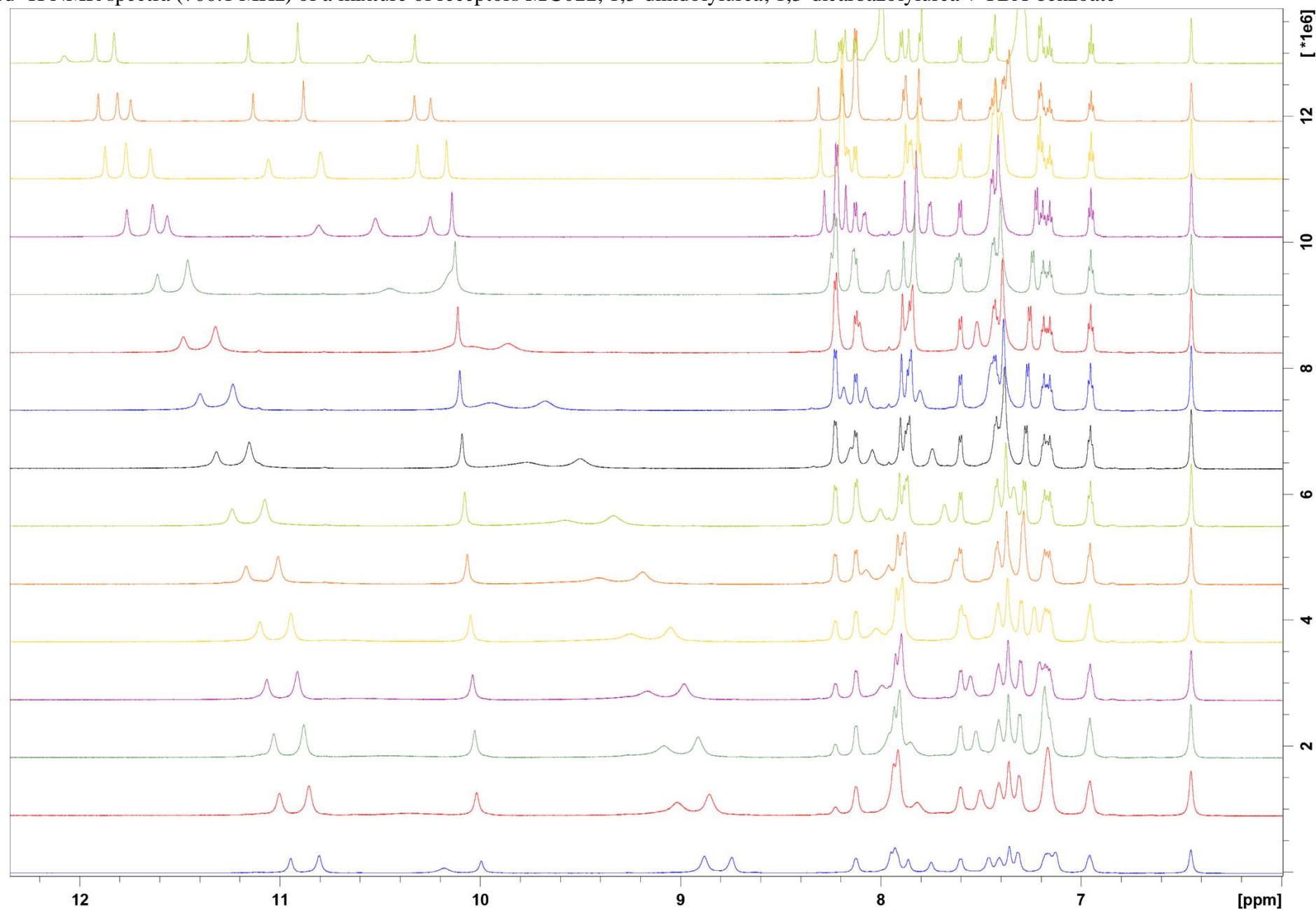
Stacked ^1H NMR spectra (700.1 MHz) of a mixture of receptors **MC009**; 1,3-diindolylurea; 1,3-dicarbazolylurea + TBA-benzoate



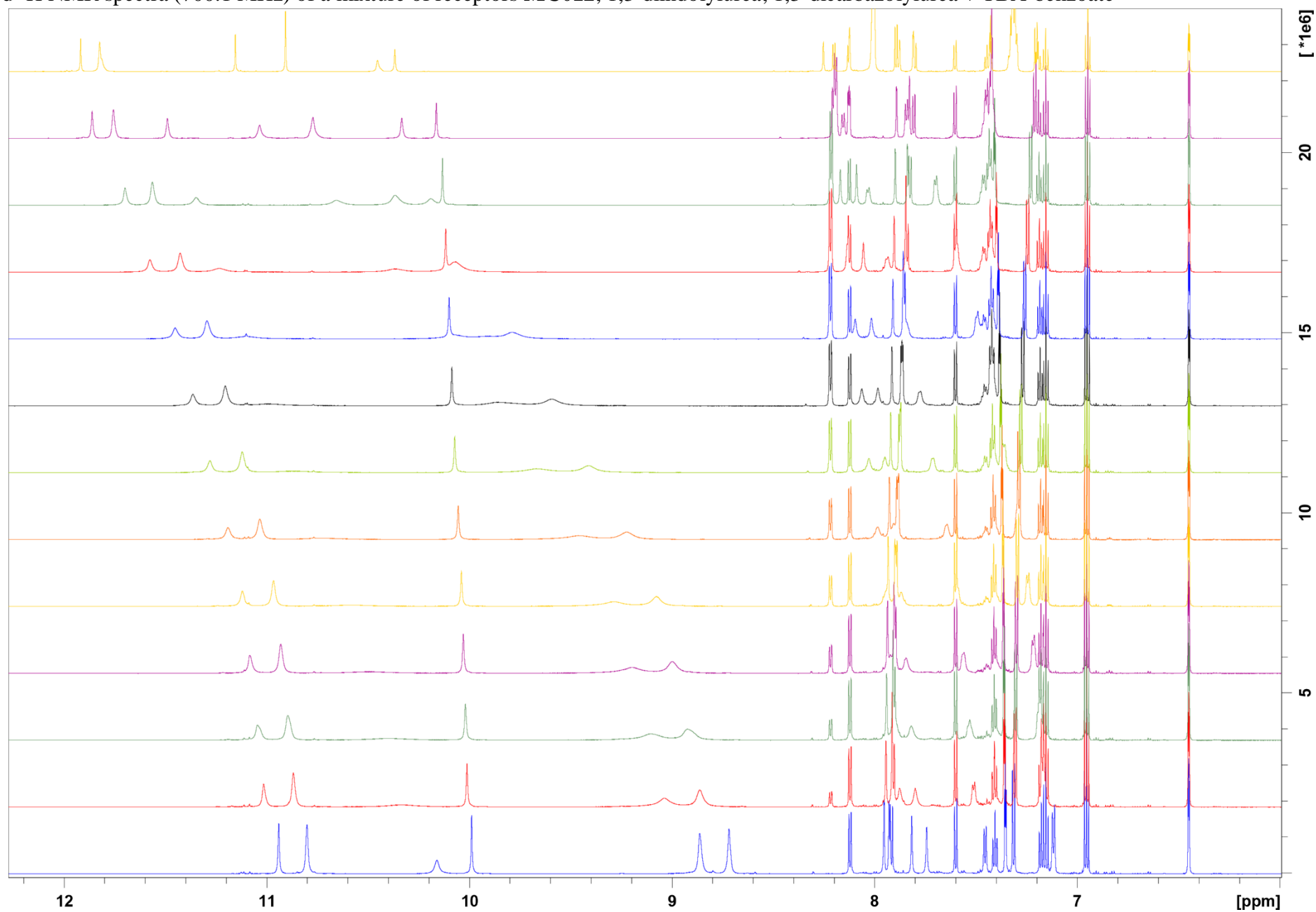
Stacked ^1H NMR spectra (700.1 MHz) of a mixture of receptors **MC010**; 1,3-diindolylurea; 1,3-dicarbazolylurea + TBA-benzoate



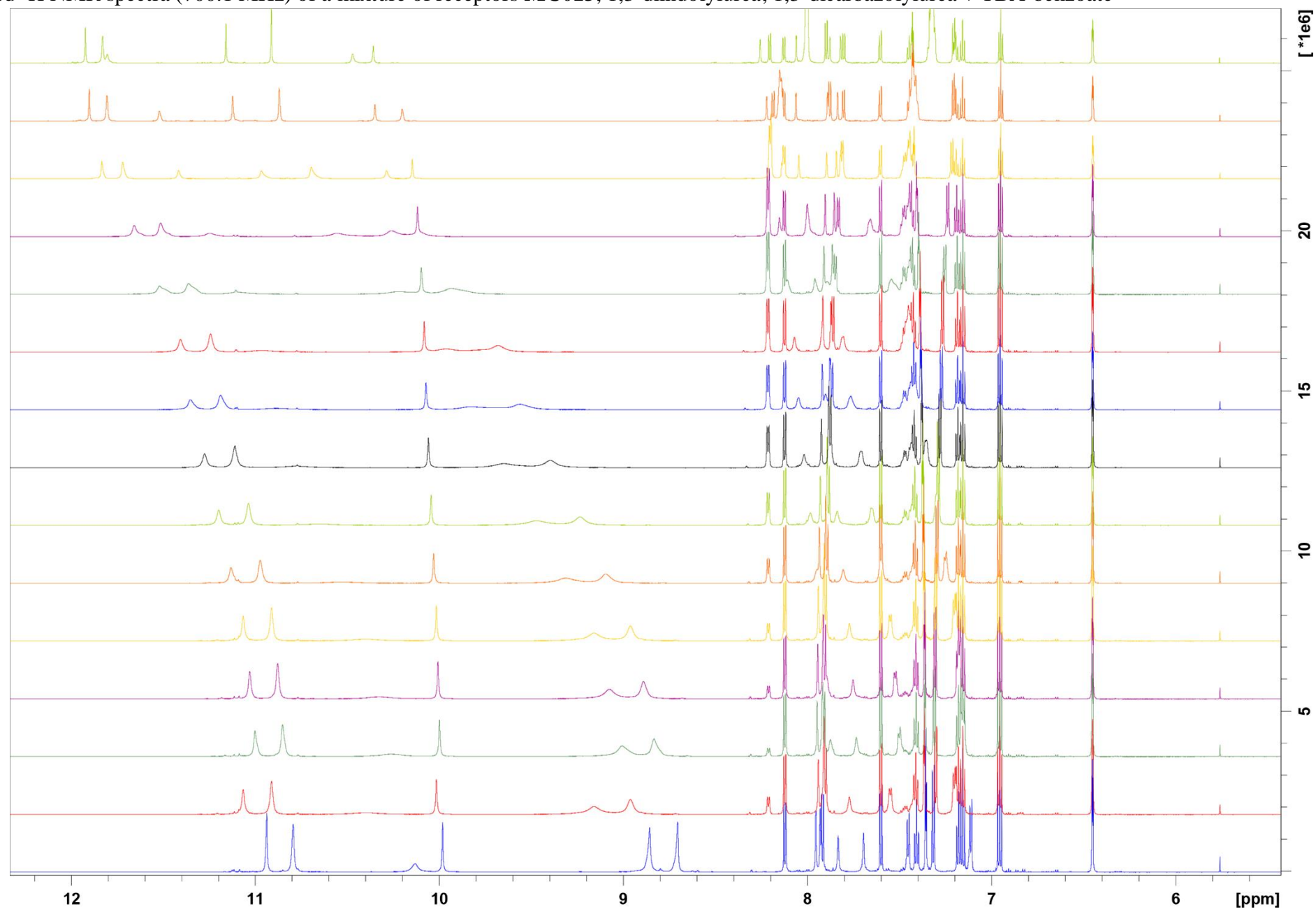
Stacked ^1H NMR spectra (700.1 MHz) of a mixture of receptors **MC011**; 1,3-diindolylurea; 1,3-dicarbazolylurea + TBA-benzoate



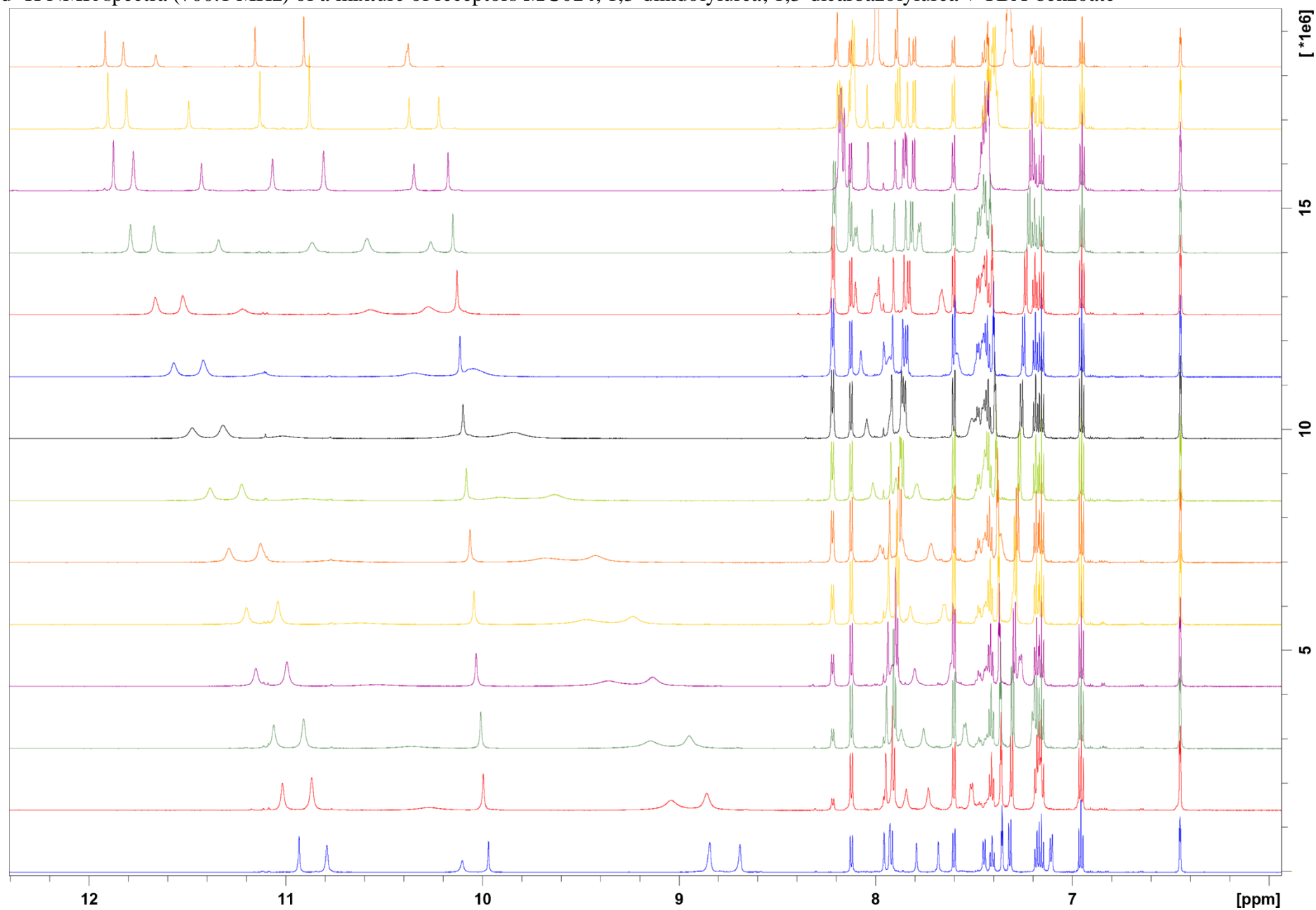
Stacked ^1H NMR spectra (700.1 MHz) of a mixture of receptors **MC012**; 1,3-diindolylurea; 1,3-dicarbazolylurea + TBA-benzoate



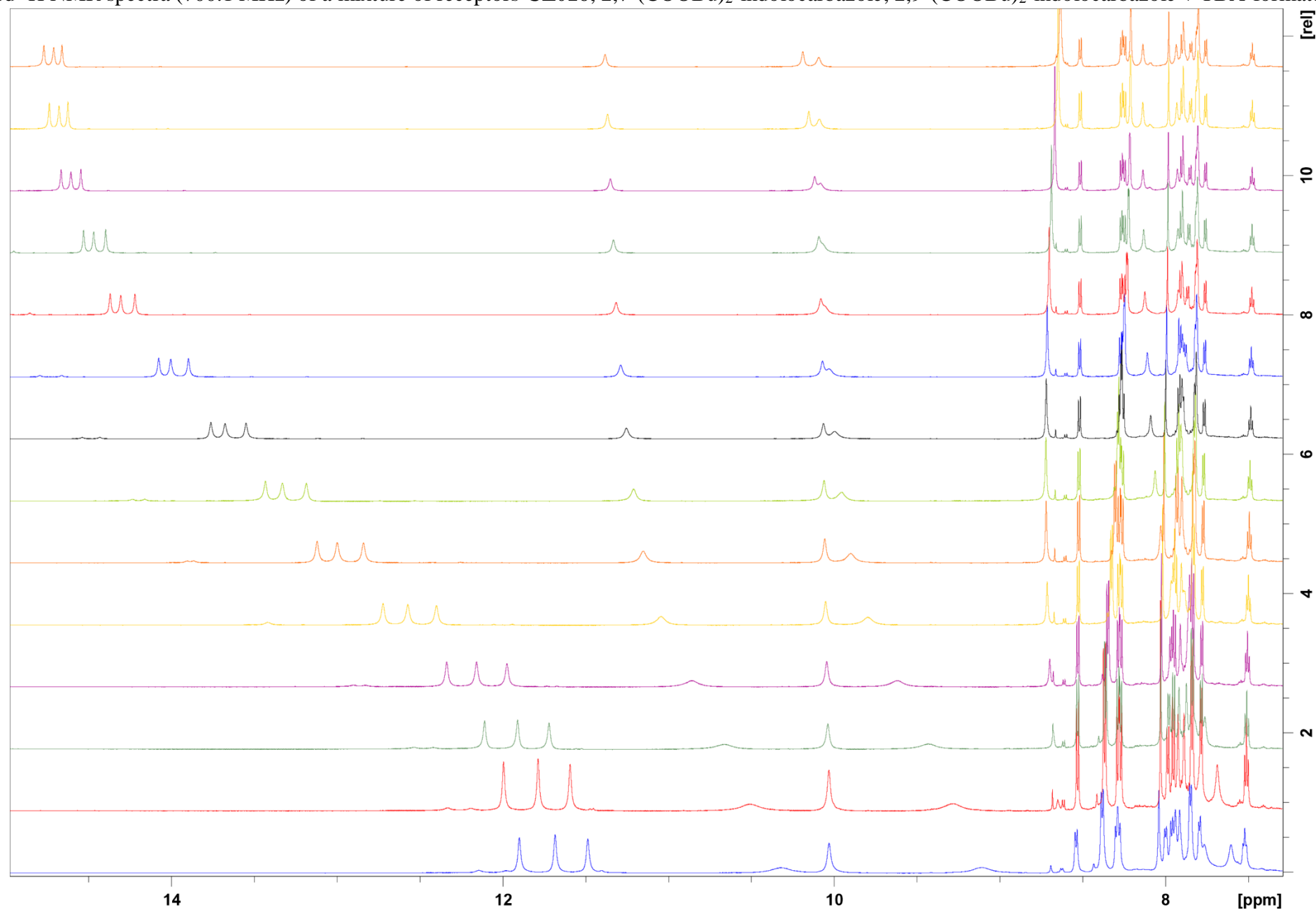
Stacked ^1H NMR spectra (700.1 MHz) of a mixture of receptors **MC013**; 1,3-diindolylurea; 1,3-dicarbazolylurea + TBA-benzoate



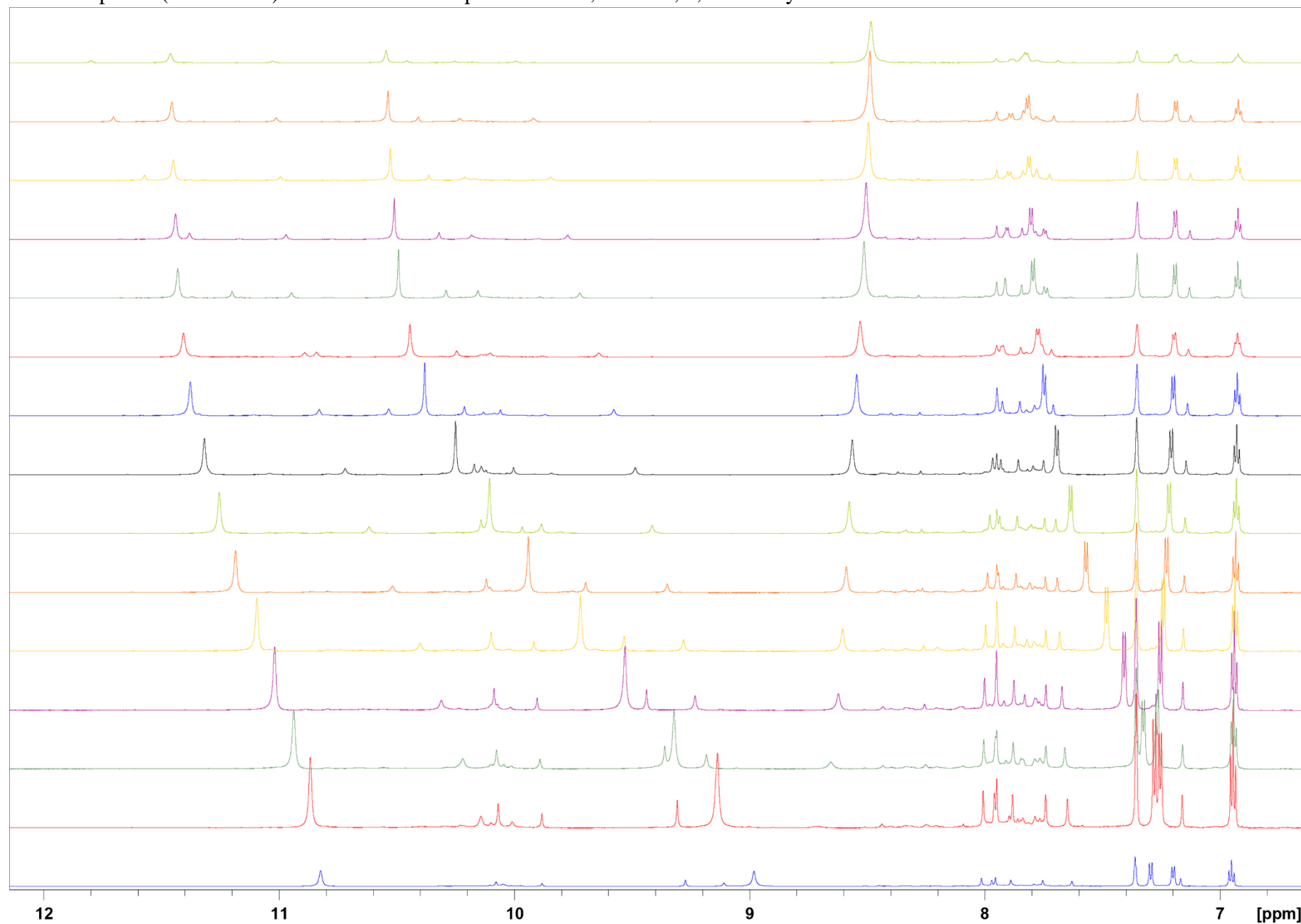
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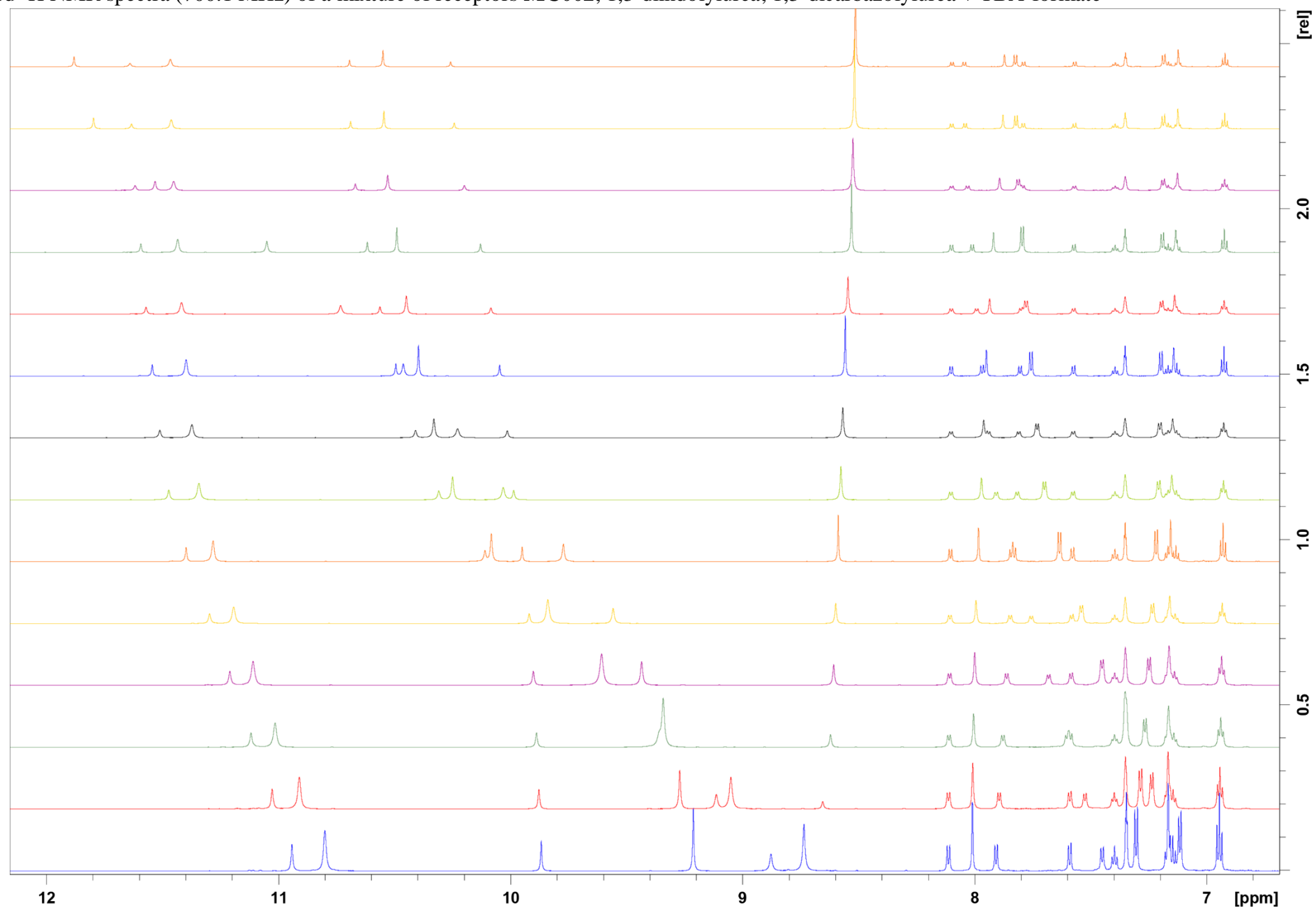
Stacked ^1H NMR spectra (700.1 MHz) of a mixture of receptors **CZ016**; 2,7-(COOBu) $_2$ -indolocarbazole; 2,9-(COOBu) $_2$ -indolocarbazole + TBA-formate



Stacked ^1H NMR spectra (700.1 MHz) of a mixture of receptors **MC001**; **MC004**; 1,3-diindolylurea + TBA-formate



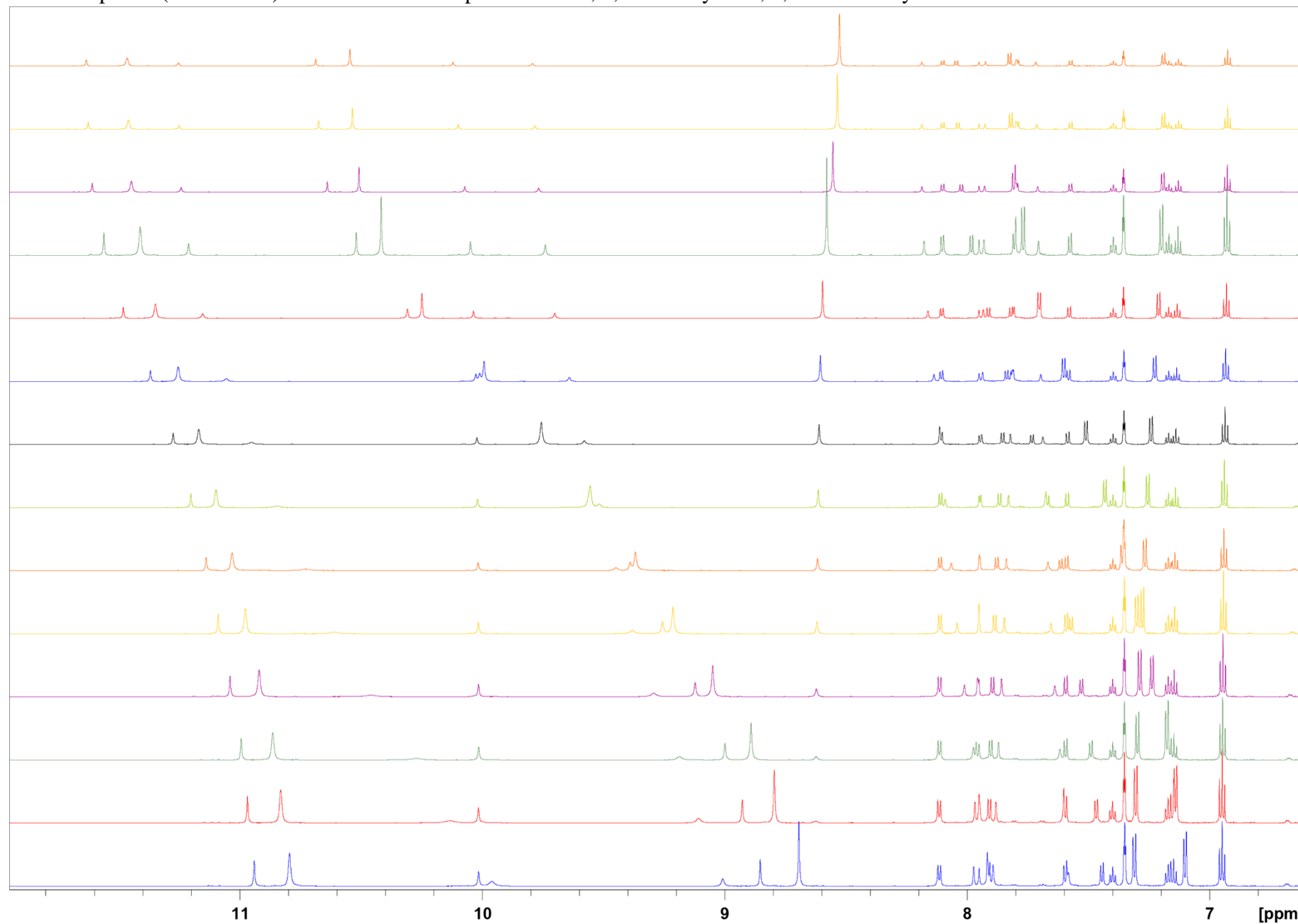
Stacked ^1H NMR spectra (700.1 MHz) of a mixture of receptors **MC002**; 1,3-diindolylurea; 1,3-dicarbazolylurea + TBA-formate



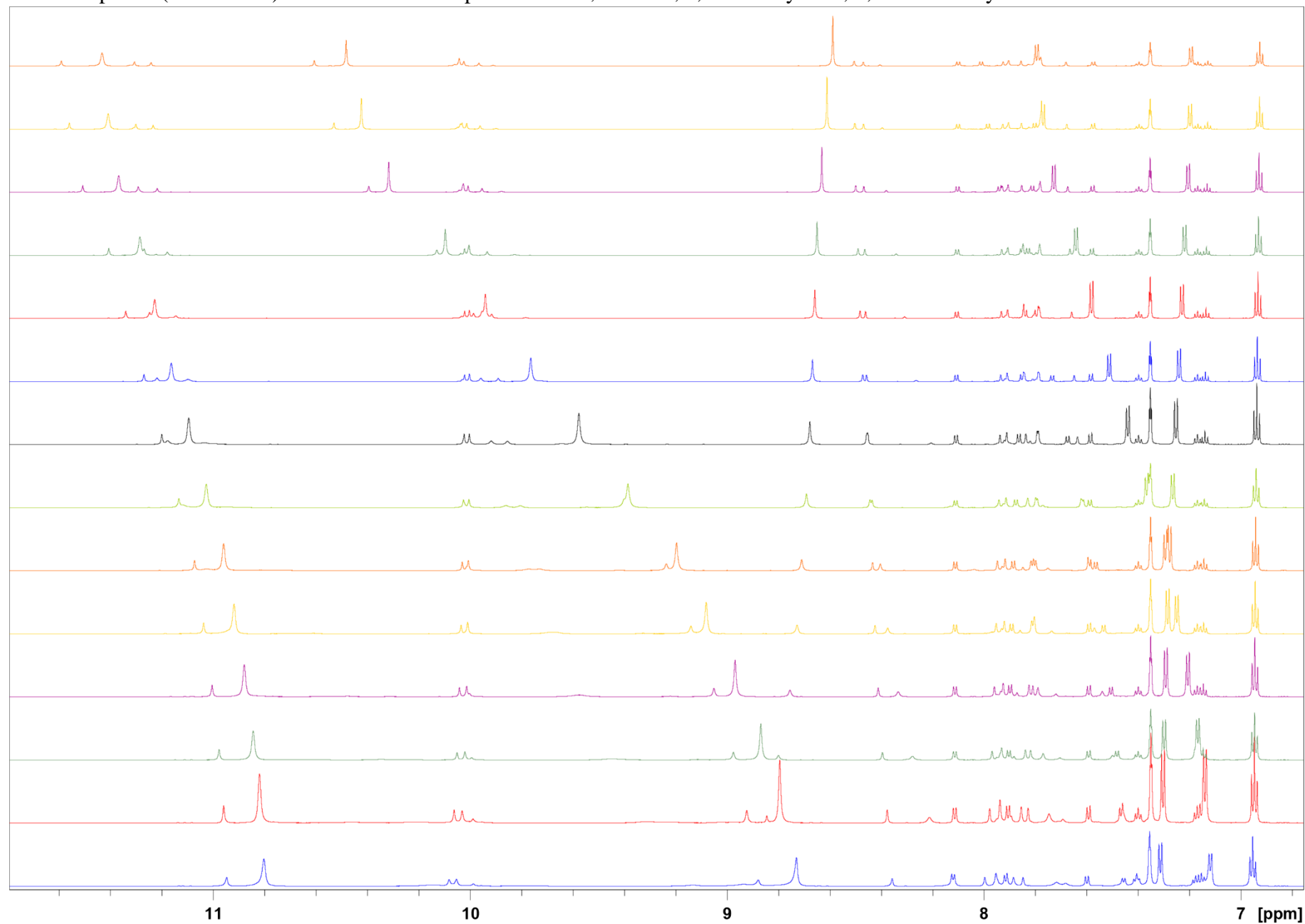
Stacked ^1H NMR spectra (700.1 MHz) of a mixture of receptors **MC003**; 1,3-diindolylurea; 1,3-dicarbazolylurea + TBA-formate



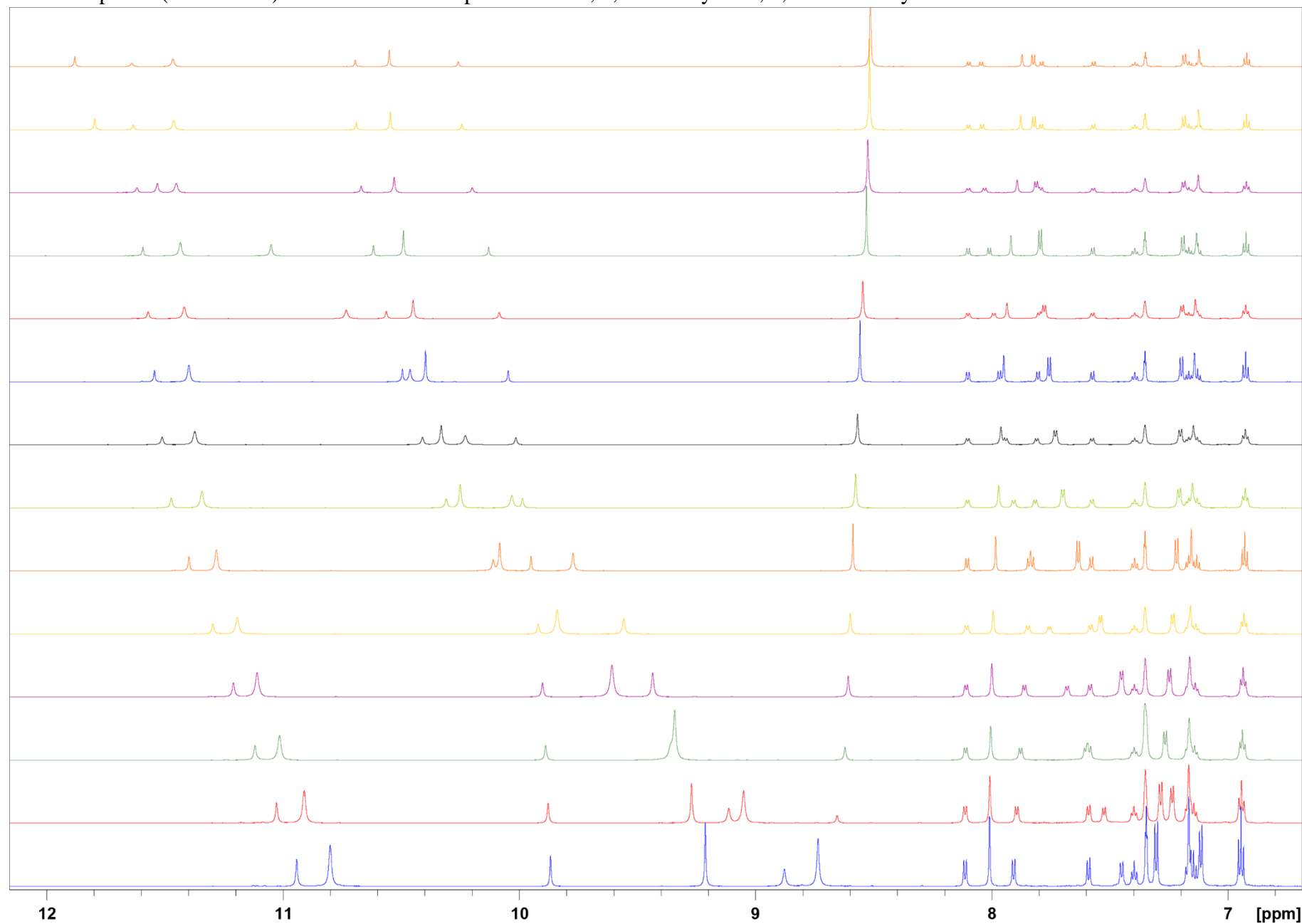
Stacked ^1H NMR spectra (700.1 MHz) of a mixture of receptors **MC005**; 1,3-diindolylurea; 1,3-dicarbazolylurea + TBA-formate



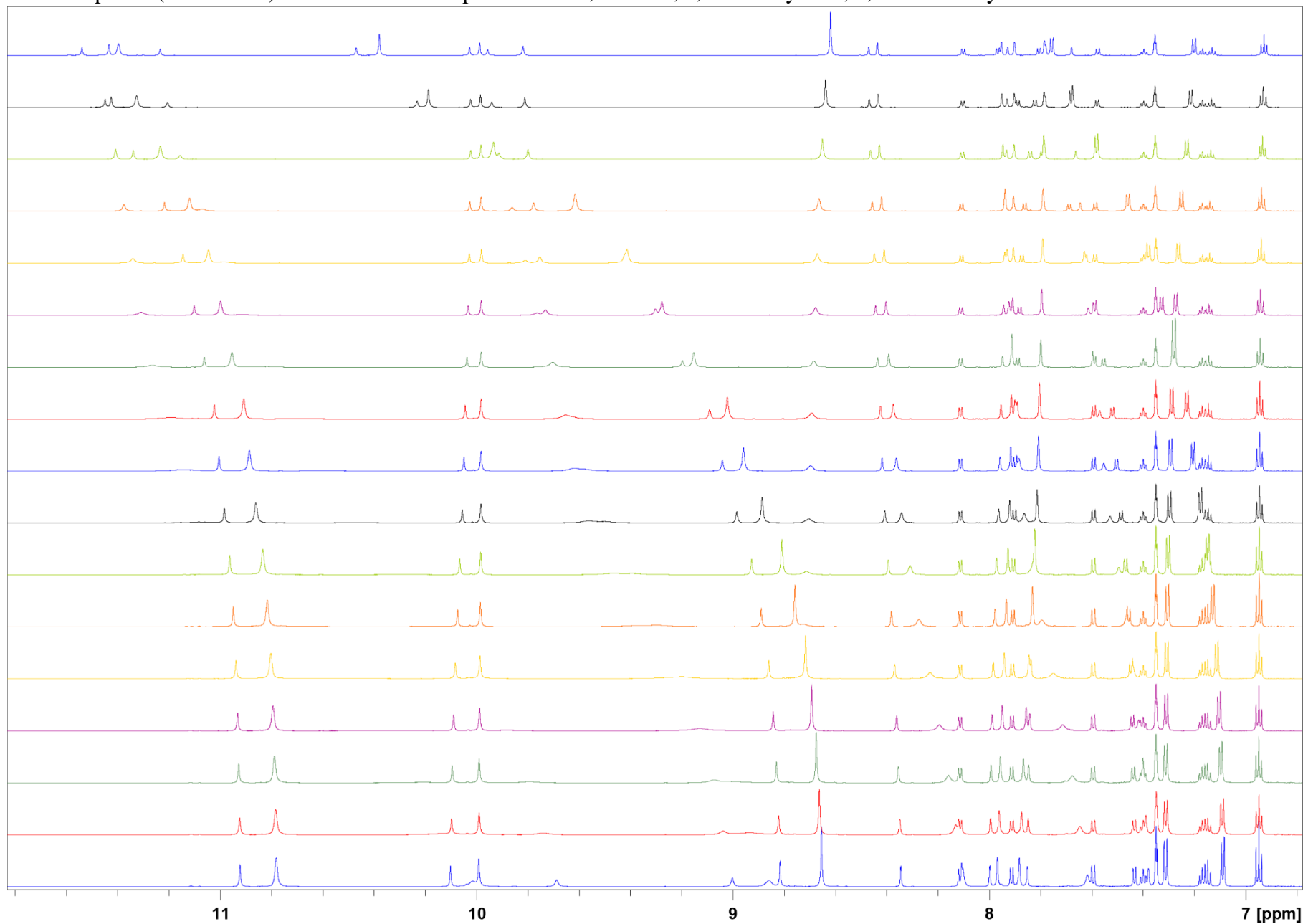
Stacked ^1H NMR spectra (700.1 MHz) of a mixture of receptors **MC006**; **MC008**; 1,3-diindolylurea; 1,3-dicarbazolylurea + TBA-formate



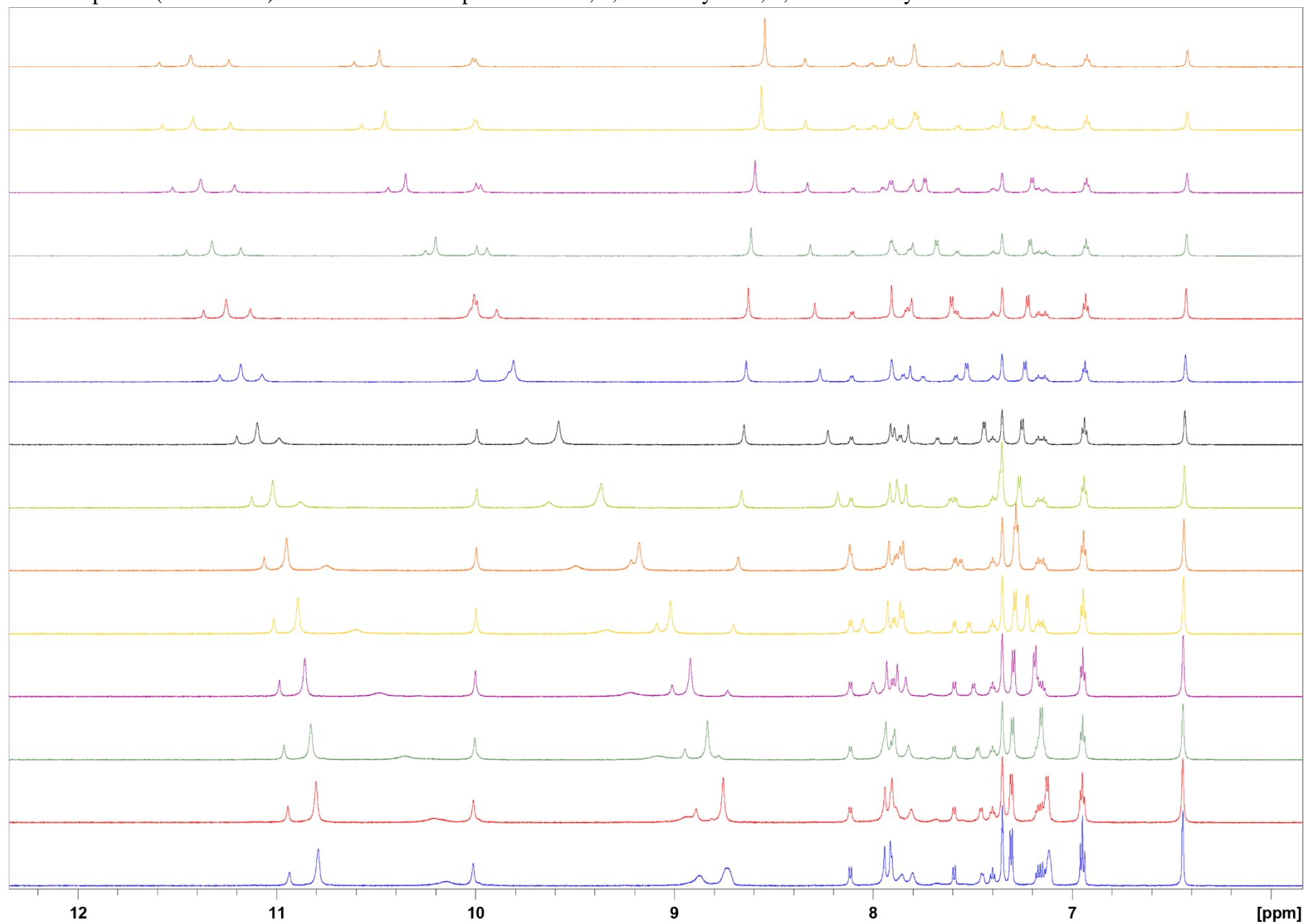
Stacked ^1H NMR spectra (700.1 MHz) of a mixture of receptors **MC007**; 1,3-diindolylurea; 1,3-dicarbazolylurea + TBA-formate



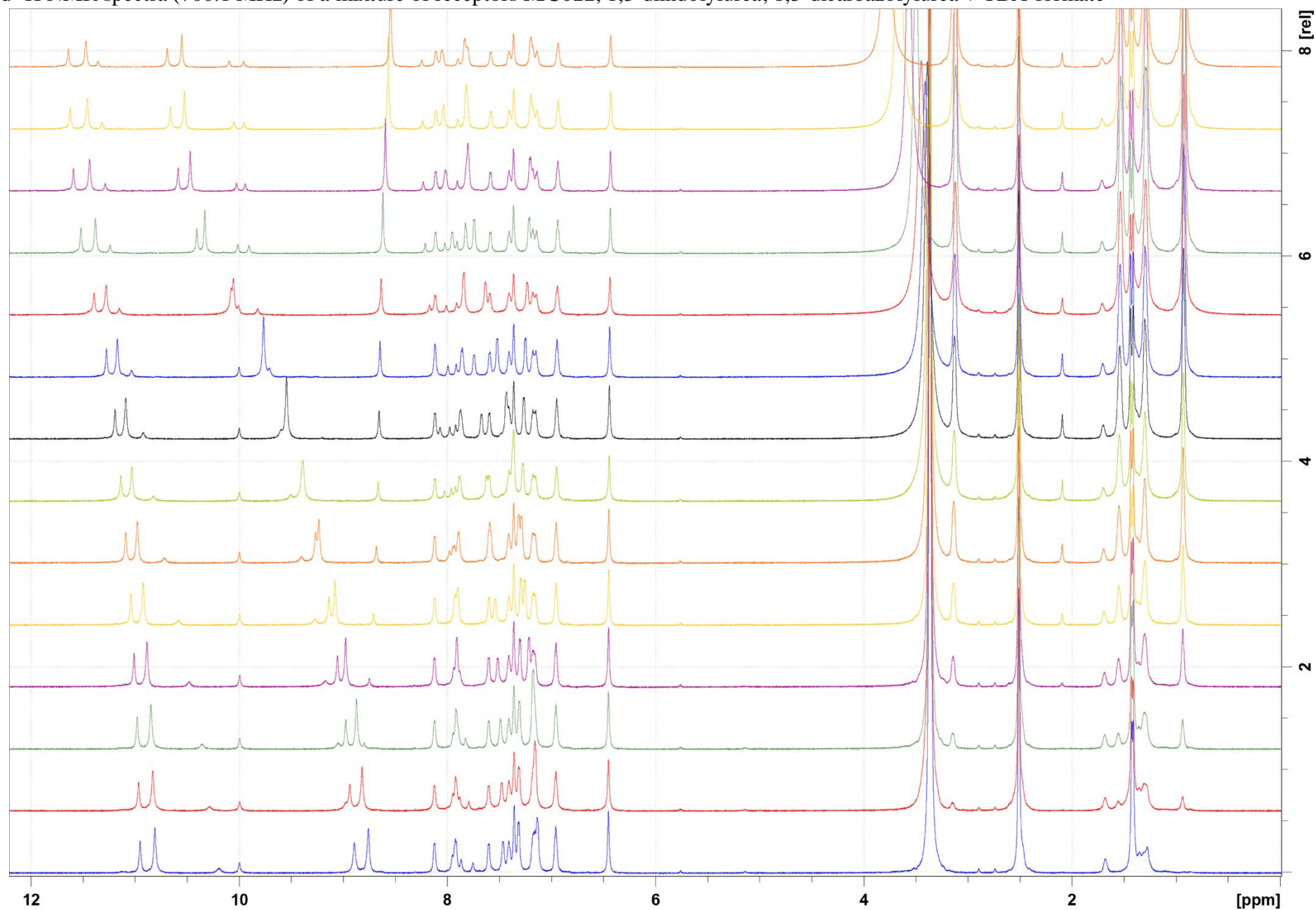
Stacked ^1H NMR spectra (700.1 MHz) of a mixture of receptors **MC008**; **MC009**; 1,3-diindolylurea; 1,3-dicarbazolylurea + TBA-formate



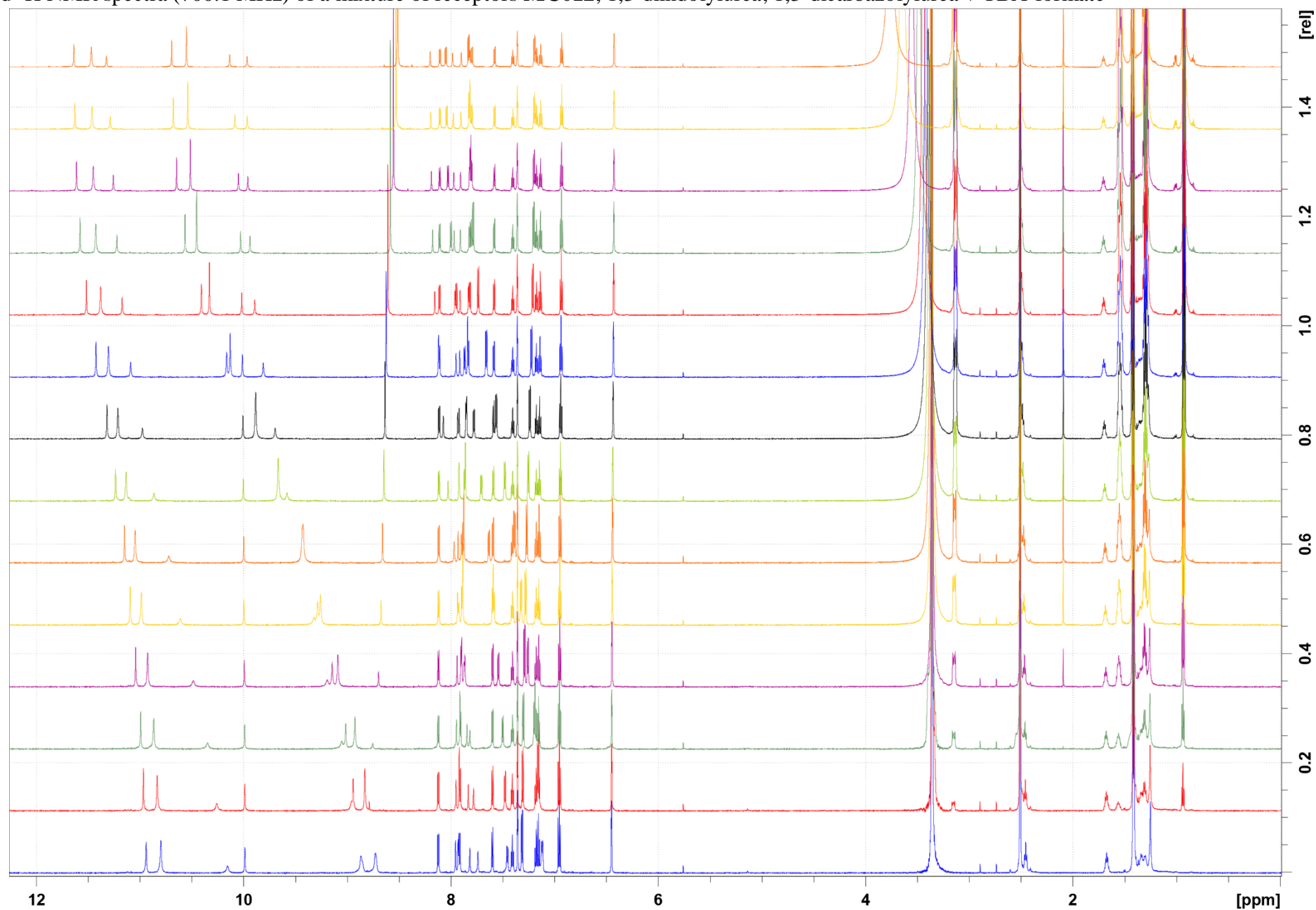
Stacked ^1H NMR spectra (700.1 MHz) of a mixture of receptors **MC010**; 1,3-diindolylurea; 1,3-dicarbazolylurea + TBA-formate



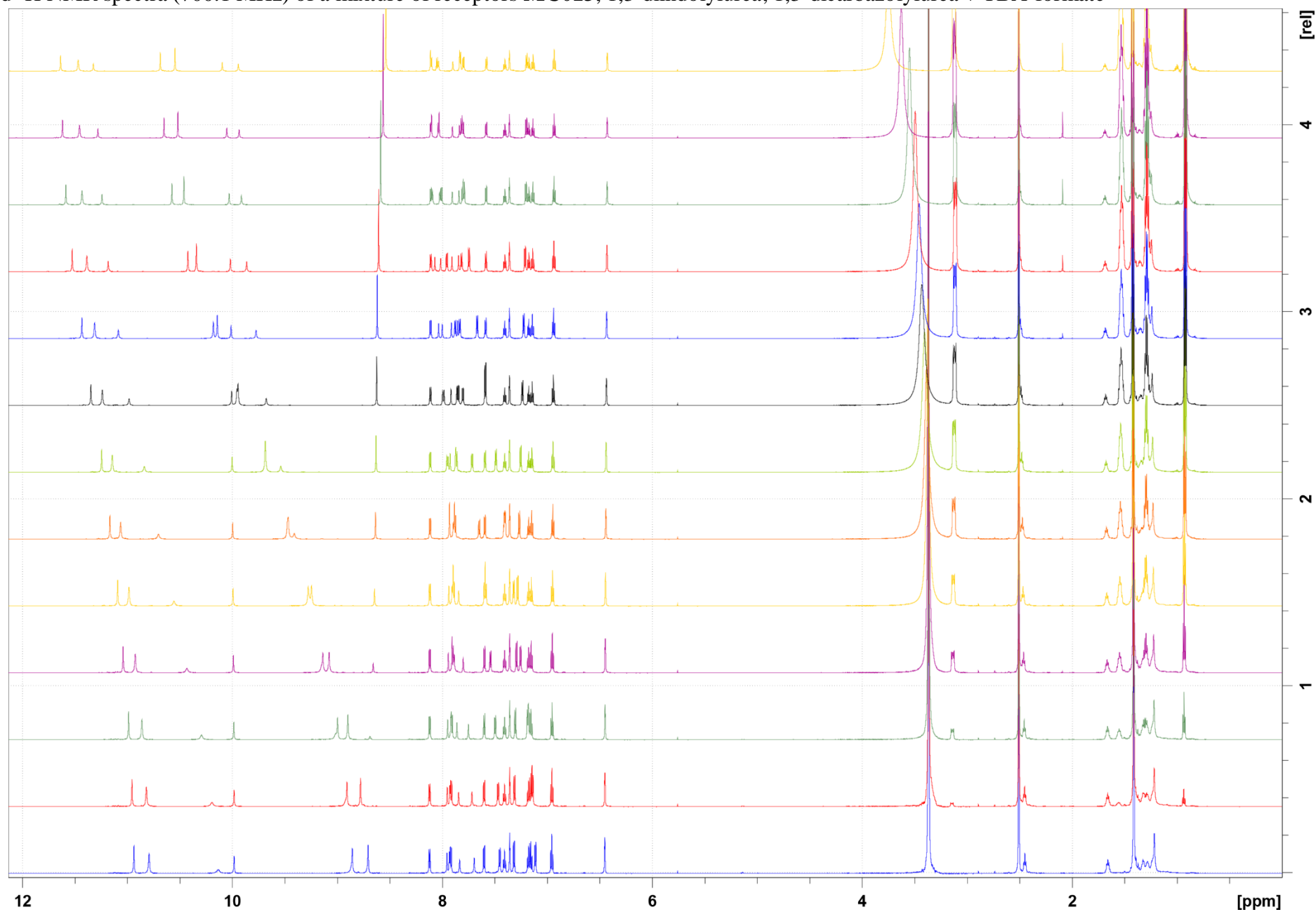
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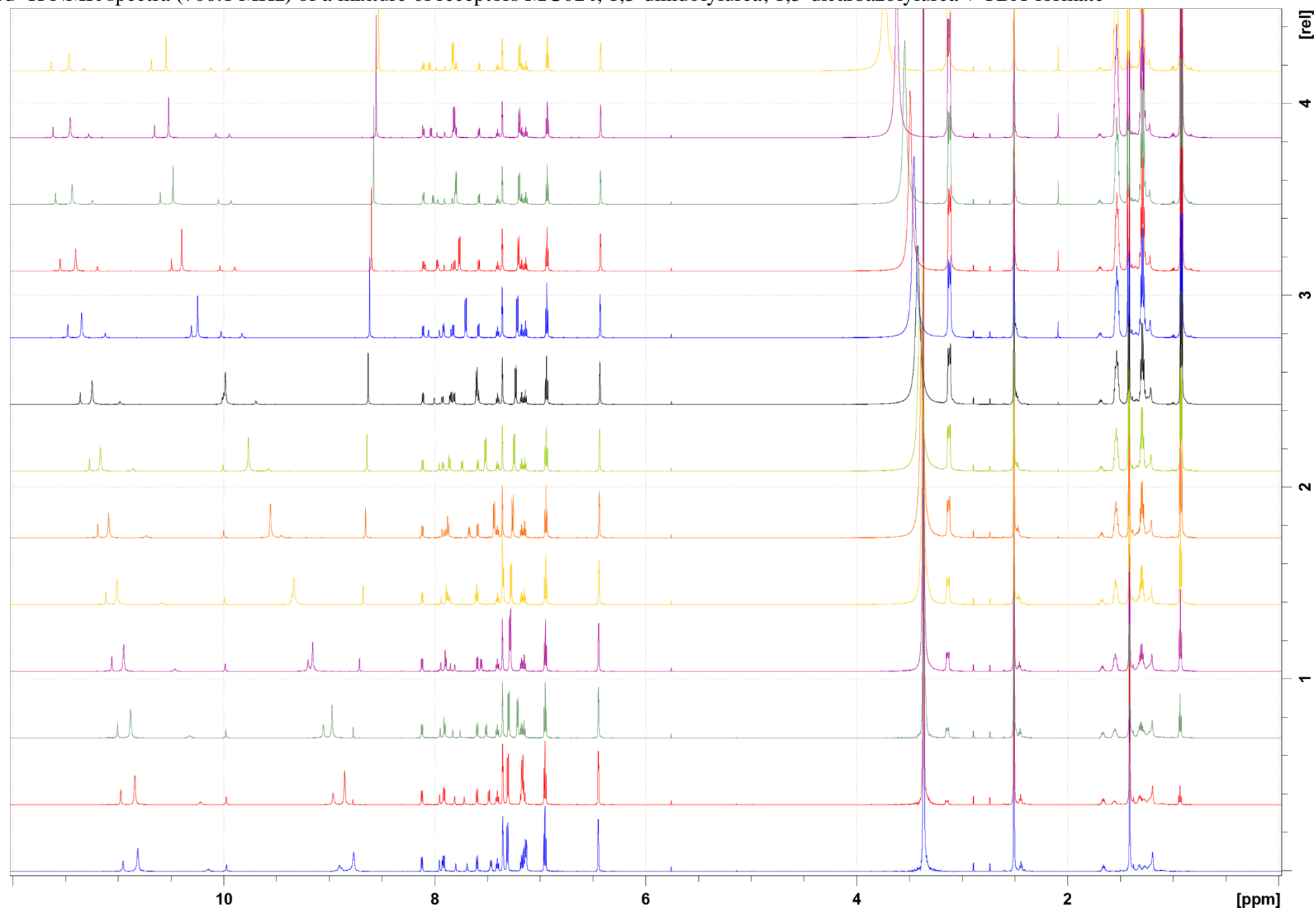
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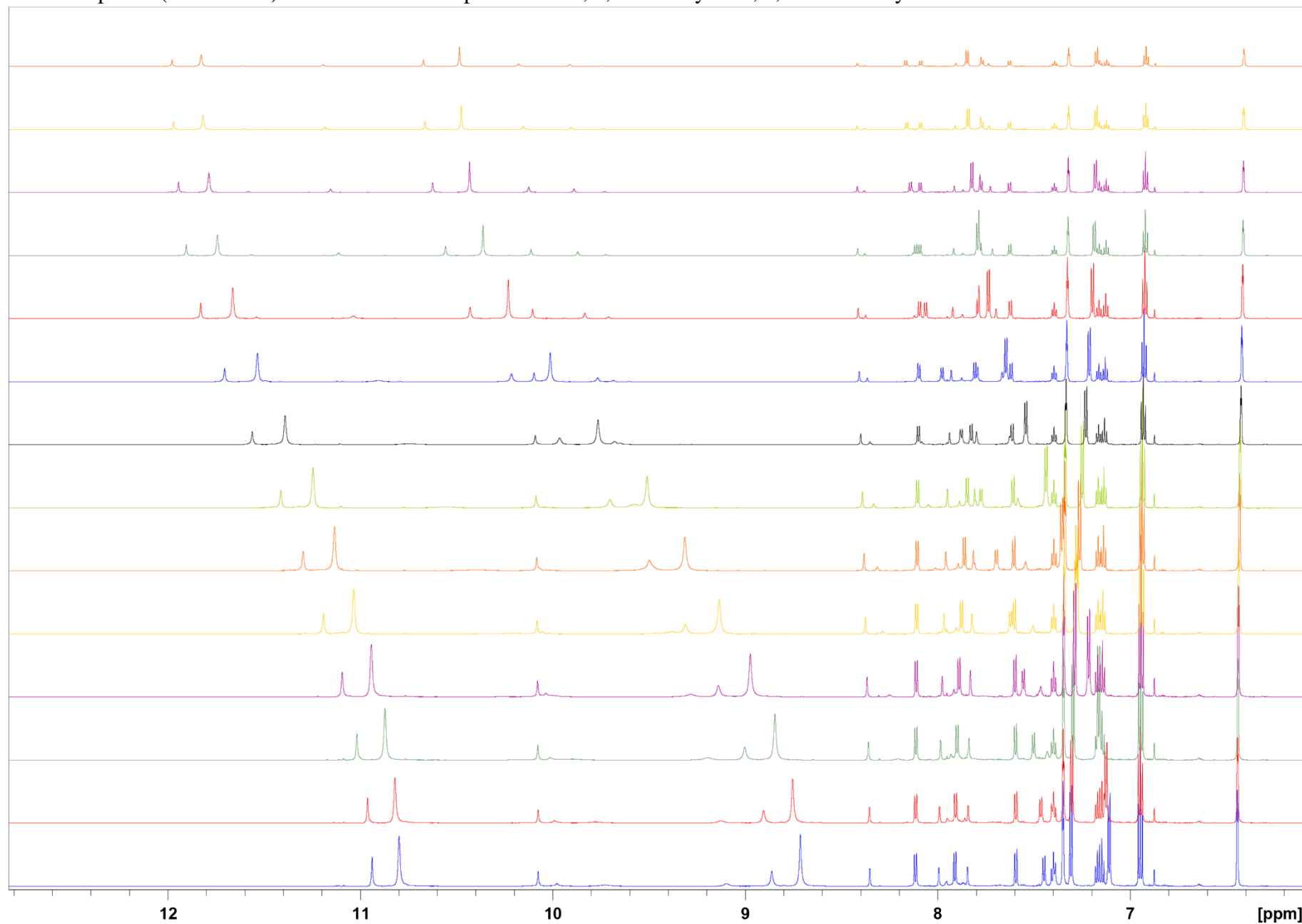
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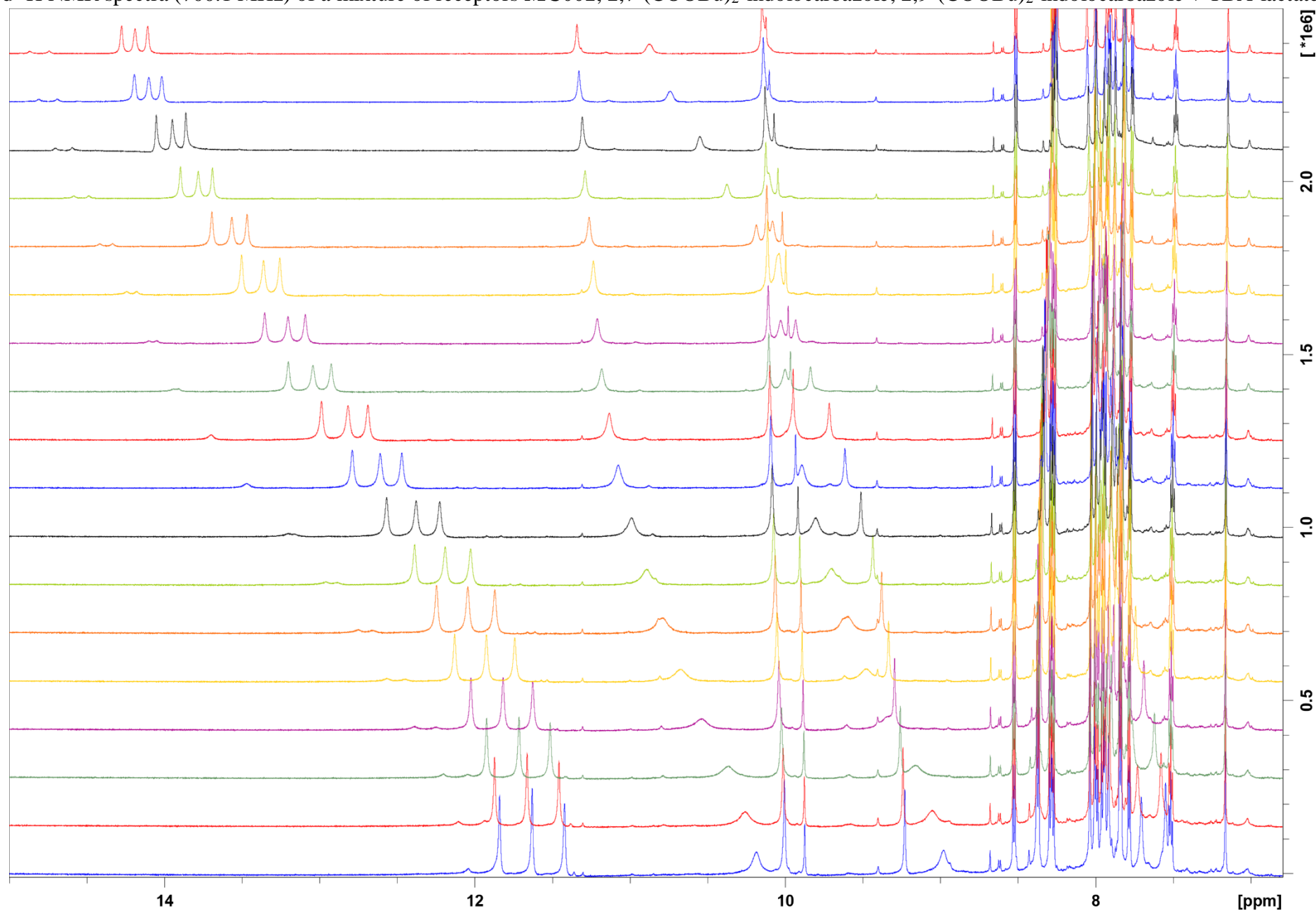
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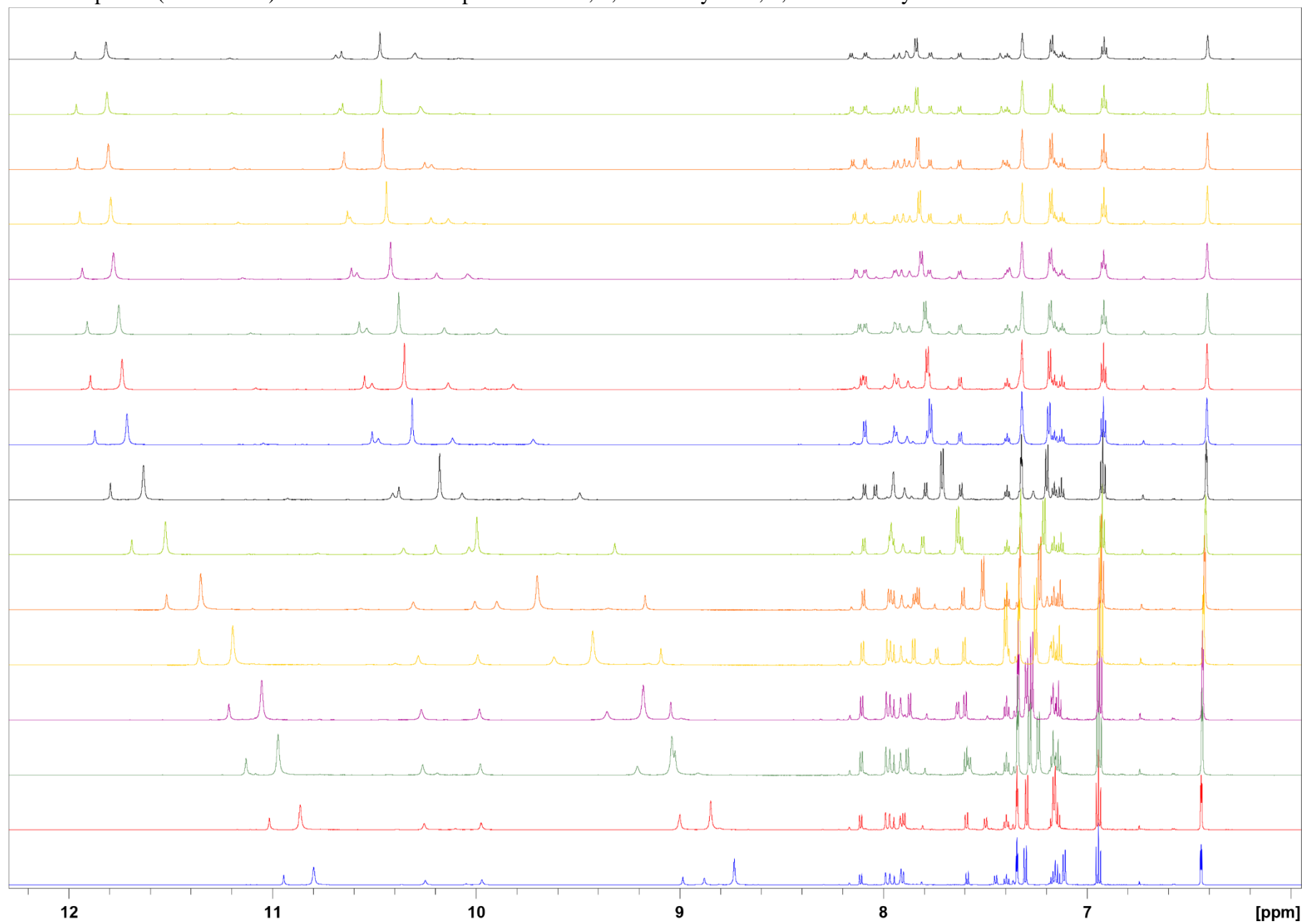
Stacked ^1H NMR spectra (700.1 MHz) of a mixture of receptors **CZ016**; 1,3-diindolylurea; 1,3-dicarbazolylurea + TBA-lactate



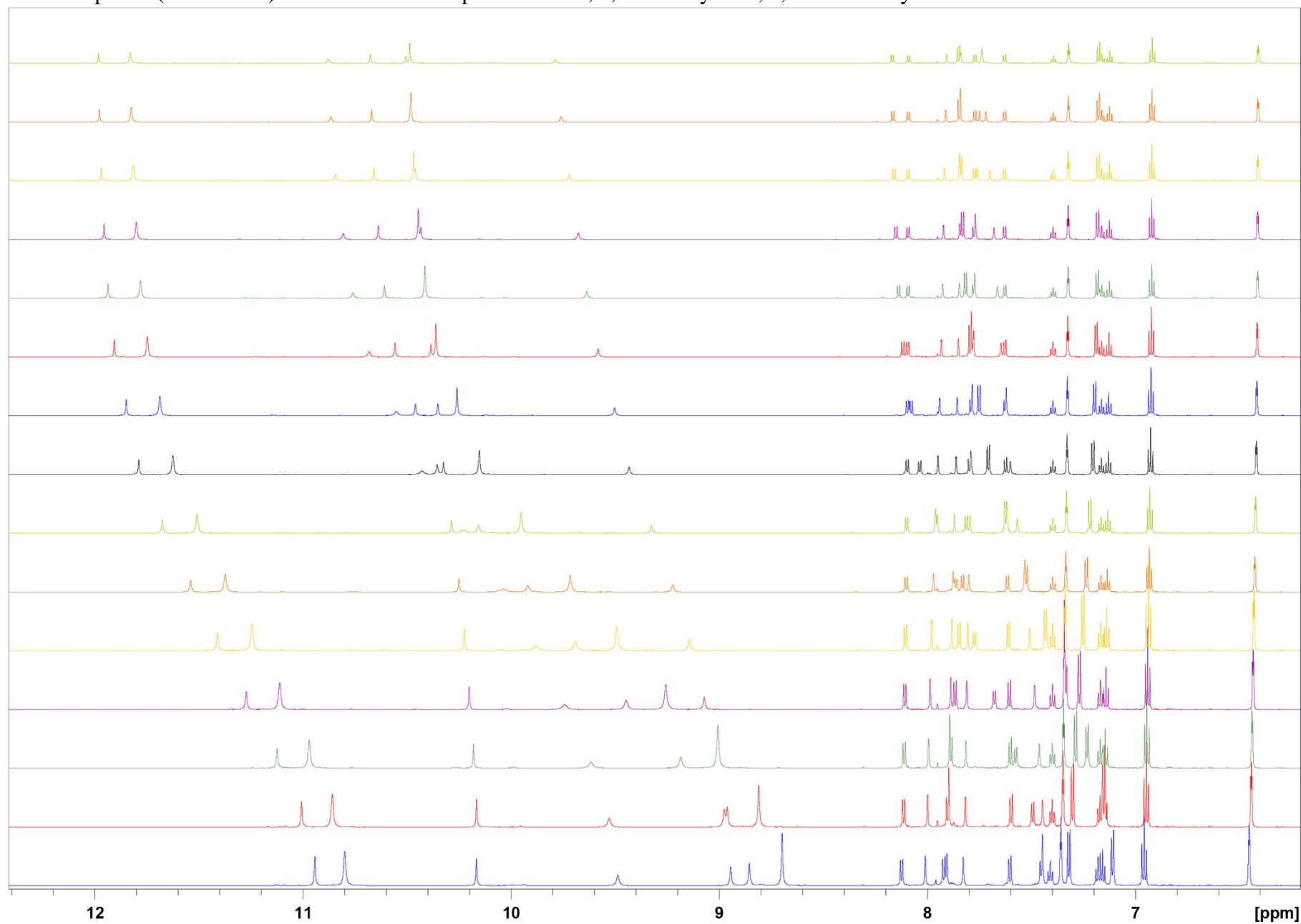
Stacked ^1H NMR spectra (700.1 MHz) of a mixture of receptors **MC001**; 2,7-(COOBu) $_2$ -indolocarbazole; 2,9-(COOBu) $_2$ -indolocarbazole + TBA-lactate



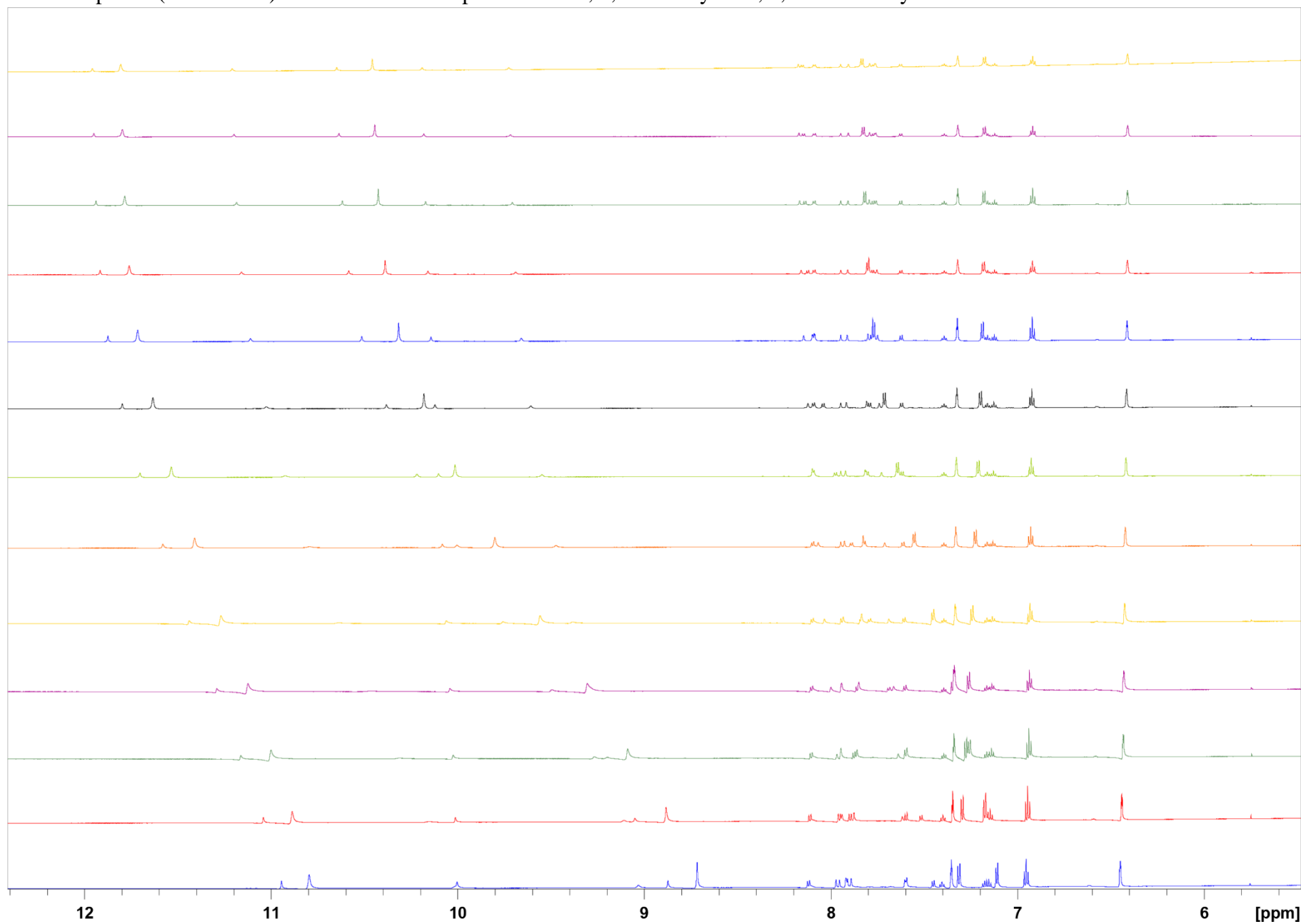
Stacked ^1H NMR spectra (700.1 MHz) of a mixture of receptors **MC003**; 1,3-diindolylurea; 1,3-dicarbazolylurea + TBA-lactate



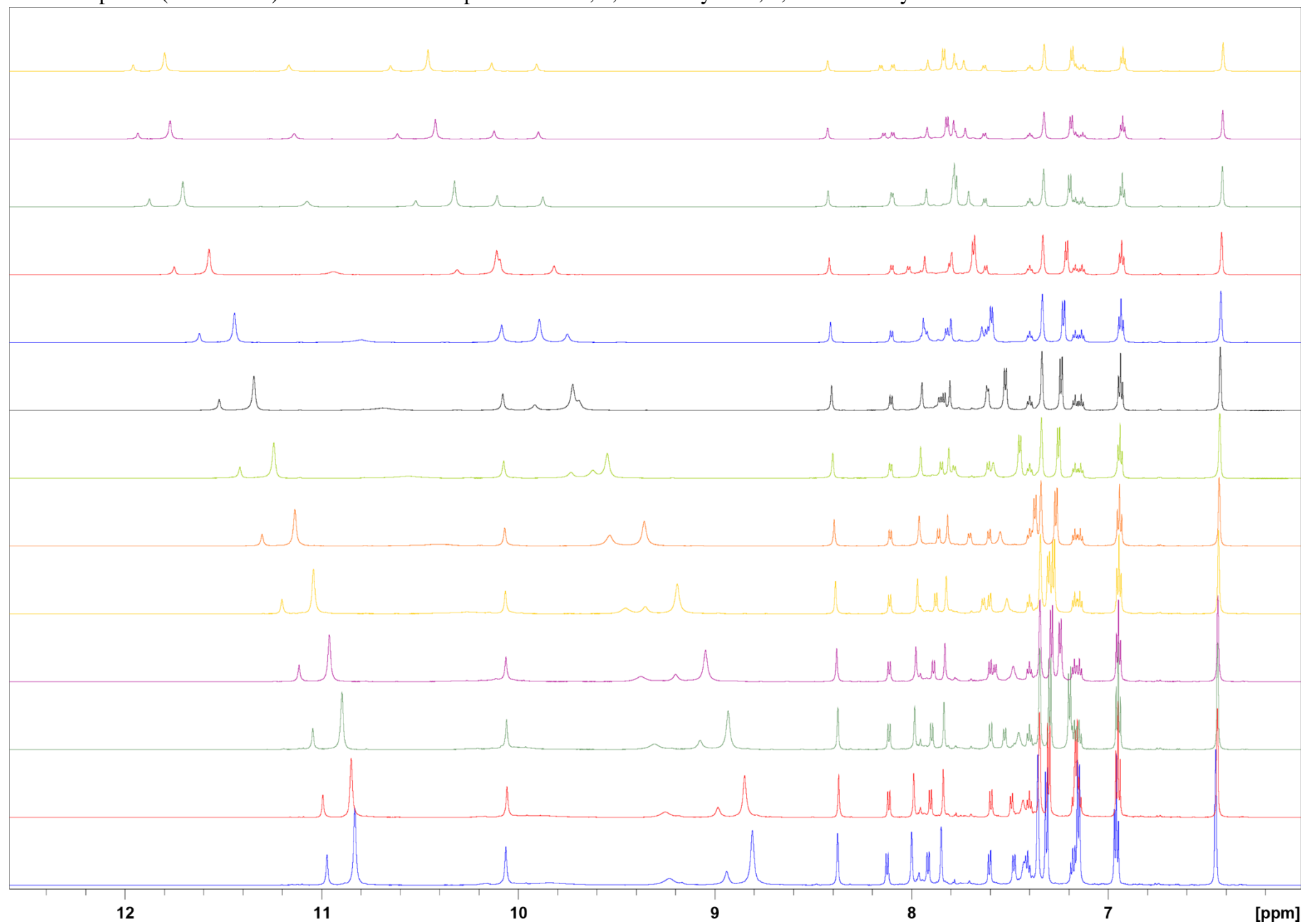
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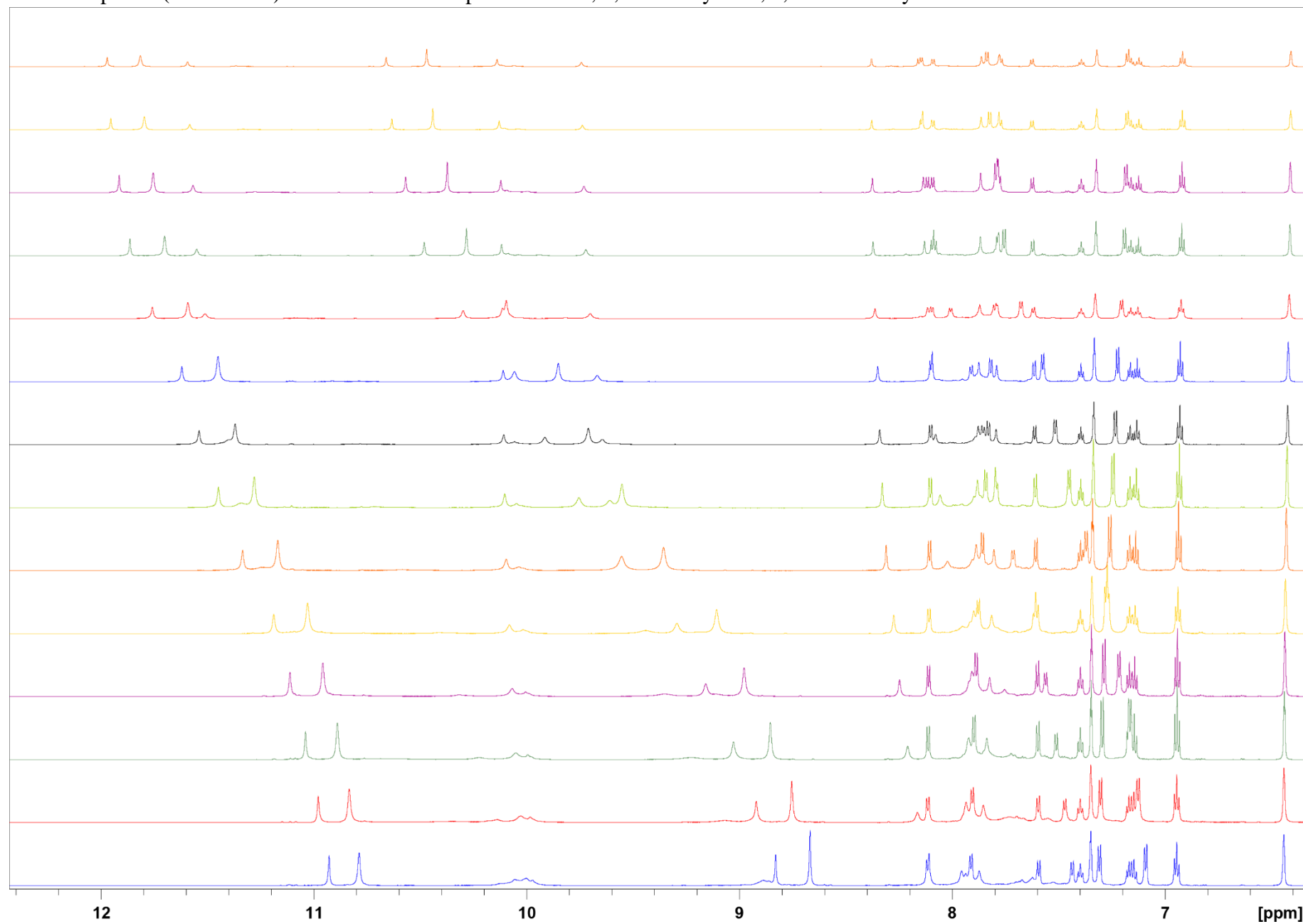
Stacked ^1H NMR spectra (700.1 MHz) of a mixture of receptors **MC005**; 1,3-diindolylurea; 1,3-dicarbazolylurea + TBA-lactate



Stacked ^1H NMR spectra (700.1 MHz) of a mixture of receptors **MC006**; 1,3-diindolylurea; 1,3-dicarbazolylurea + TBA-lactate



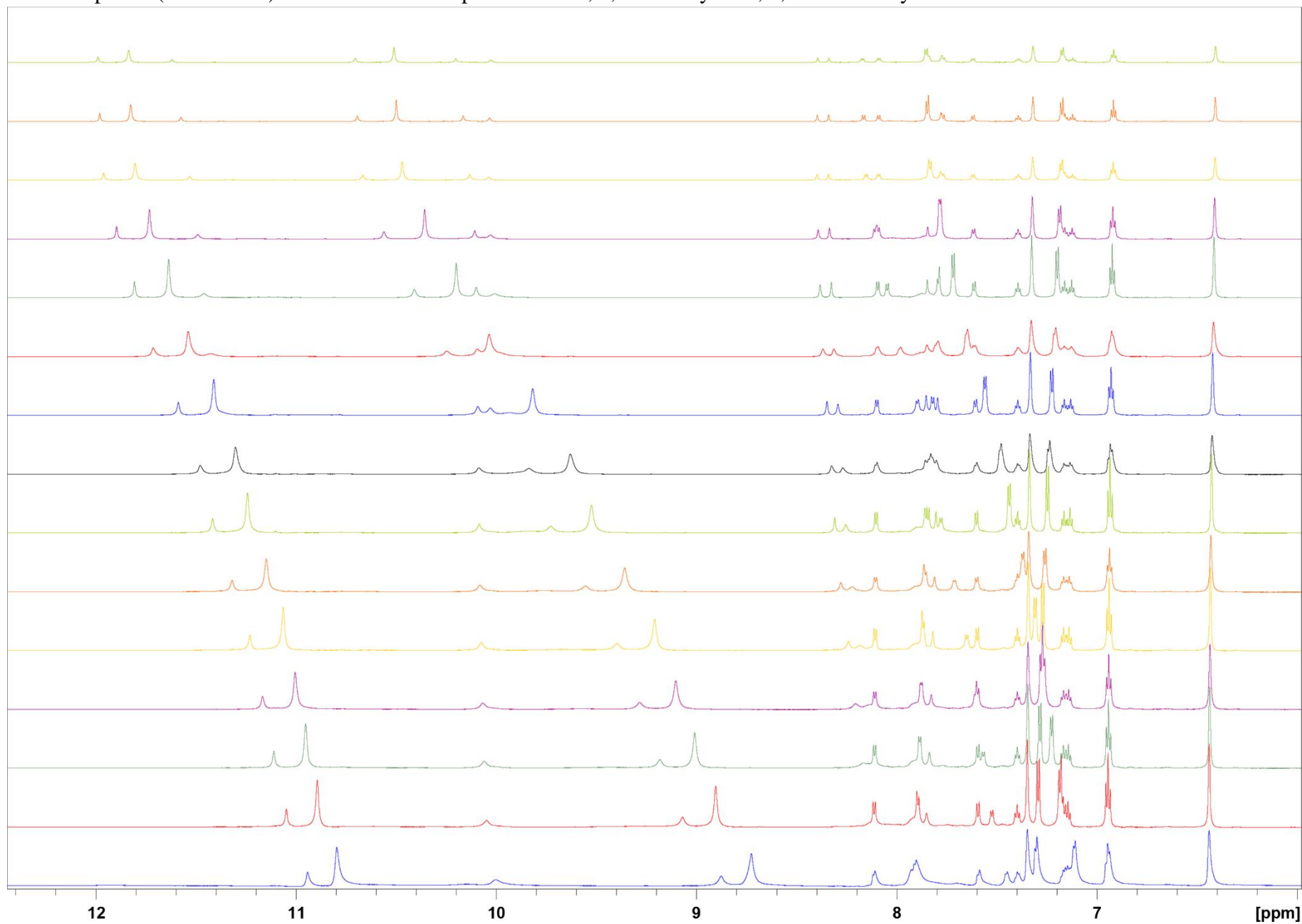
Stacked ^1H NMR spectra (700.1 MHz) of a mixture of receptors **MC007**; 1,3-diindolylurea; 1,3-dicarbazolylurea + TBA-lactate



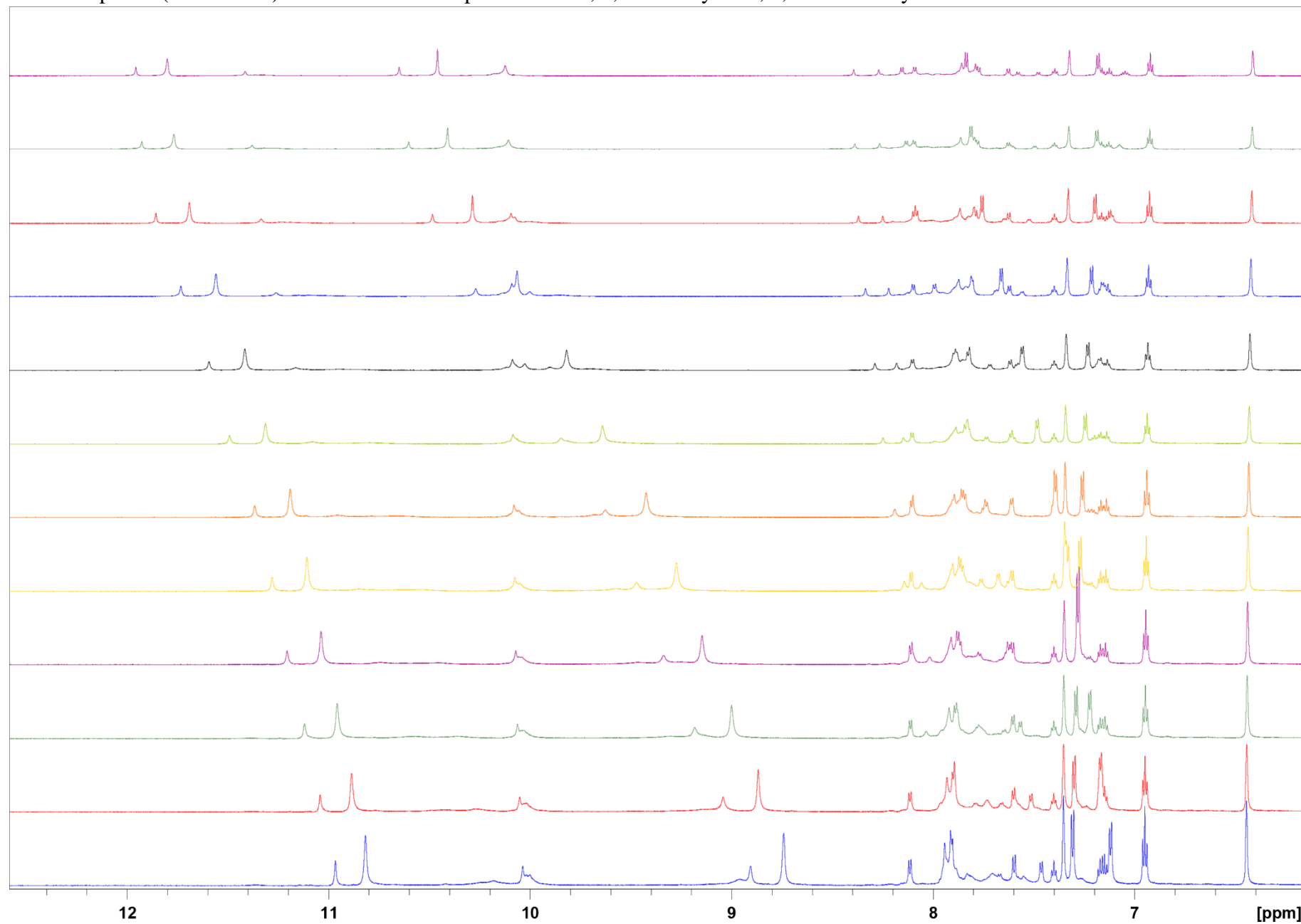
Stacked ^1H NMR spectra (700.1 MHz) of a mixture of receptors **MC008**; 1,3-diindolylurea; 1,3-dicarbazolylurea + TBA-lactate



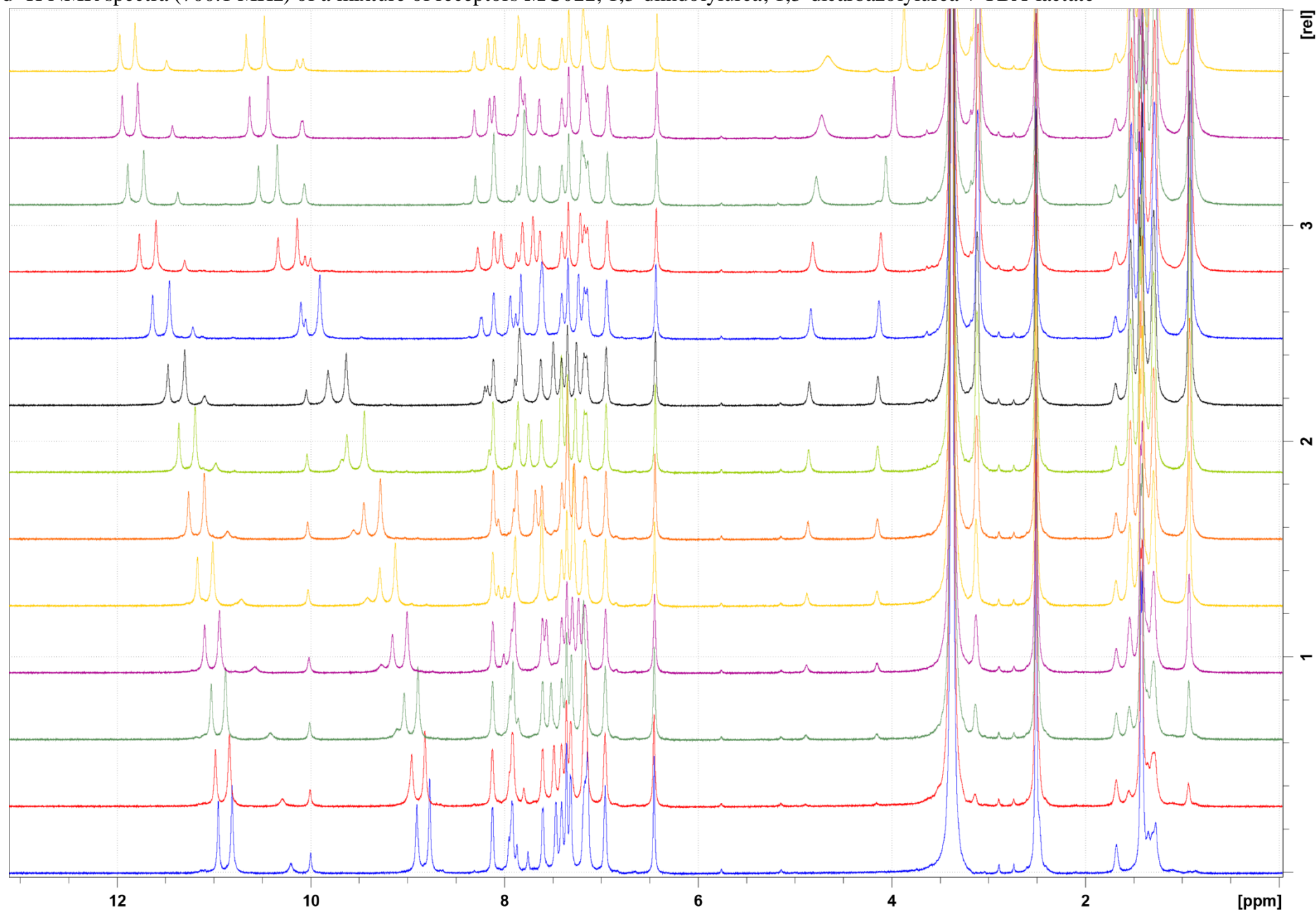
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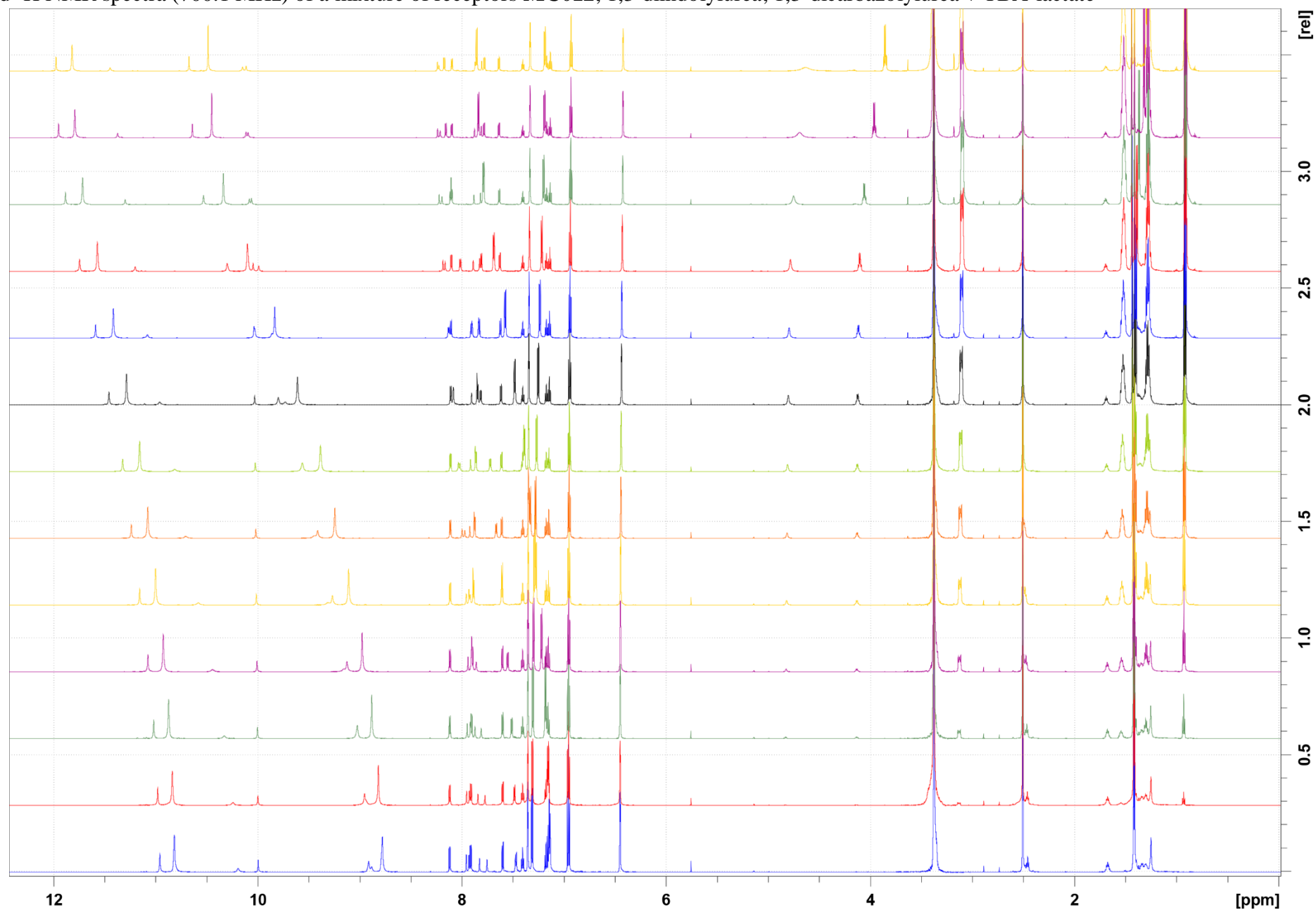
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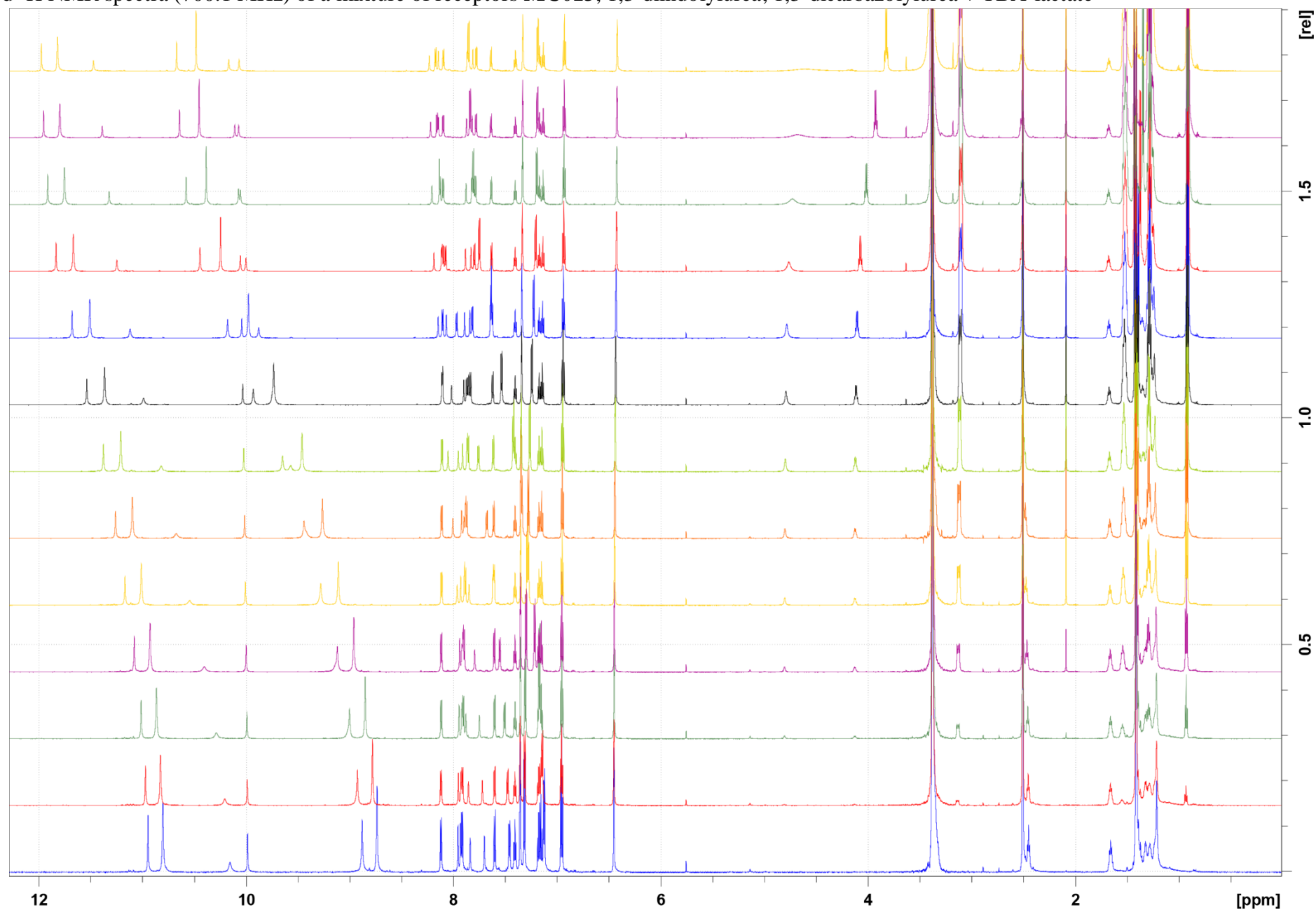
Stacked ^1H NMR spectra (700.1 MHz) of a mixture of receptors **MC011**; 1,3-diindolylurea; 1,3-dicarbazolylurea + TBA-lactate



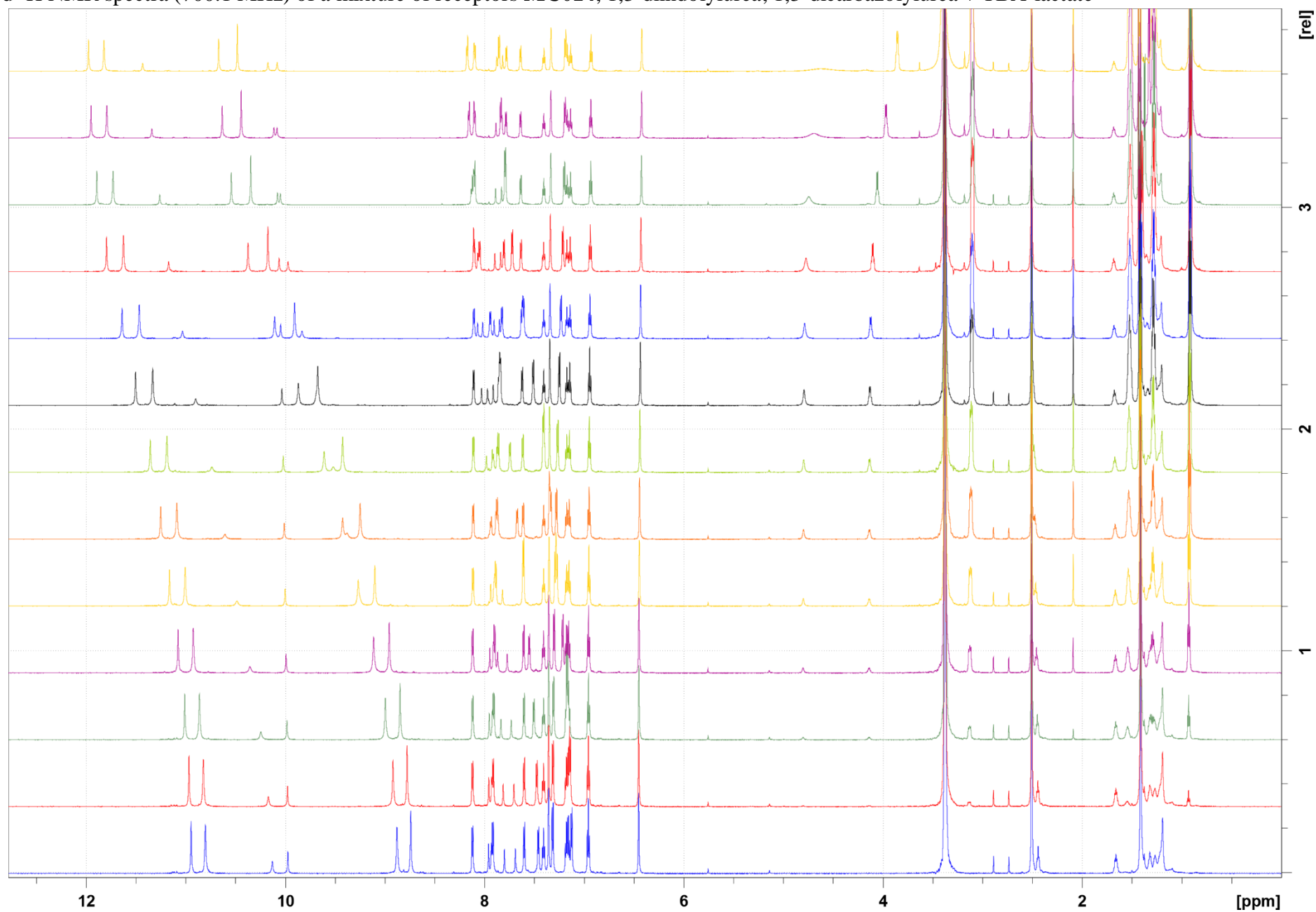
Stacked ^1H NMR spectra (700.1 MHz) of a mixture of receptors **MC012**; 1,3-diindolylurea; 1,3-dicarbazolylurea + TBA-lactate



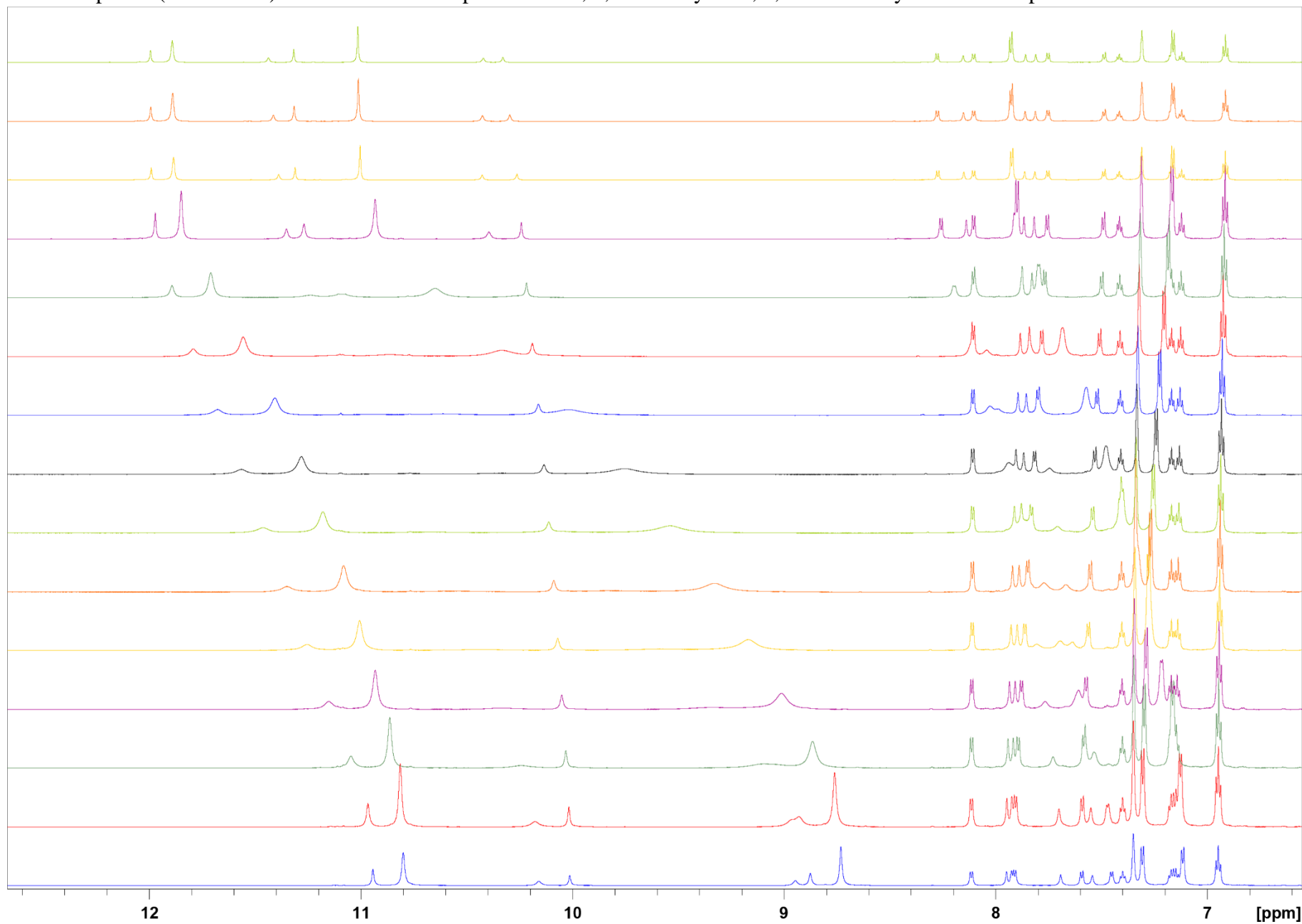
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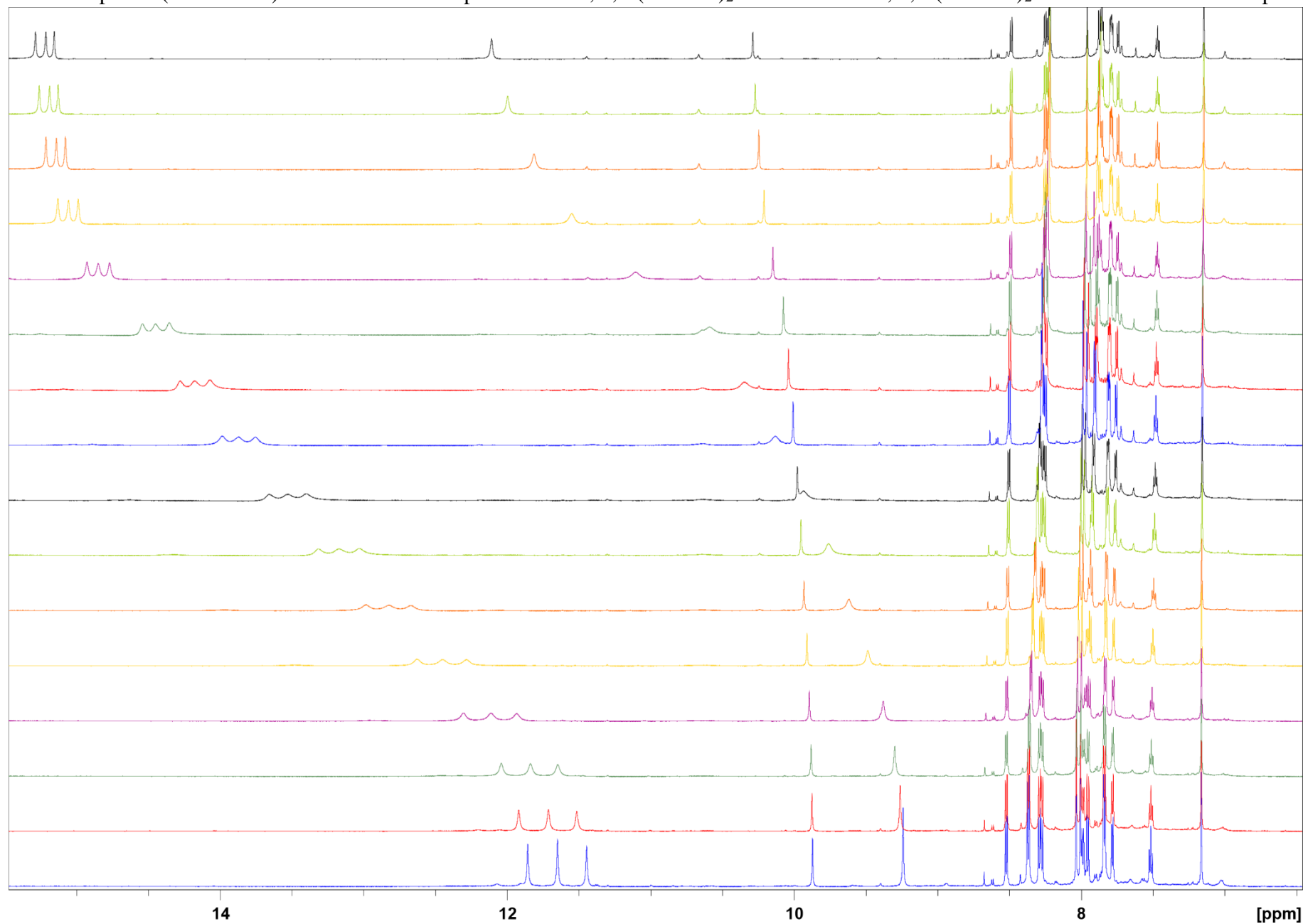
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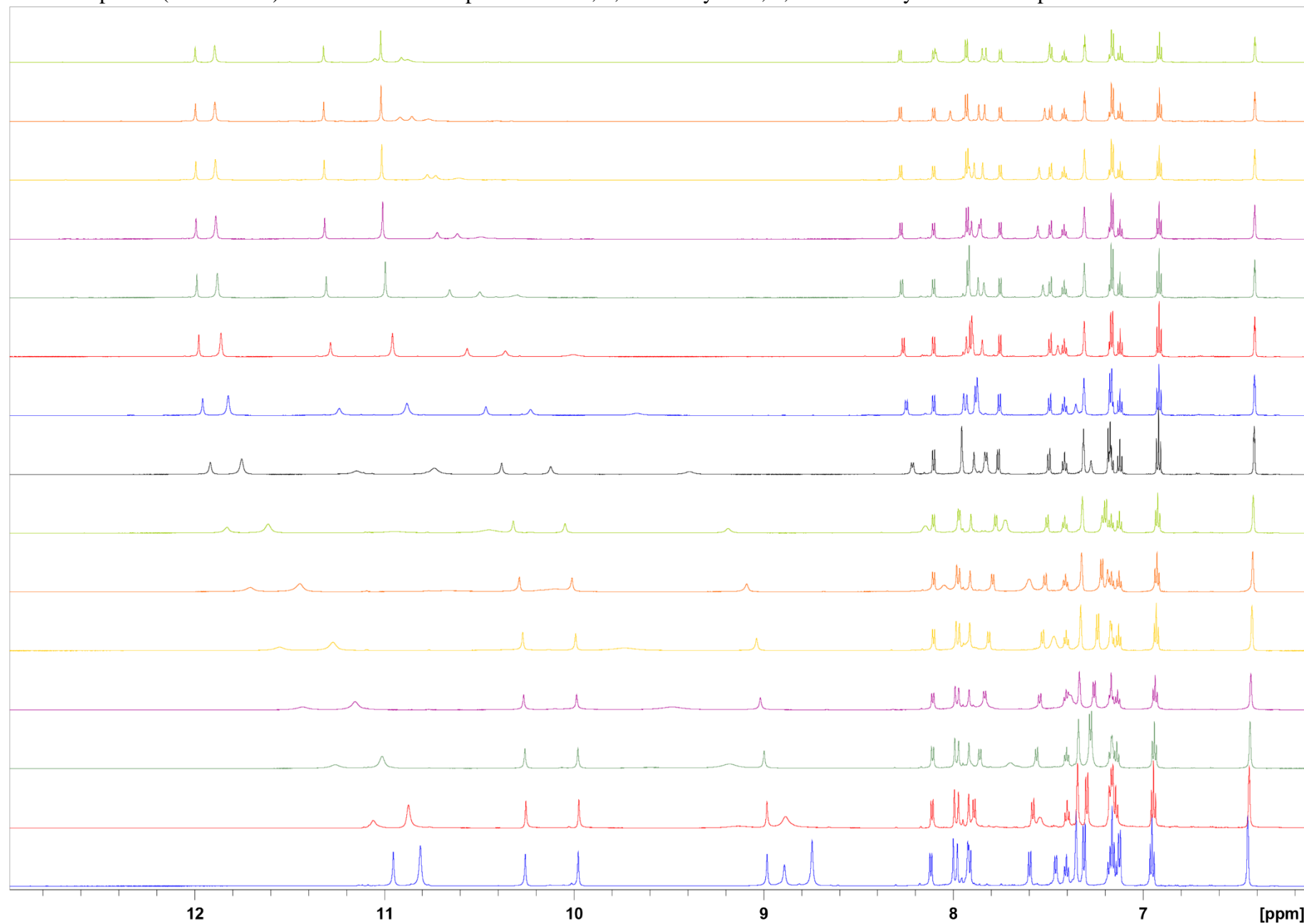
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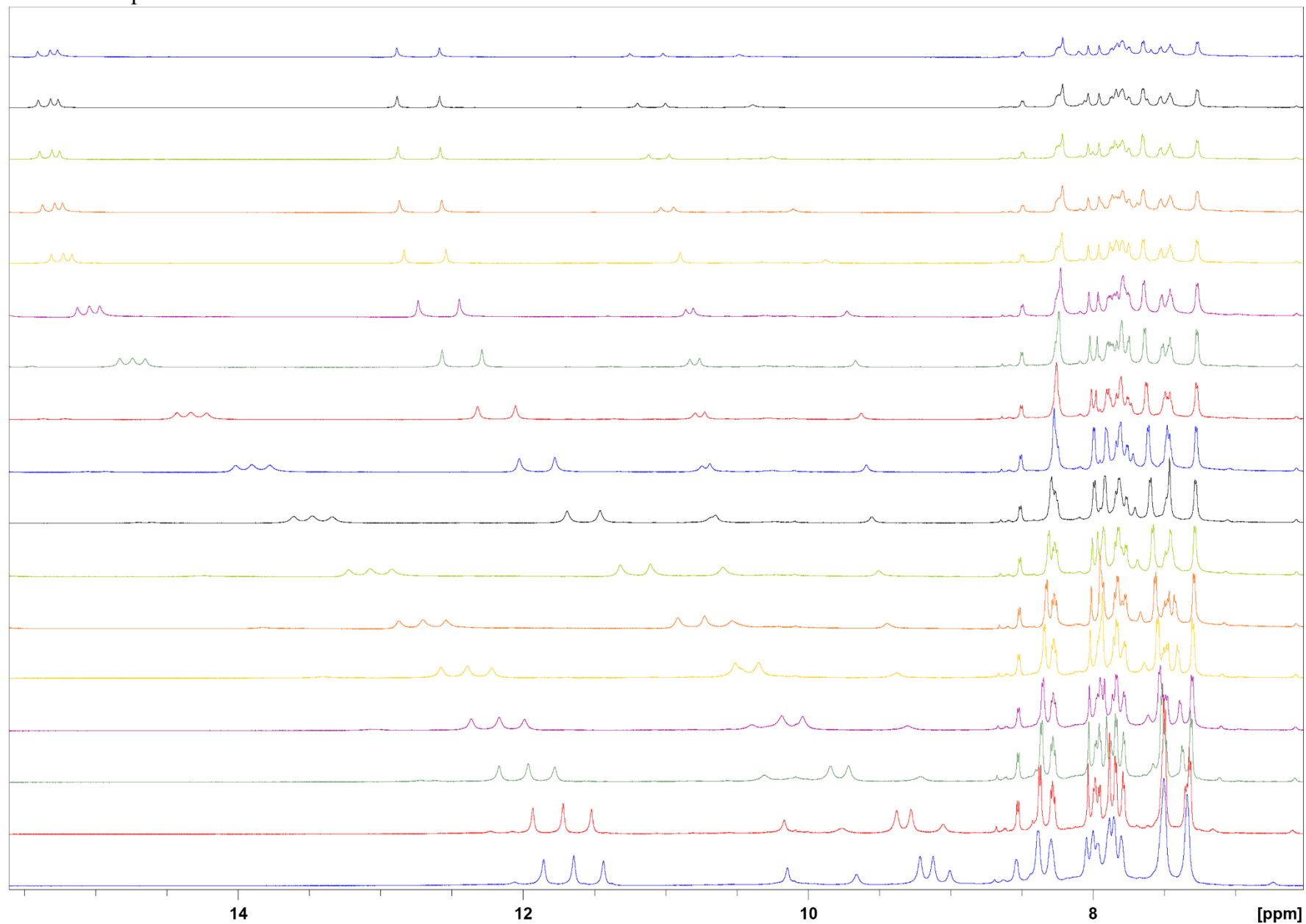
Stacked ^1H NMR spectra (700.1 MHz) of a mixture of receptors **MC001**; 2,7-(COOBu) $_2$ -indolocarbazole; 2,9-(COOBu) $_2$ -indolocarbazole + TBA-pivalate



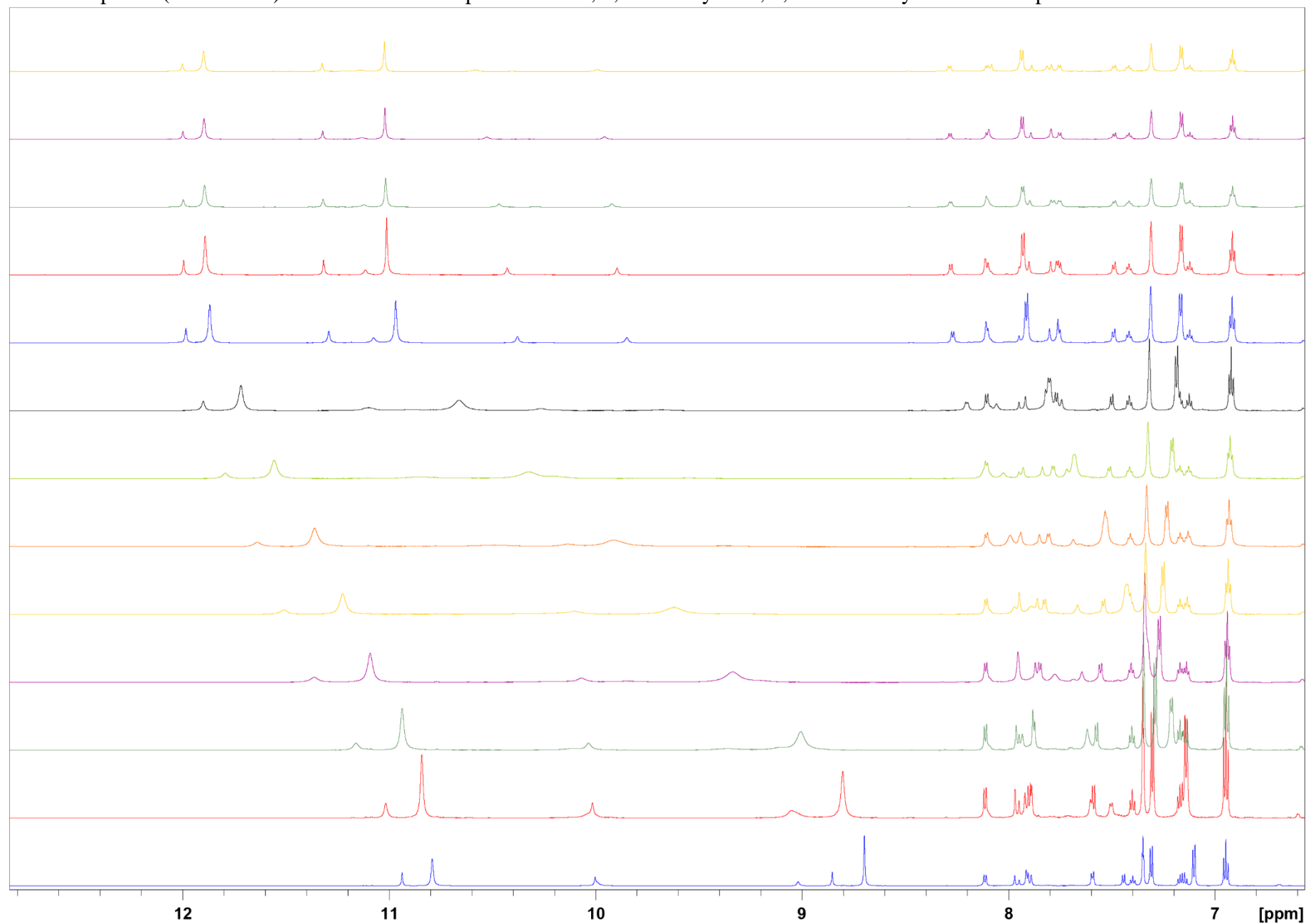
Stacked ^1H NMR spectra (700.1 MHz) of a mixture of receptors **MC003**; 1,3-diindolylurea; 1,3-dicarbazolylurea + TBA-pivalate



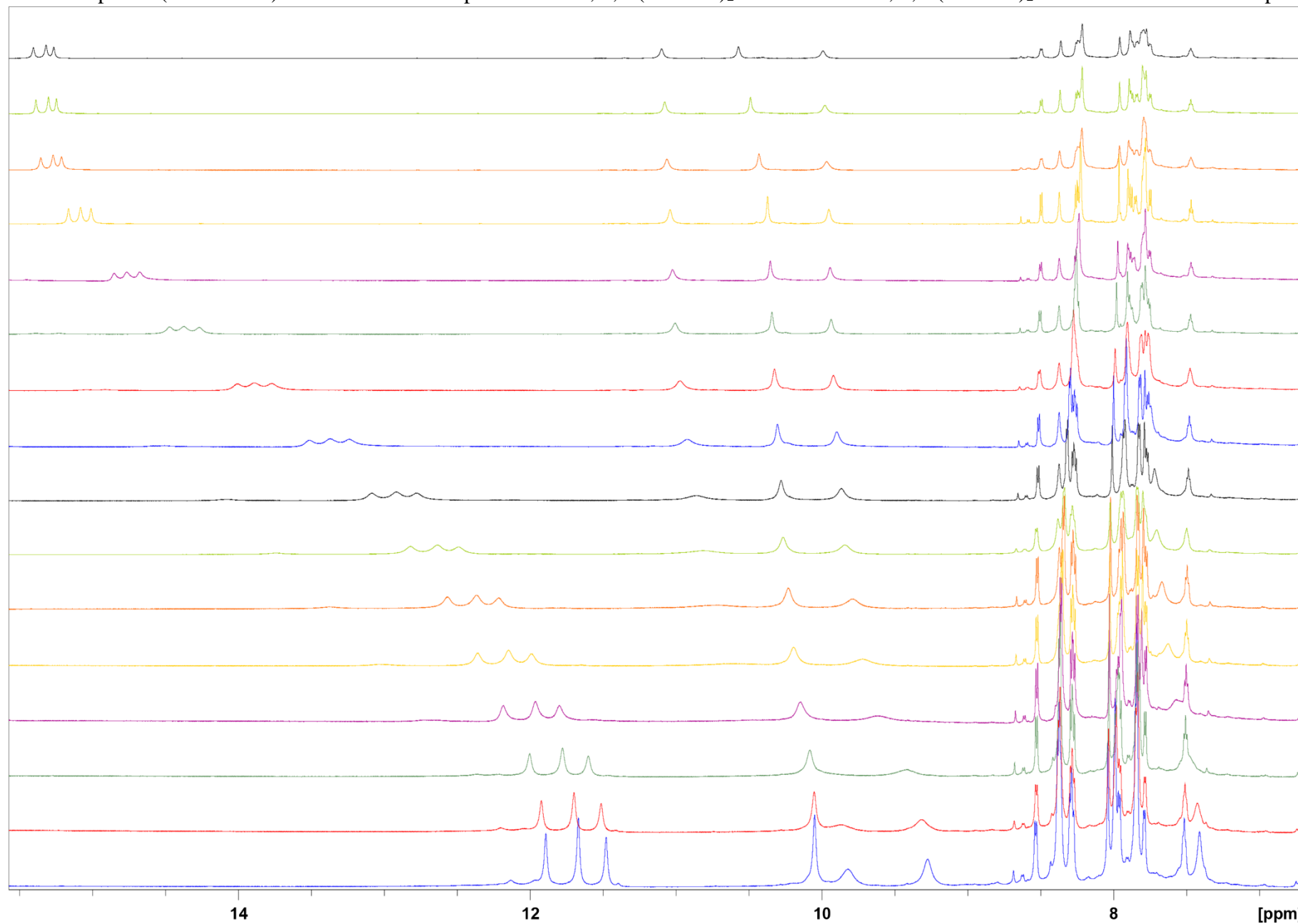
Stacked ^1H NMR spectra (700.1 MHz) of a mixture of receptors **MC004**; 3,4,4- Cl_3 -diphenylurea; 2,7-(COOBu) $_2$ -indolocarbazole; 2,9-(COOBu) $_2$ -indolocarbazole + TBA-pivalate



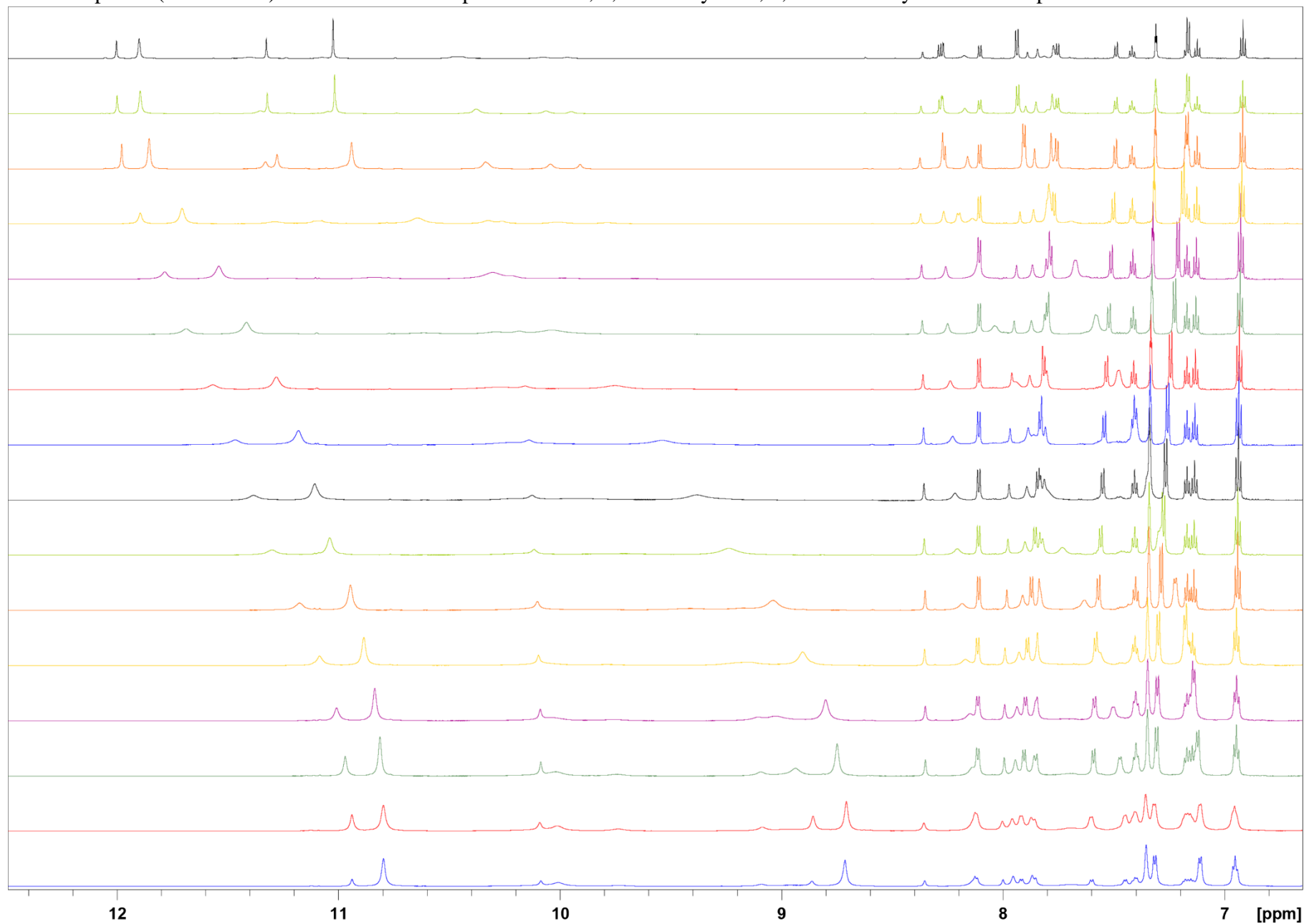
Stacked ^1H NMR spectra (700.1 MHz) of a mixture of receptors **MC005**; 1,3-diindolylurea; 1,3-dicarbazolyurea + TBA-pivalate



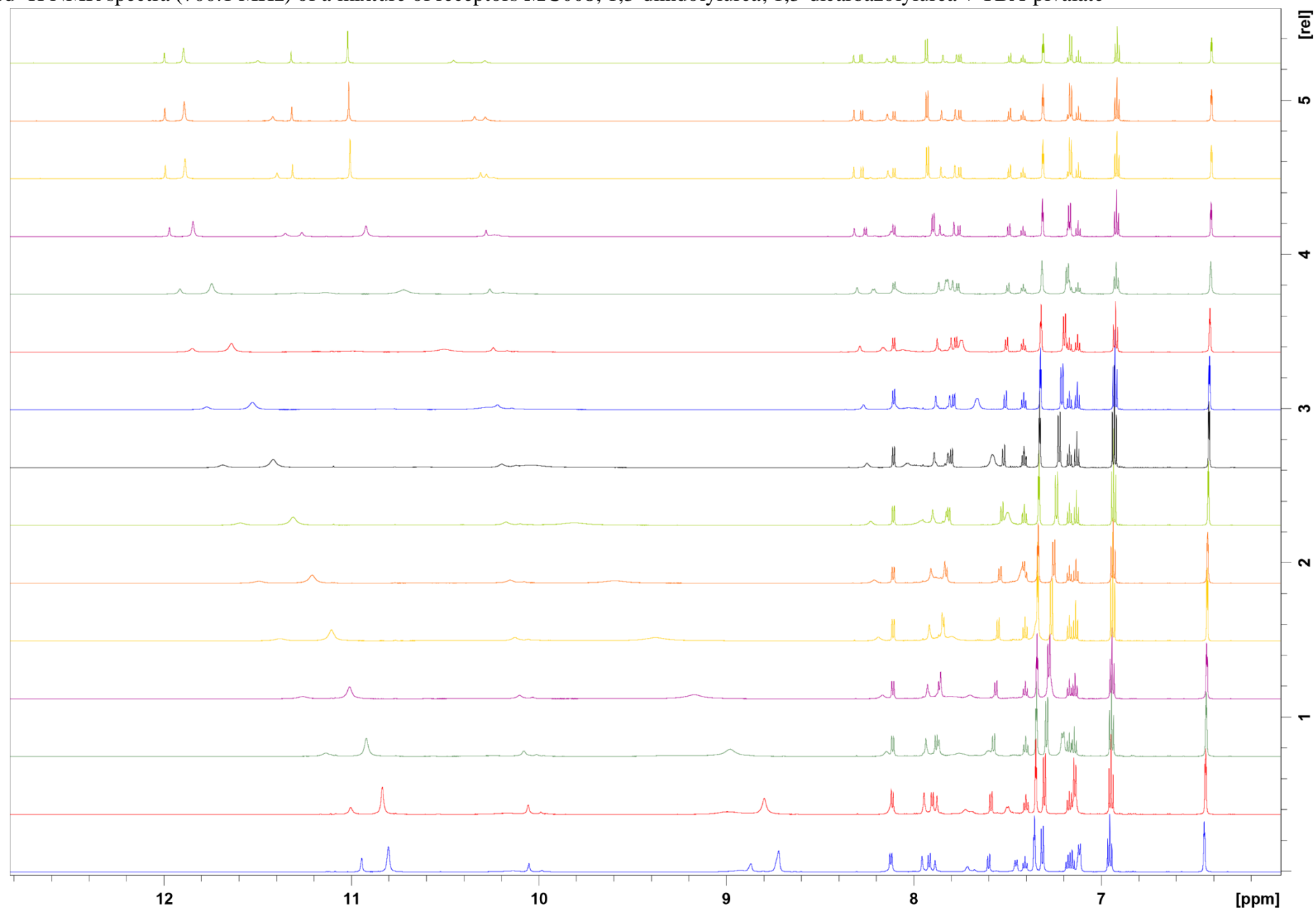
Stacked ^1H NMR spectra (700.1 MHz) of a mixture of receptors **MC006**; 2,7-(COOBu) $_2$ -indolocarbazole; 2,9-(COOBu) $_2$ -indolocarbazole + TBA-pivalate



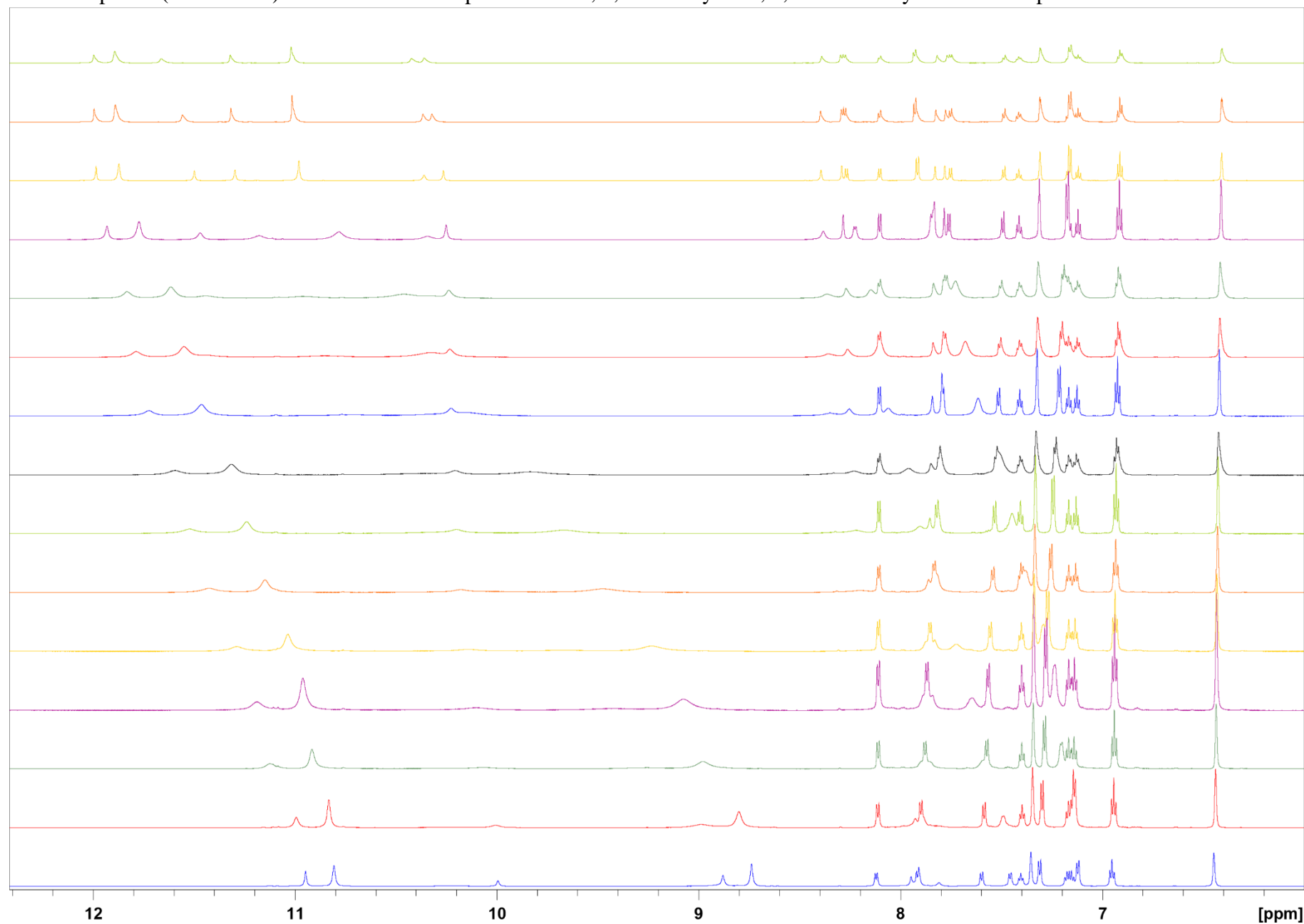
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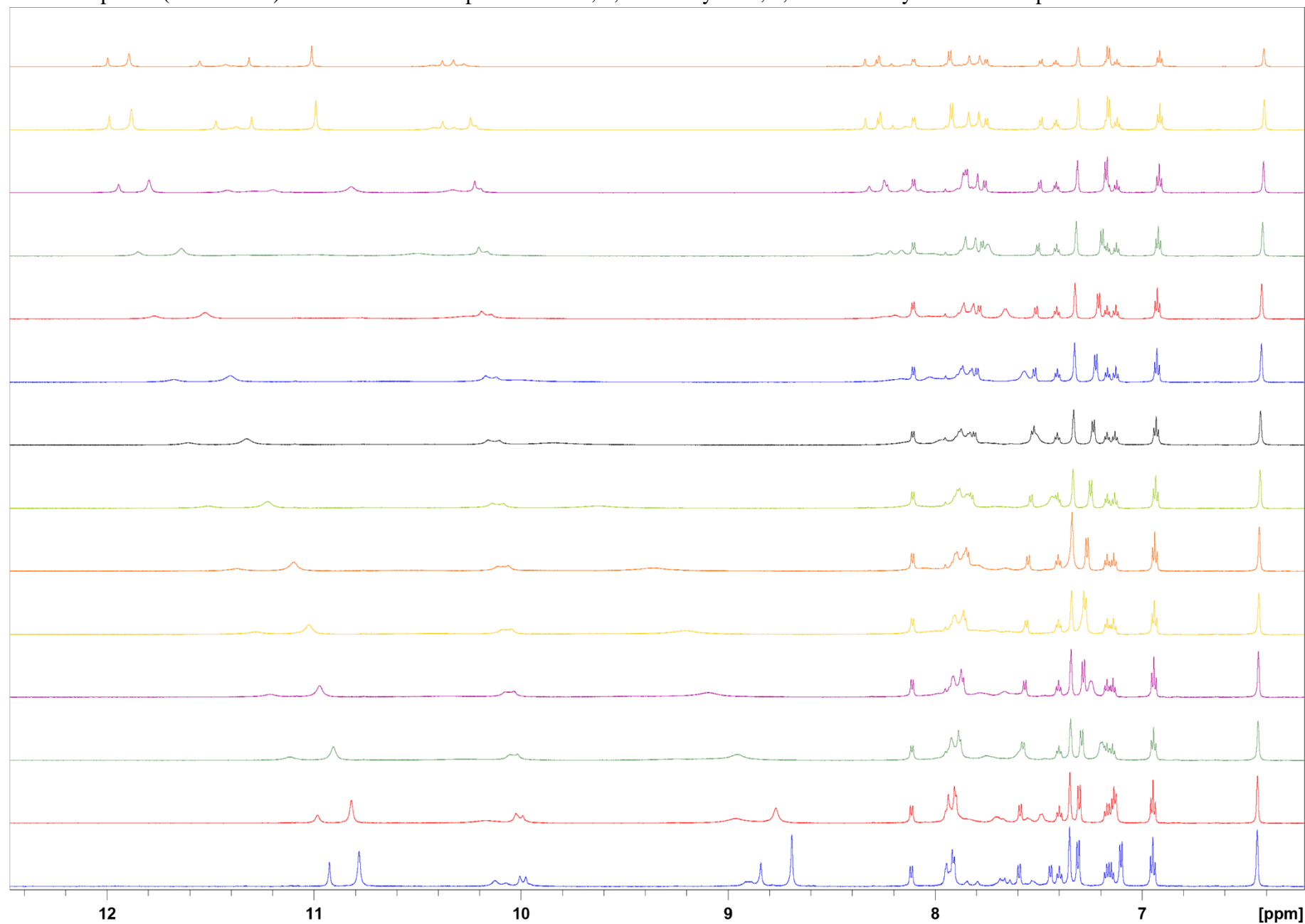
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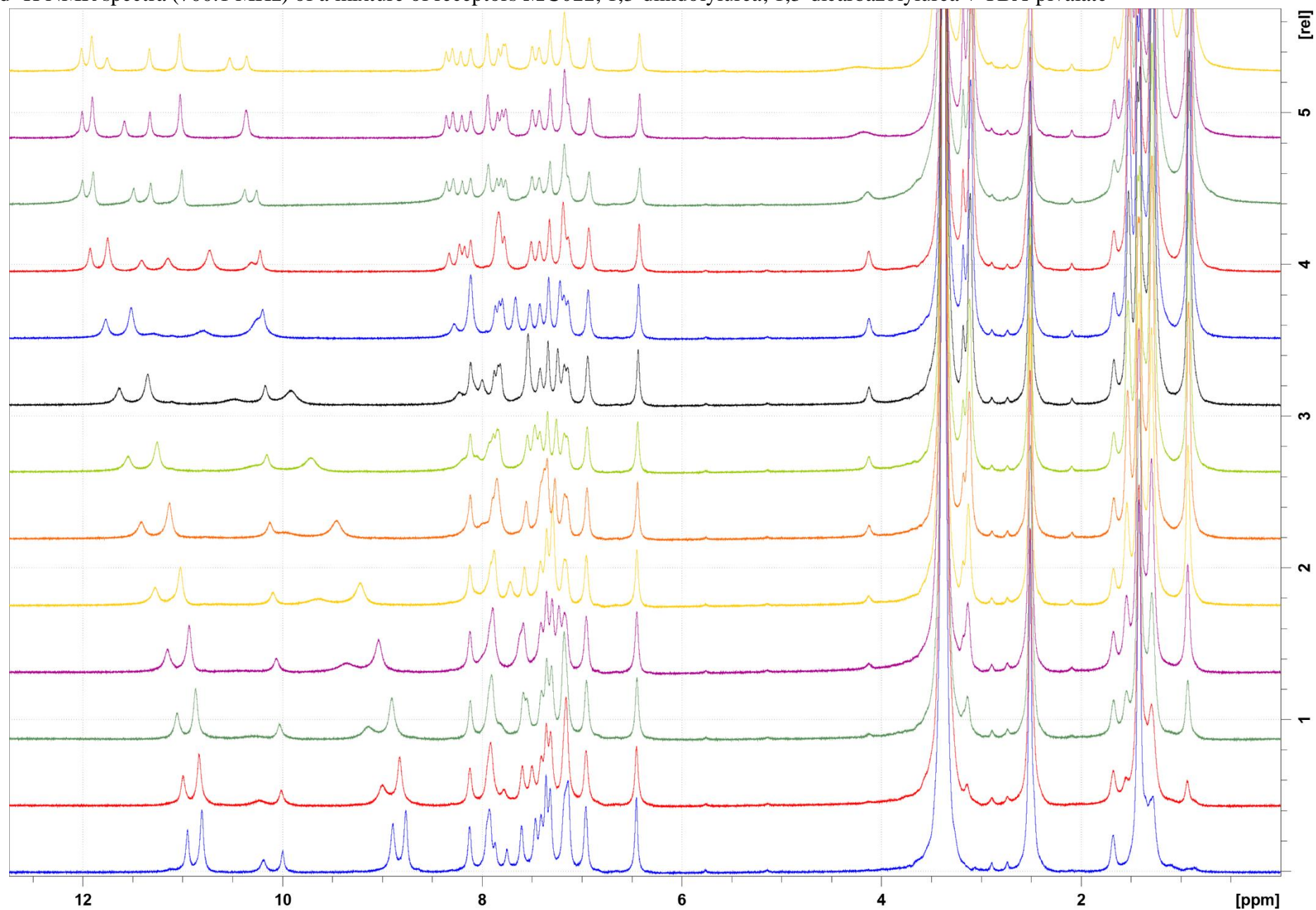
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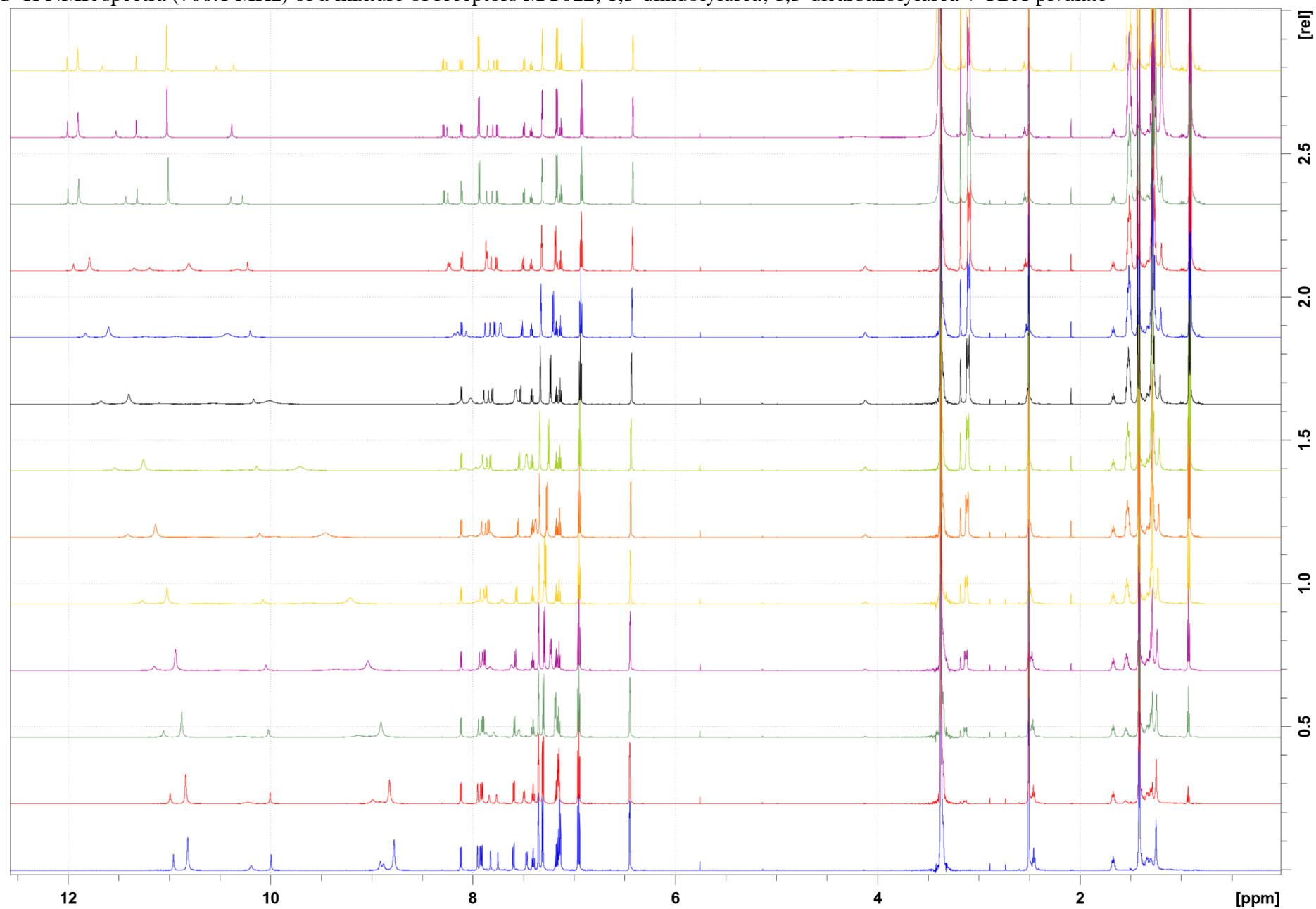
Stacked ^1H NMR spectra (700.1 MHz) of a mixture of receptors **MC010**; 1,3-diindolylurea; 1,3-dicarbazolylurea + TBA-pivalate



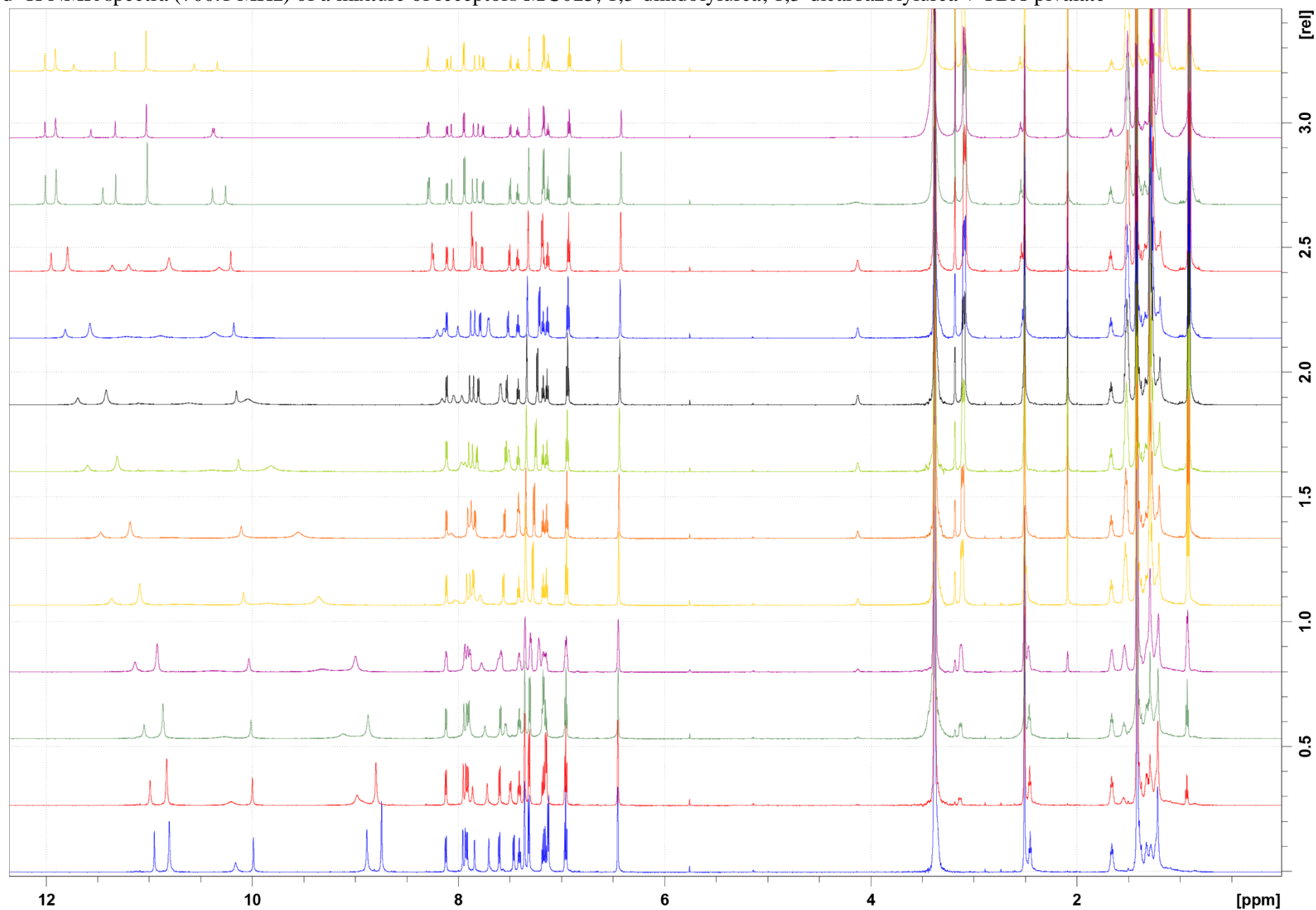
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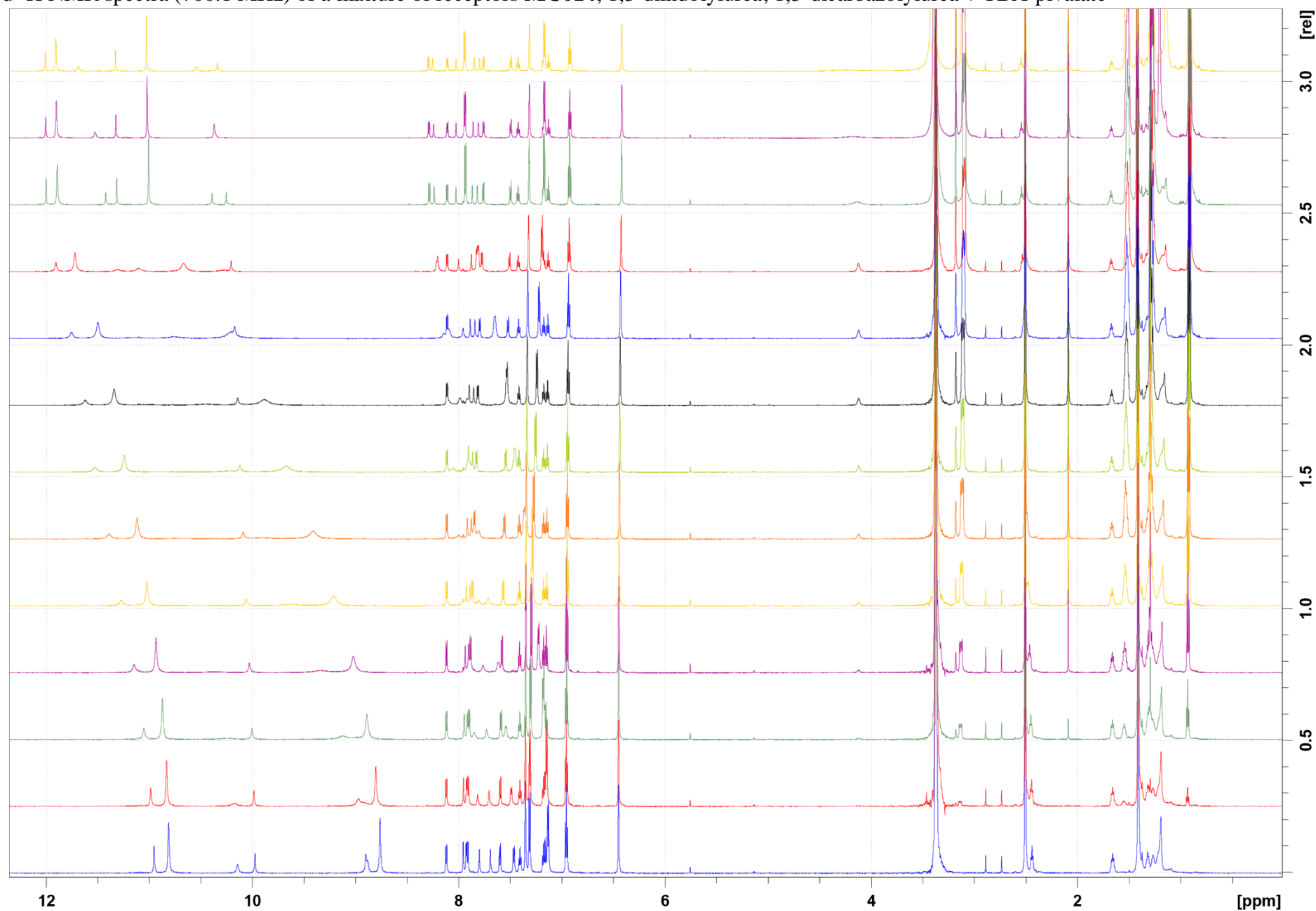
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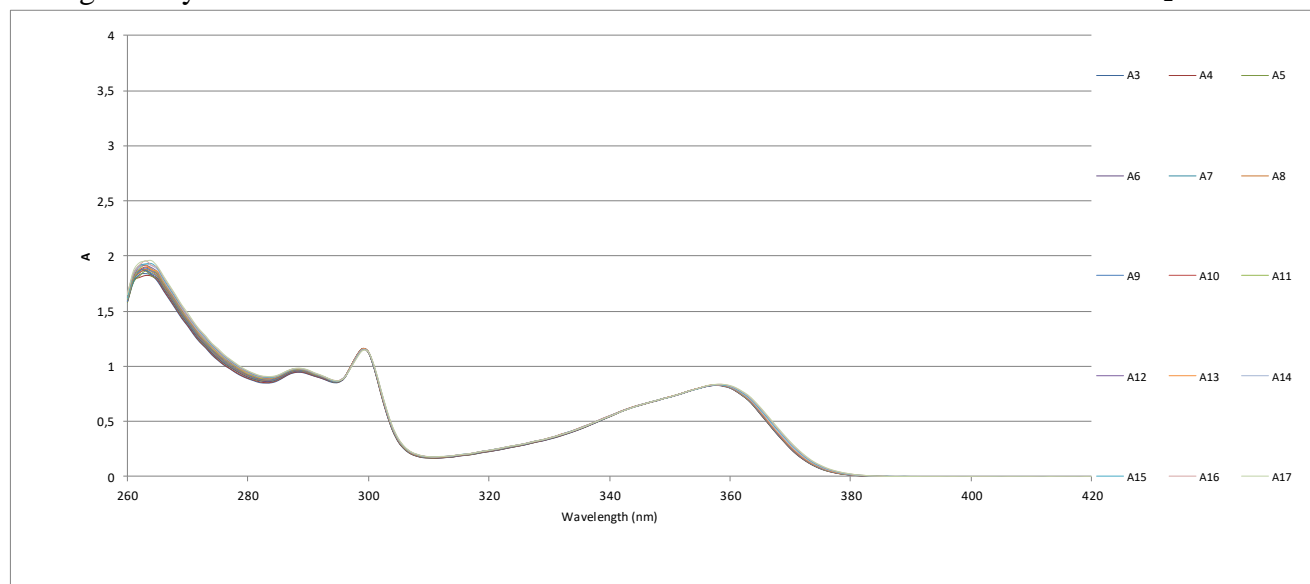
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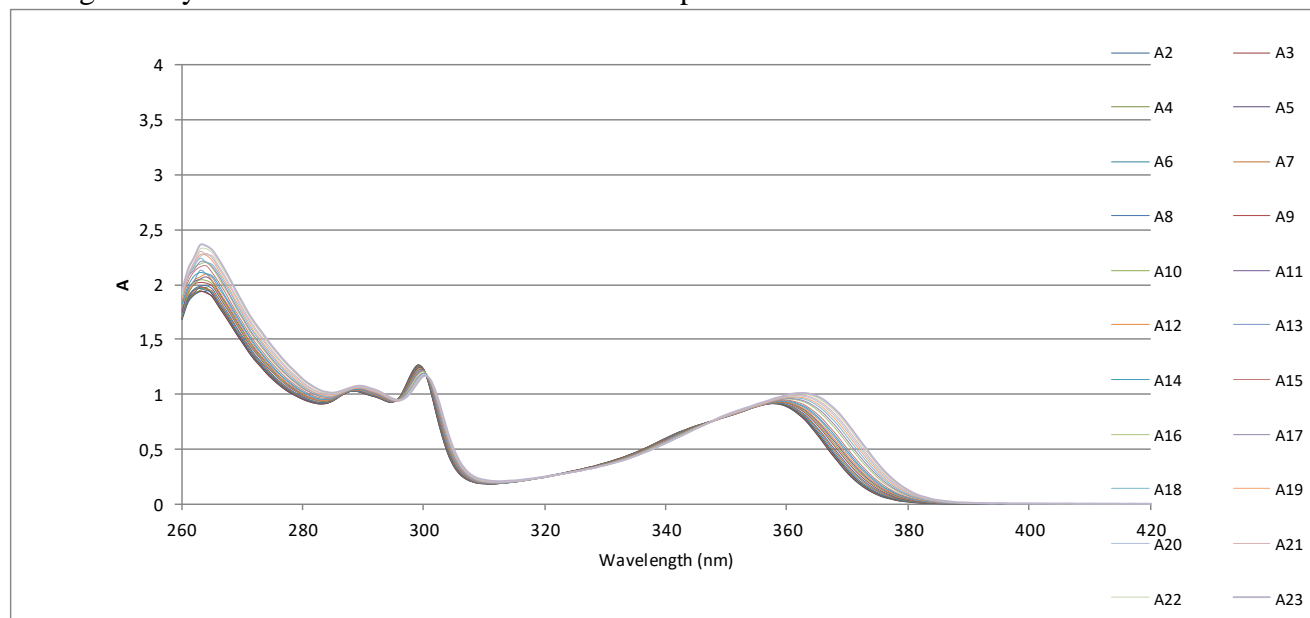
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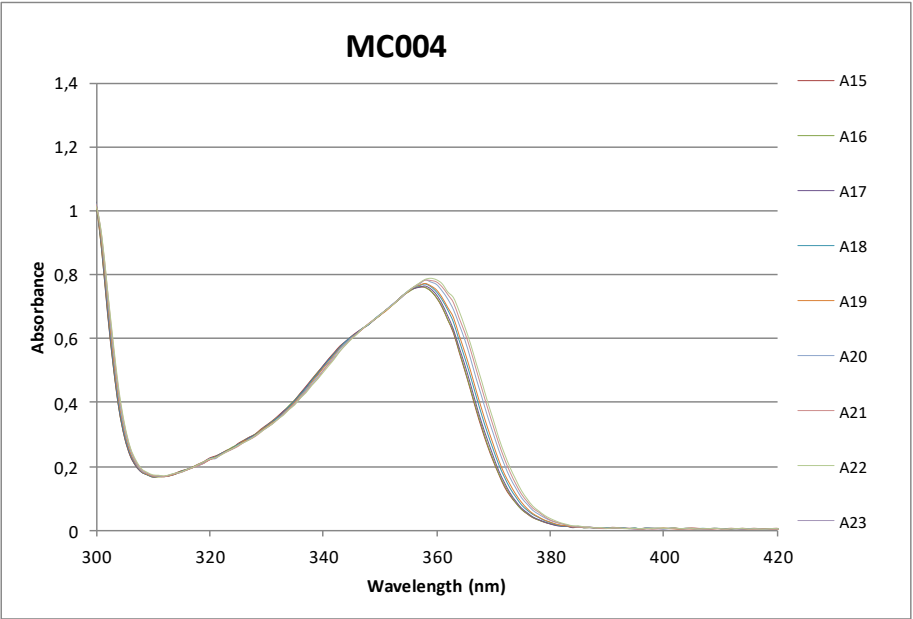
UV-vis spectra of absolute binding affinity measurement for **MC002** with TBA-lactate in 99.5%:0.5% m/m DMSO-H₂O



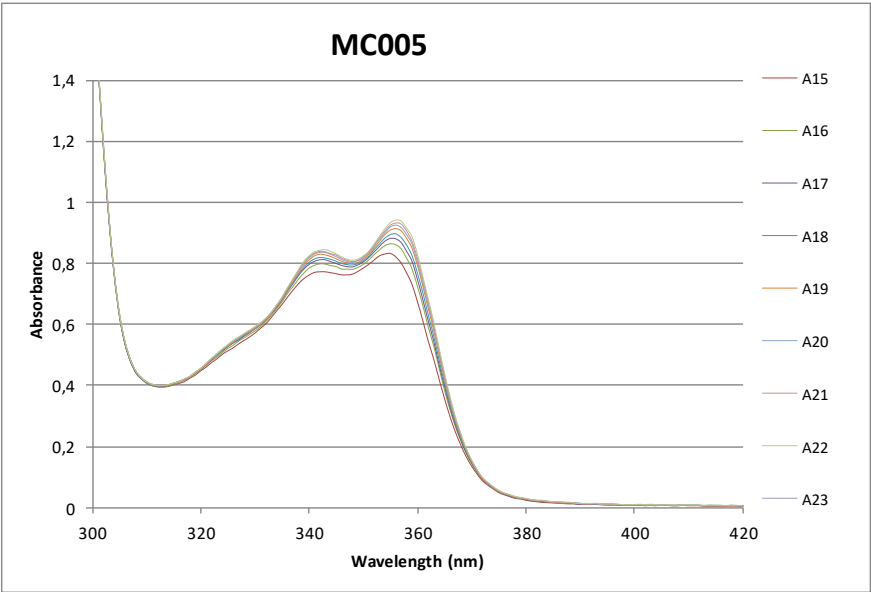
UV-vis spectra of absolute binding affinity measurement for **MC002** with TBA-pivalate 99.5%:0.5% m/m DMSO-H₂O



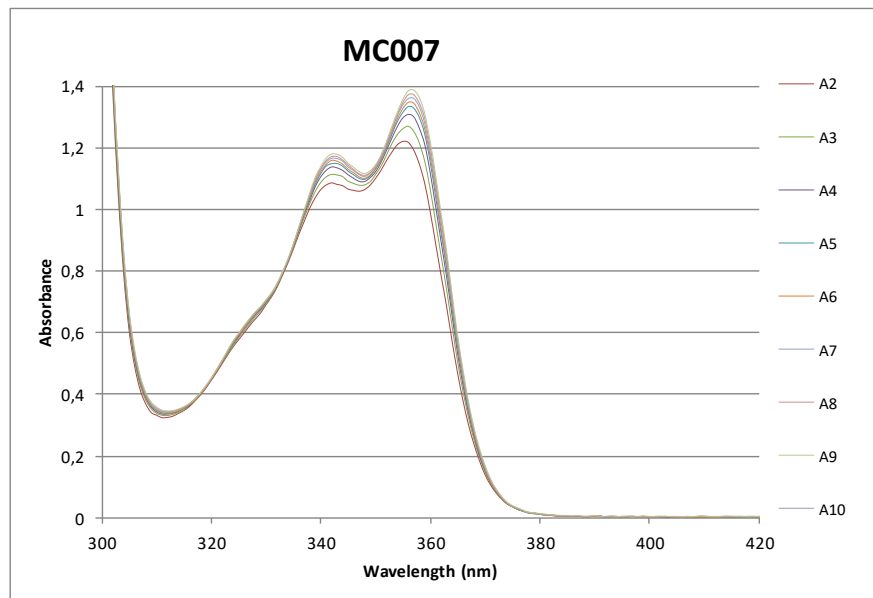
UV-vis spectra of absolute binding affinity measurement for **MC004** with TBA-acetate 90.0%:10.0% m/m DMSO-H₂O



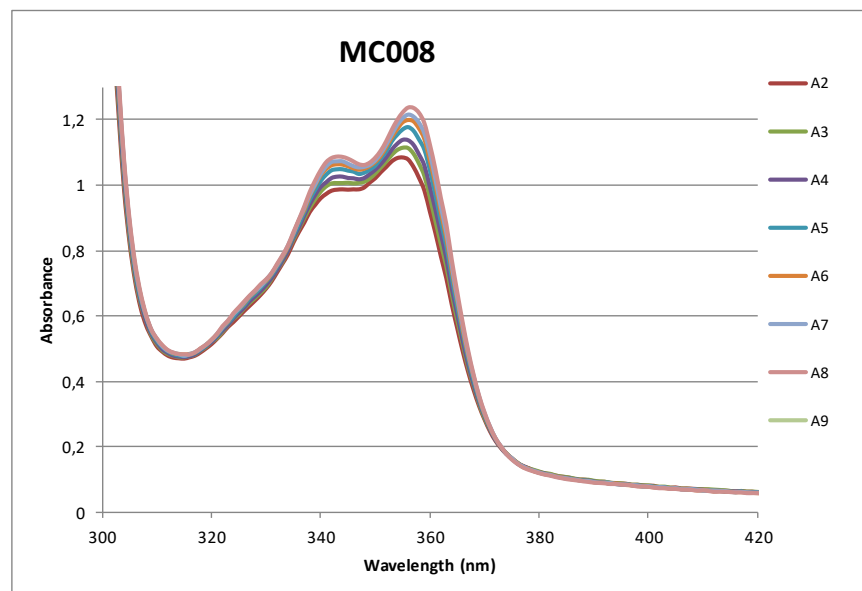
UV-vis spectra of absolute binding affinity measurement for **MC005** with TBA-acetate 90.0%:10.0% m/m DMSO-H₂O



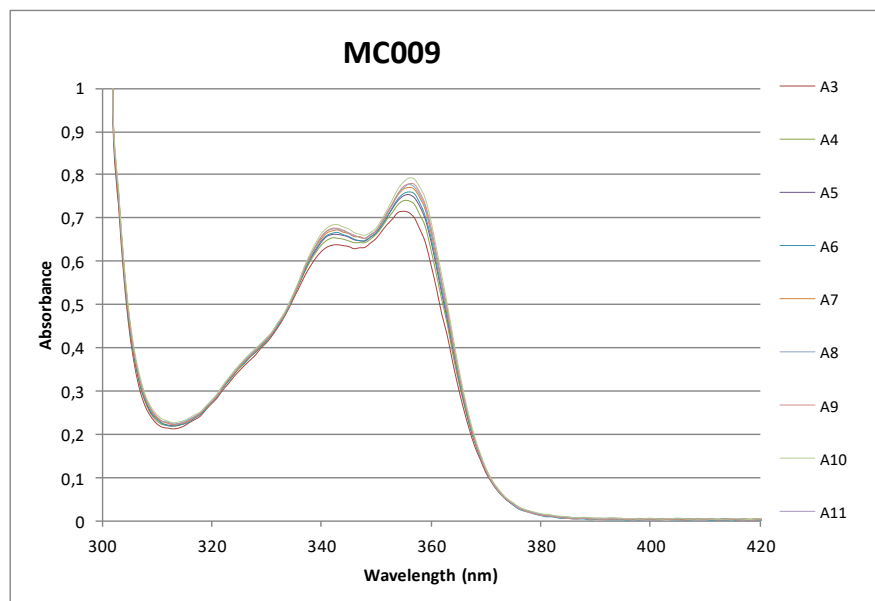
UV-vis spectra of absolute binding affinity measurement for **MC007** with TBA-acetate 90.0%:10.0% m/m DMSO-H₂O



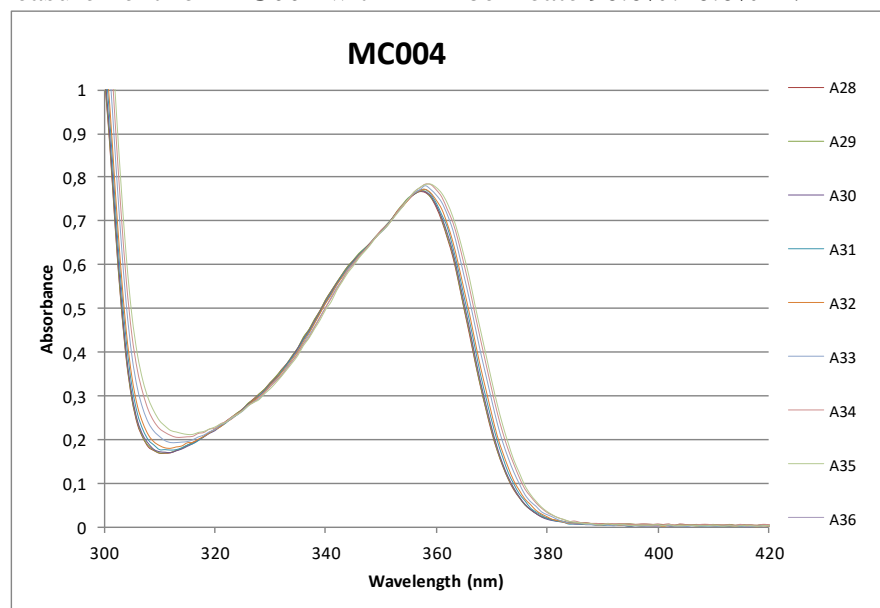
UV-vis spectra of absolute binding affinity measurement for **MC008** with TBA-acetate 90.0%:10.0% m/m DMSO-H₂O



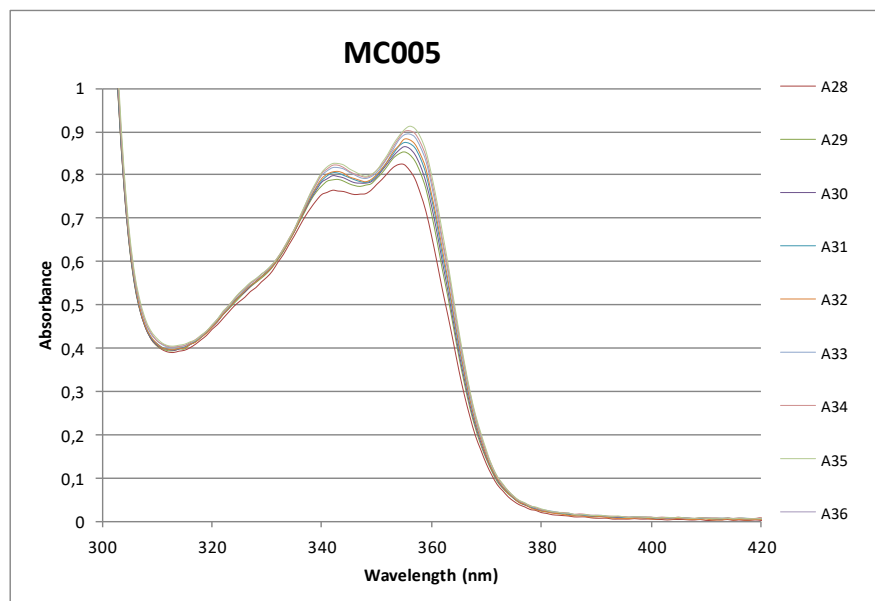
UV-vis spectra of absolute binding affinity measurement for **MC009** with TBA-acetate 90.0%:10.0% m/m DMSO-H₂O



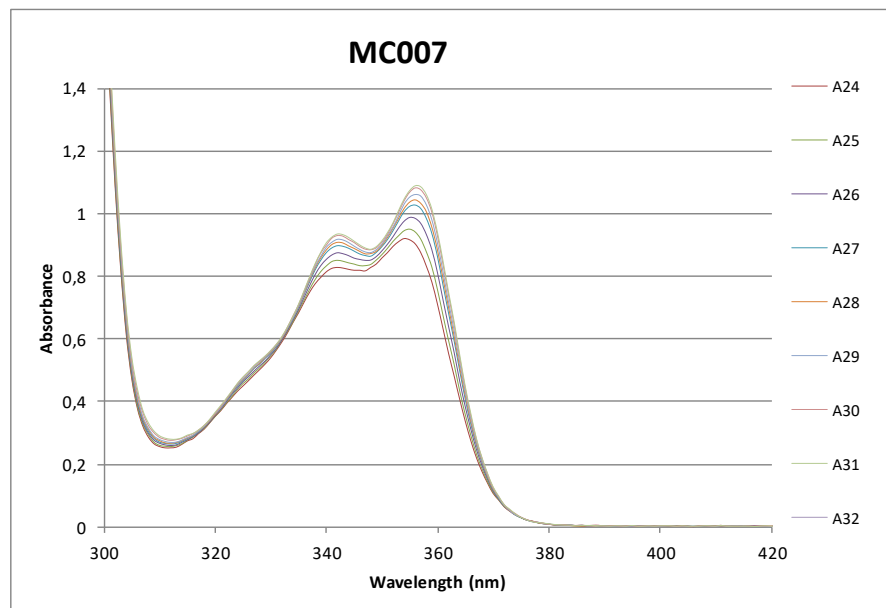
UV-vis spectra of absolute binding affinity measurement for **MC004** with TBA-benzoate 90.0%:10.0% m/m DMSO-H₂O



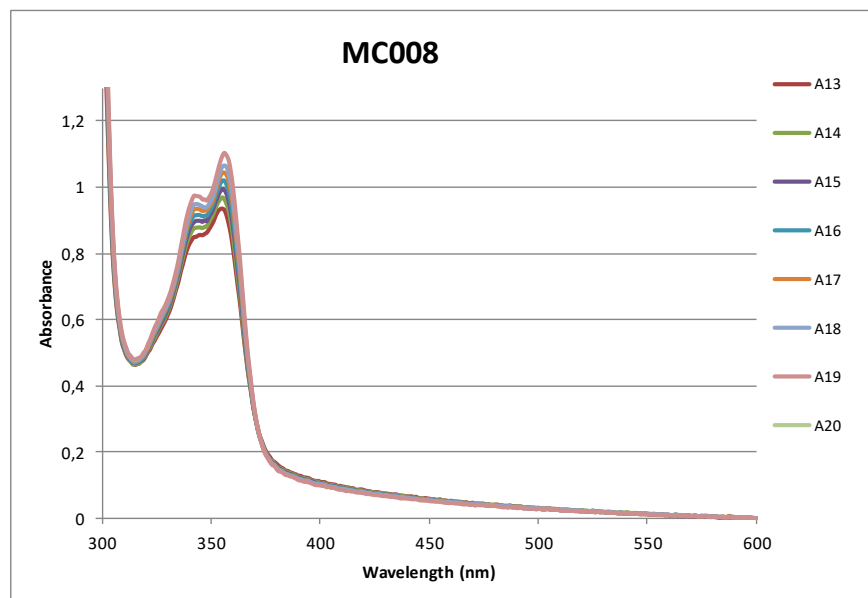
UV-vis spectra of absolute binding affinity measurement for **MC005** with TBA-benzoate 90.0%:10.0% m/m DMSO-H₂O



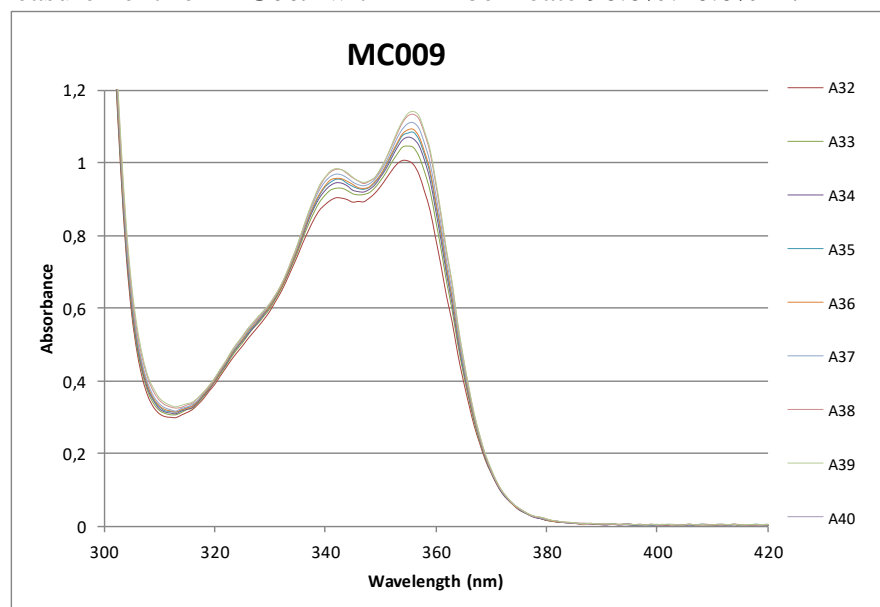
UV-vis spectra of absolute binding affinity measurement for **MC007** with TBA-benzoate 90.0%:10.0% m/m DMSO-H₂O



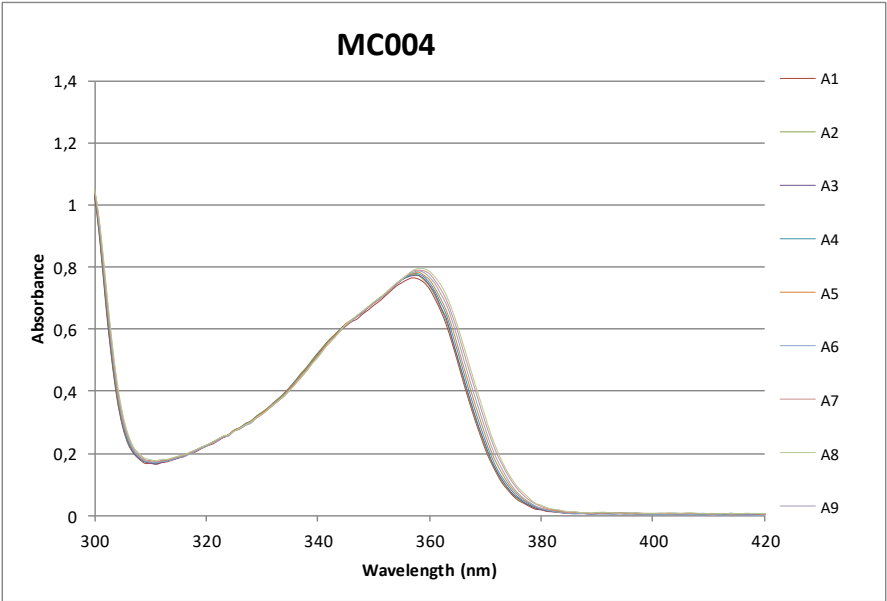
UV-vis spectra of absolute binding affinity measurement for **MC008** with TBA-benzoate 90.0%:10.0% m/m DMSO-H₂O



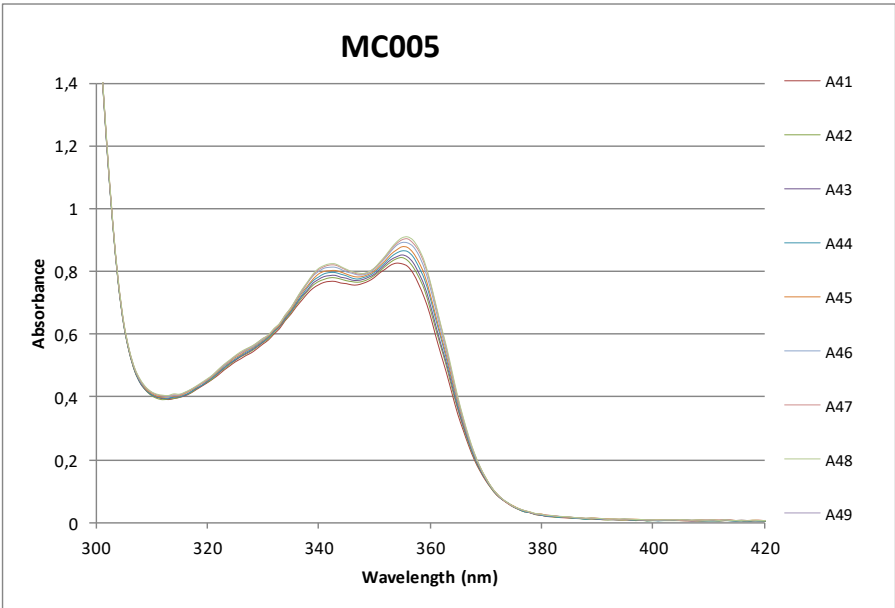
UV-vis spectra of absolute binding affinity measurement for **MC009** with TBA-benzoate 90.0%:10.0% m/m DMSO-H₂O



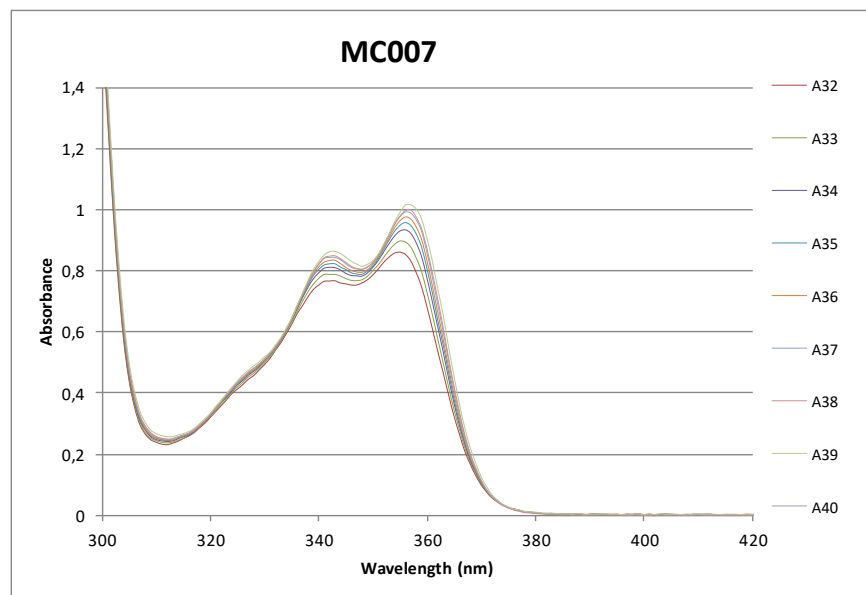
UV-vis spectra of absolute binding affinity measurement for **MC004** with TBA-formate 90.0%:10.0% m/m DMSO-H₂O



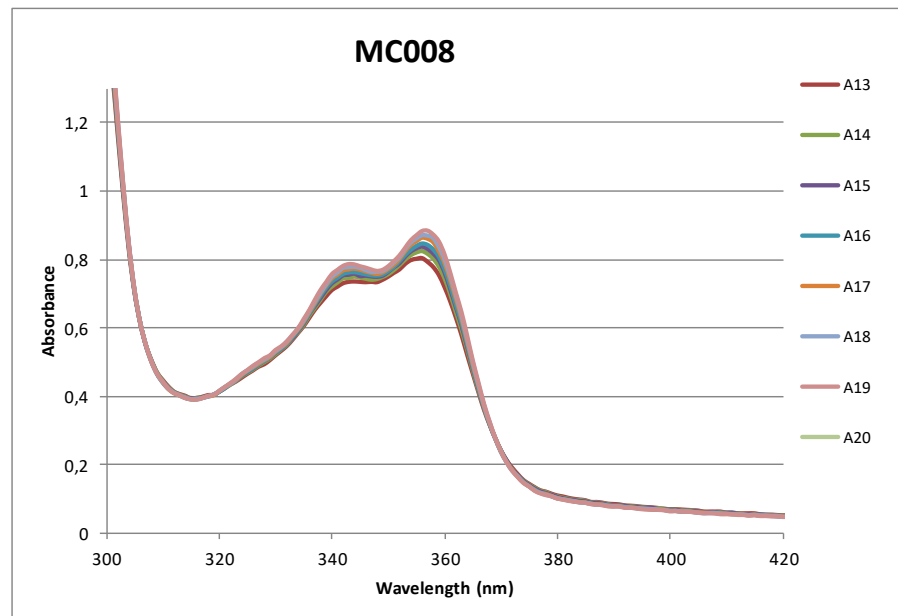
UV-vis spectra of absolute binding affinity measurement for **MC005** with TBA-formate 90.0%:10.0% m/m DMSO-H₂O



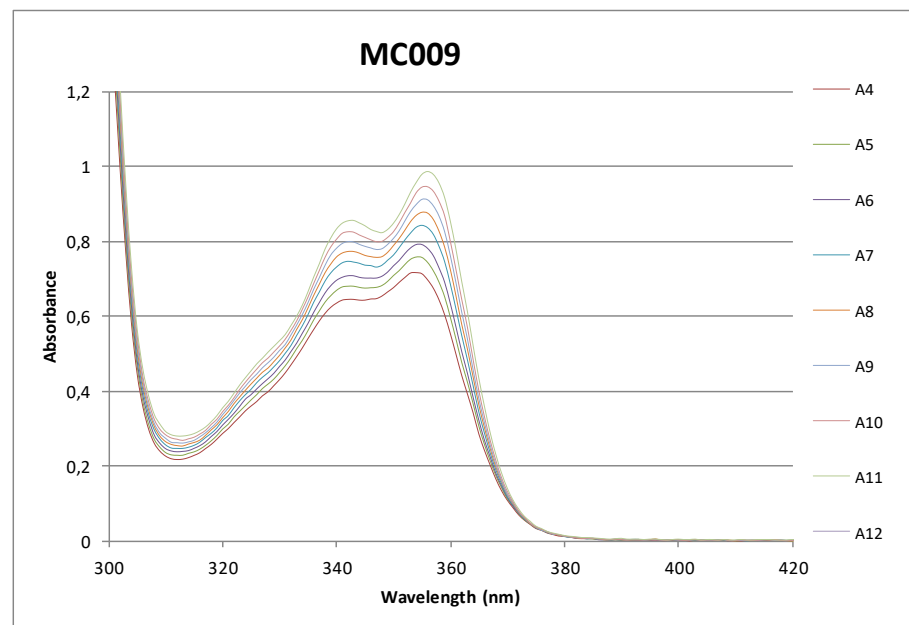
UV-vis spectra of absolute binding affinity measurement for **MC007** with TBA-formate 90.0%:10.0% m/m DMSO-H₂O



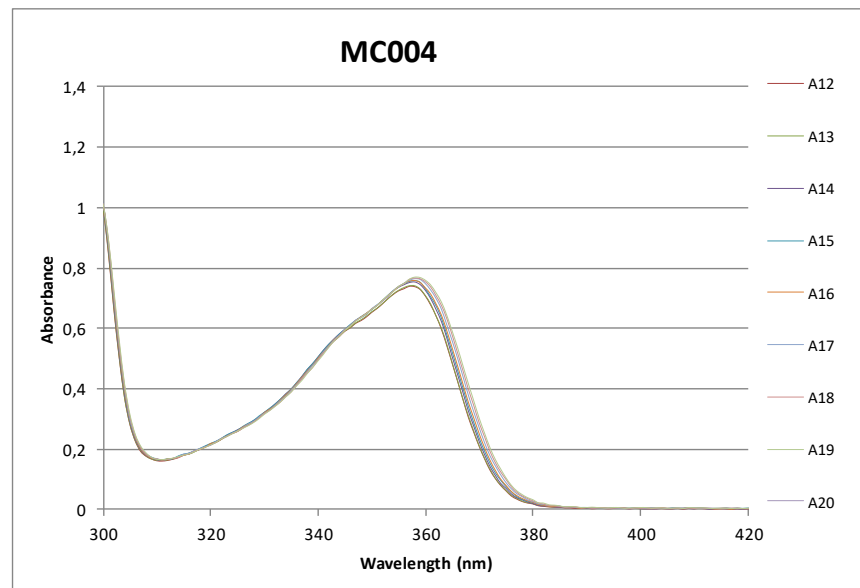
UV-vis spectra of absolute binding affinity measurement for **MC008** with TBA-formate 90.0%:10.0% m/m DMSO-H₂O



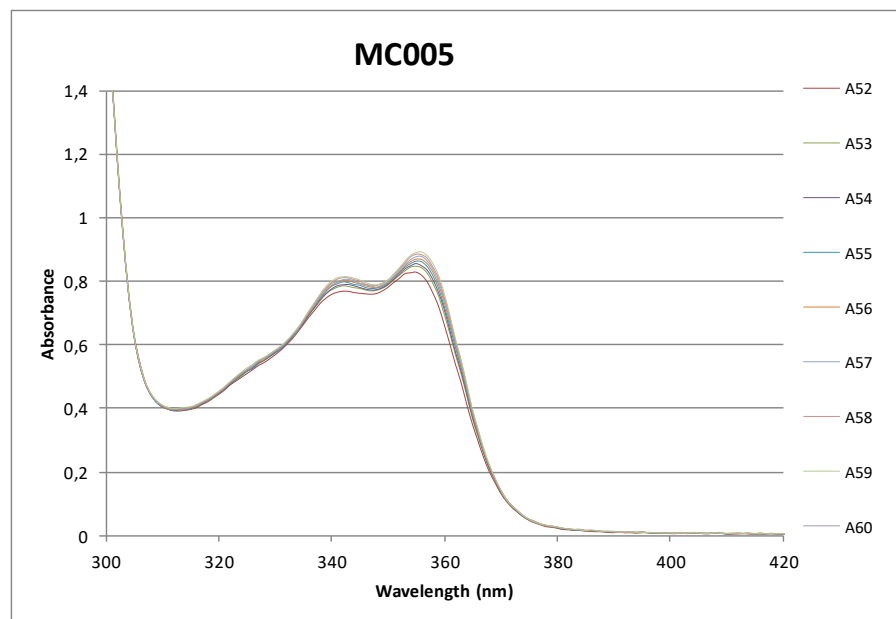
UV-vis spectra of absolute binding affinity measurement for **MC009** with TBA-formate 90.0%:10.0% m/m DMSO-H₂O



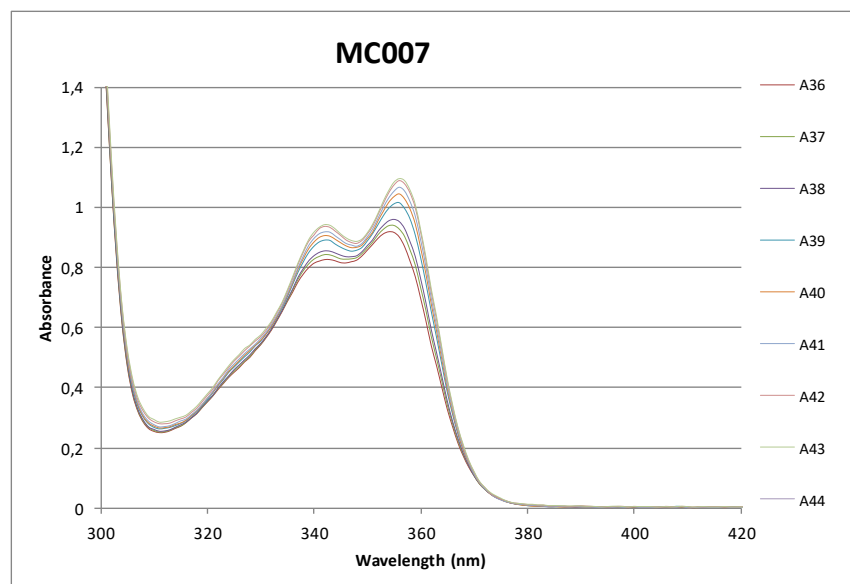
UV-vis spectra of absolute binding affinity measurement for **MC004** with TBA-lactate 90.0%:10.0% m/m DMSO-H₂O



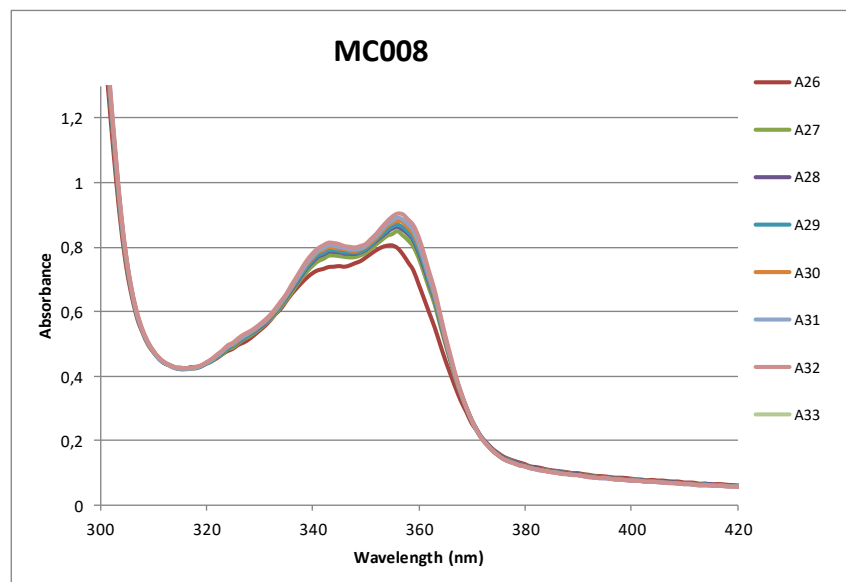
UV-vis spectra of absolute binding affinity measurement for **MC005** with TBA-lactate 90.0%:10.0% m/m DMSO-H₂O



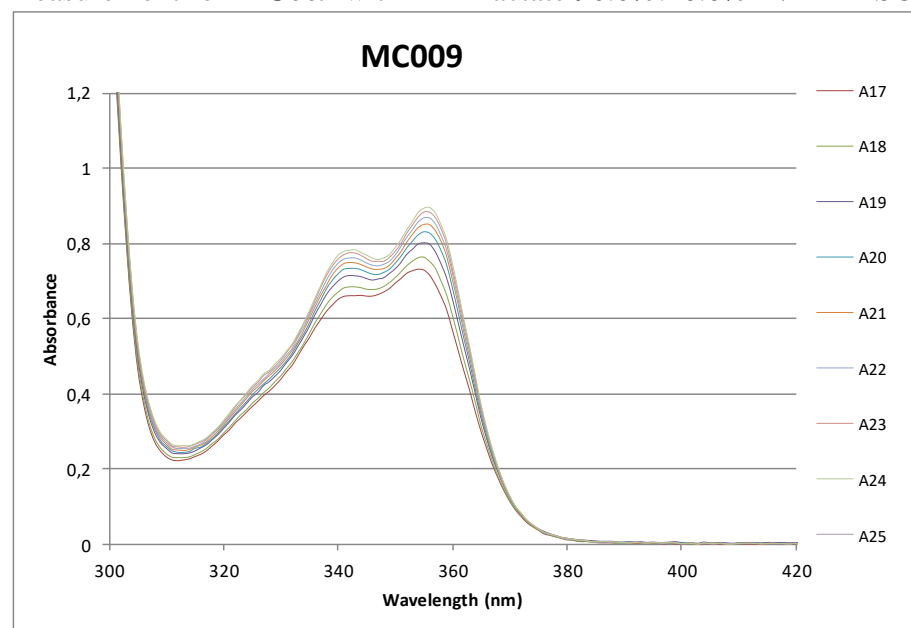
UV-vis spectra of absolute binding affinity measurement for **MC007** with TBA-lactate 90.0%:10.0% m/m DMSO-H₂O



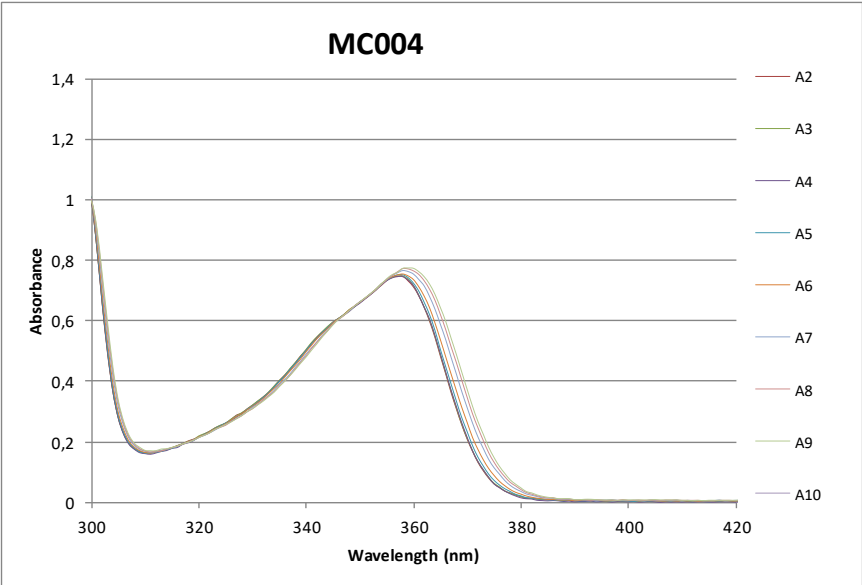
UV-vis spectra of absolute binding affinity measurement for **MC008** with TBA-lactate 90.0%:10.0% m/m DMSO-H₂O



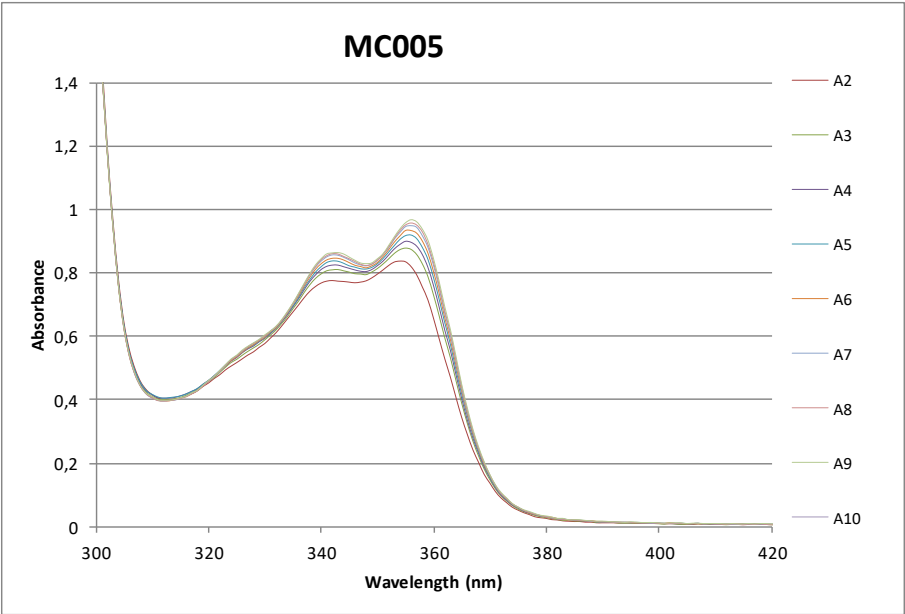
UV-vis spectra of absolute binding affinity measurement for **MC009** with TBA-lactate 90.0%:10.0% m/m DMSO-H₂O



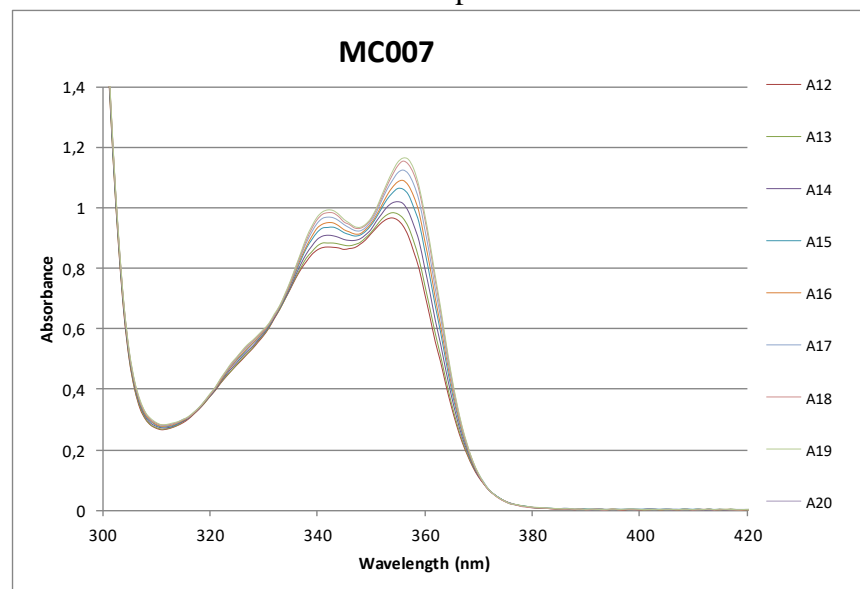
UV-vis spectra of absolute binding affinity measurement for **MC004** with TBA-pivalate 90.0%:10.0% m/m DMSO-H₂O



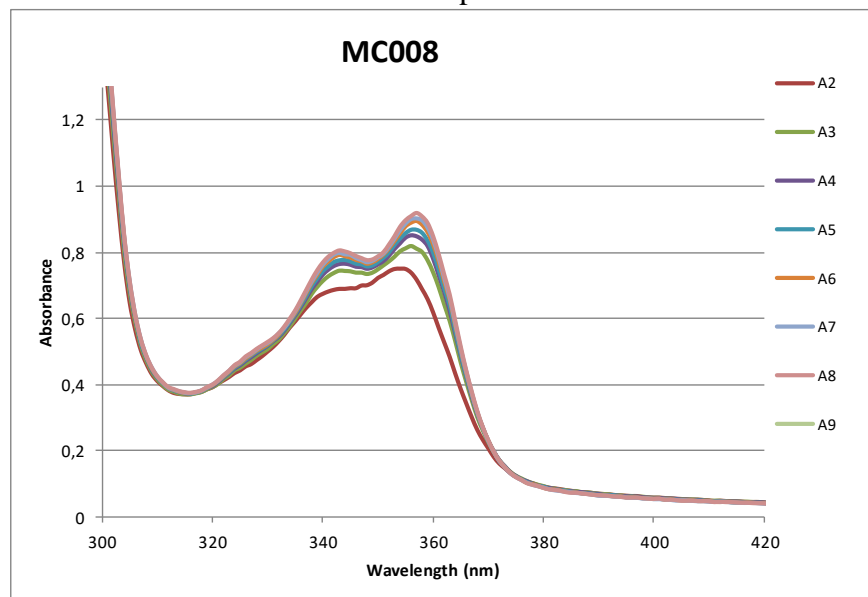
UV-vis spectra of absolute binding affinity measurement for **MC005** with TBA-pivalate 90.0%:10.0% m/m DMSO-H₂O



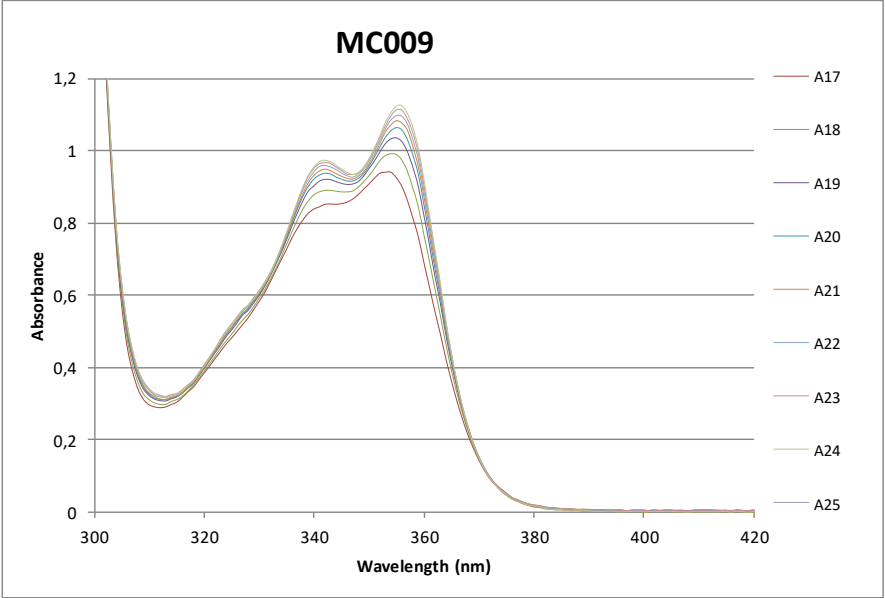
UV-vis spectra of absolute binding affinity measurement for **MC007** with TBA-pivalate 90.0%:10.0% m/m DMSO-H₂O



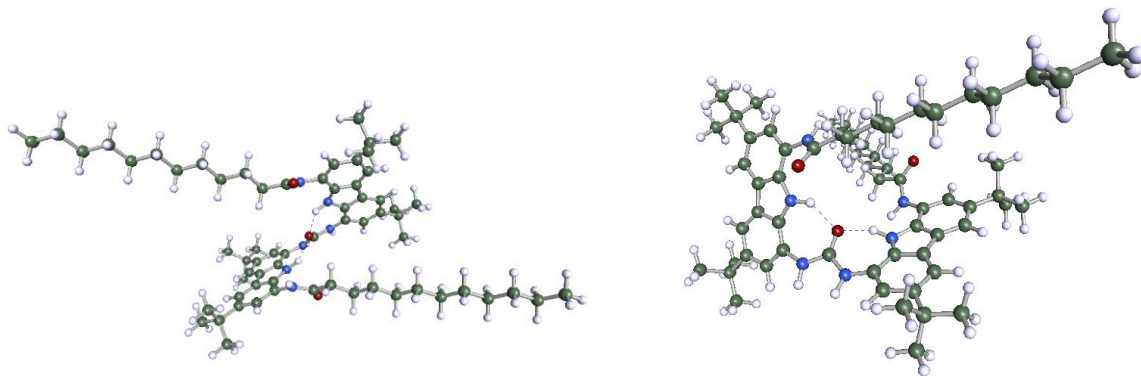
UV-vis spectra of absolute binding affinity measurement for **MC008** with TBA-pivalate 90.0%:10.0% m/m DMSO-H₂O



UV-vis spectra of absolute binding affinity measurement for **MC009** with TBA-pivalate 90.0%:10.0% m/m DMSO-H2O

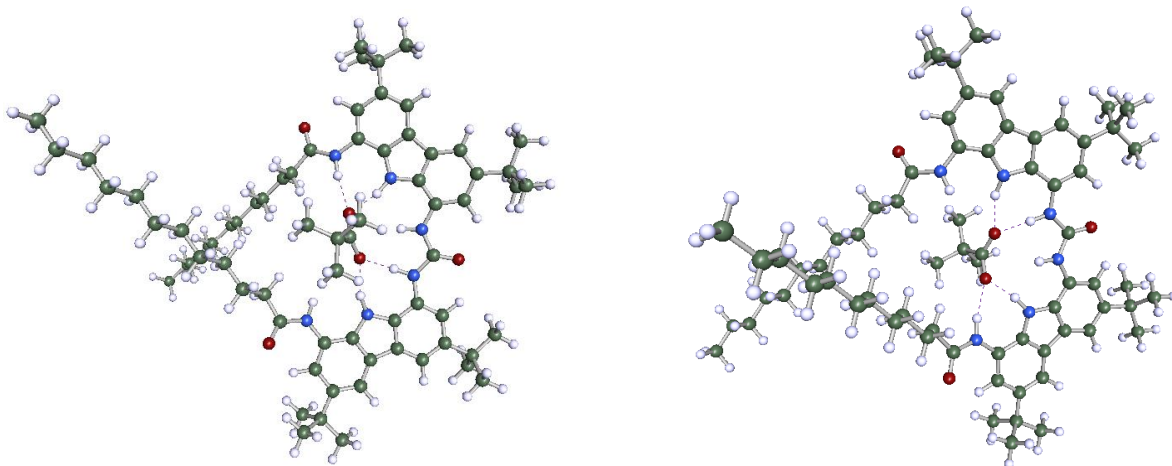


Lowest energy conformation of receptor **CZ016**



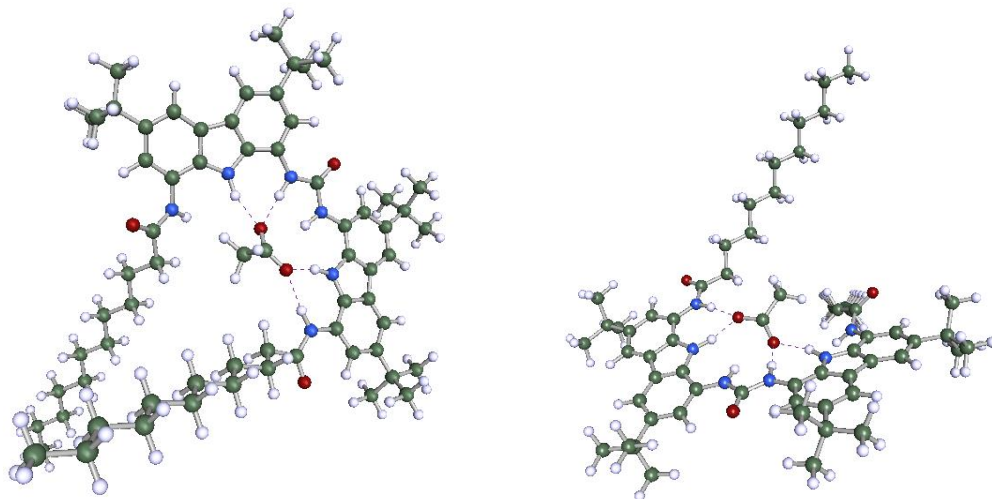
$G = -1939212.08127$ kcal/mol

Lowest energy conformation of receptor **CZ016** with pivalate anion



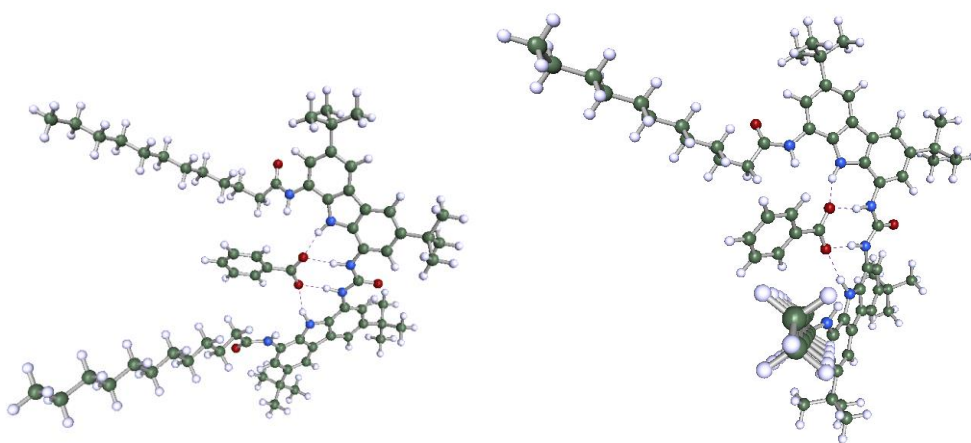
$G = -2156792.28741$ kcal/mol

Lowest energy conformation of receptor **CZ016** with acetate anion



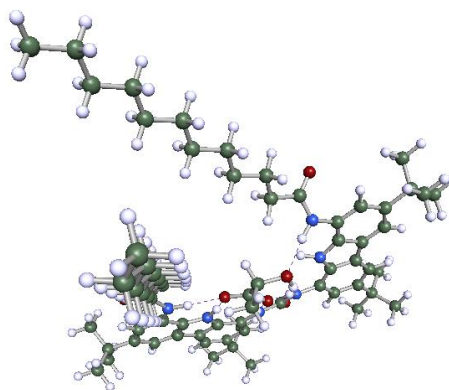
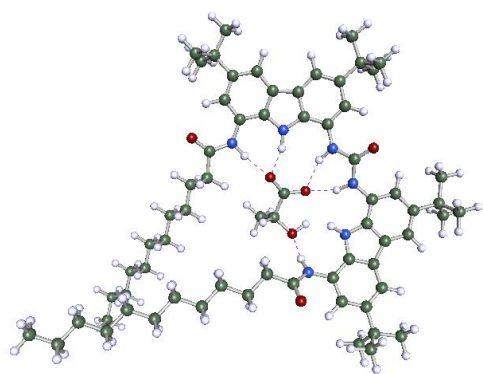
$G = -2082766.14645 \text{ kcal/mol}$

Lowest energy conformation of receptor **CZ016** with benzoate anion



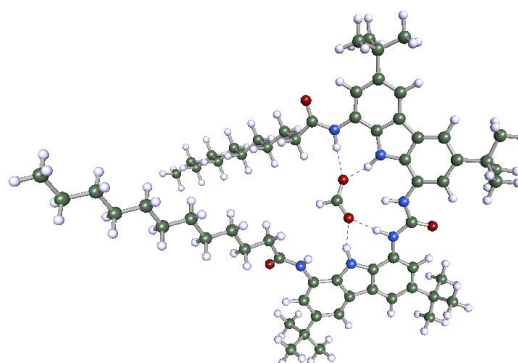
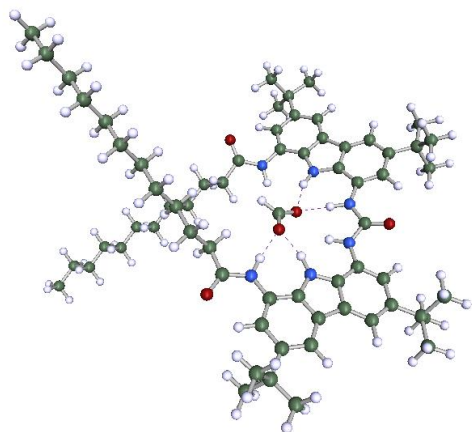
$G = 2203124.50120 \text{ kcal/mol}$

Lowest energy conformation of receptor **CZ016** with lactate anion



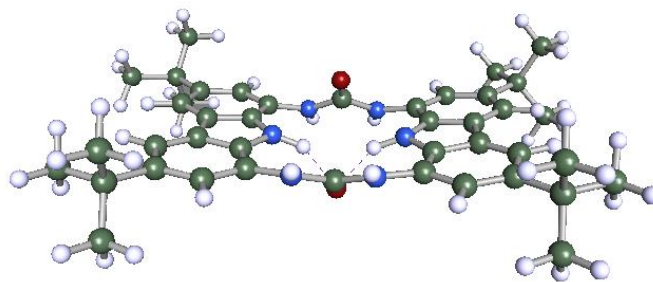
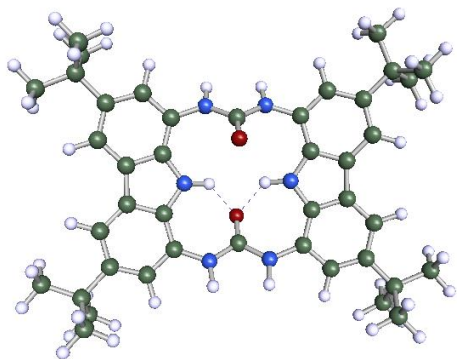
$G = -2154669.70262$ kcal/mol

Lowest energy conformation of receptor **CZ016** with formate anion



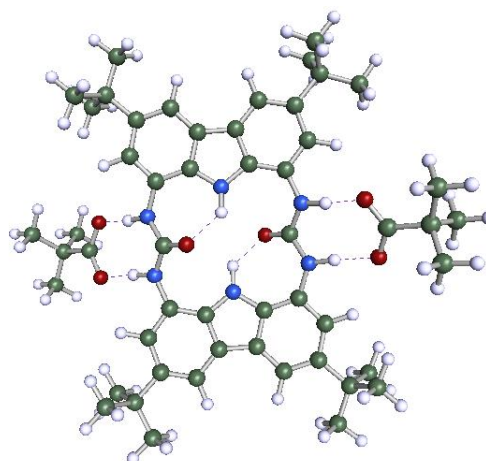
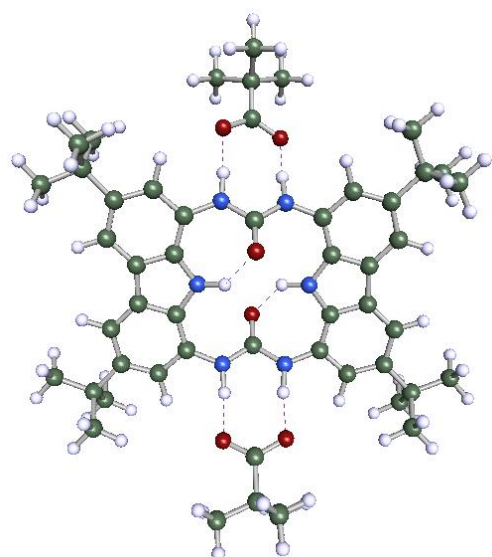
$G = -2058084.08381$ kcal/mol

Lowest energy conformation of receptor **MC001**



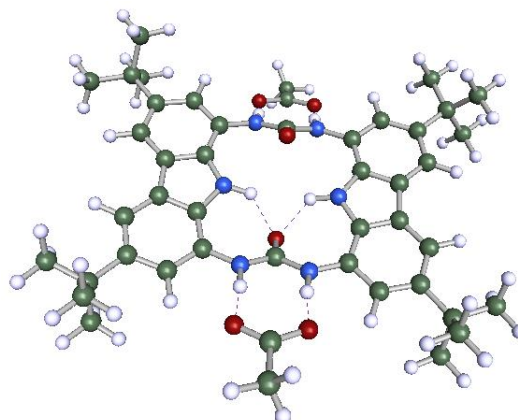
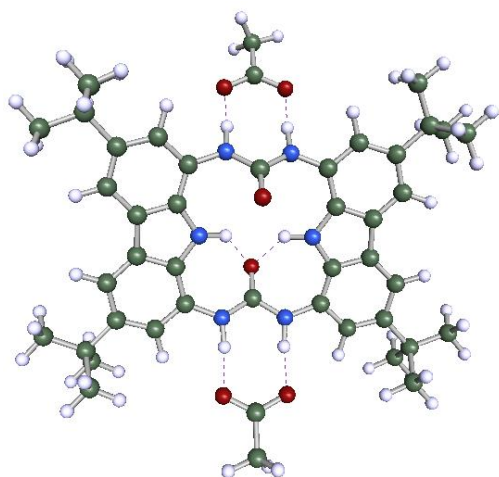
$G = -1324377.24934$ kcal/mol

Lowest energy conformation of receptor **MC001** with pivalate anion



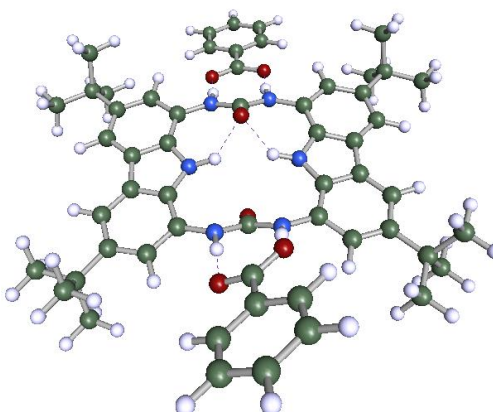
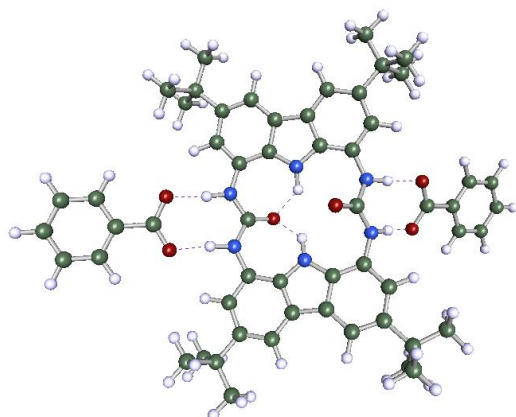
$G = -1759540.52493$ kcal/mol

Lowest energy conformation of receptor **MC001** with acetate anion



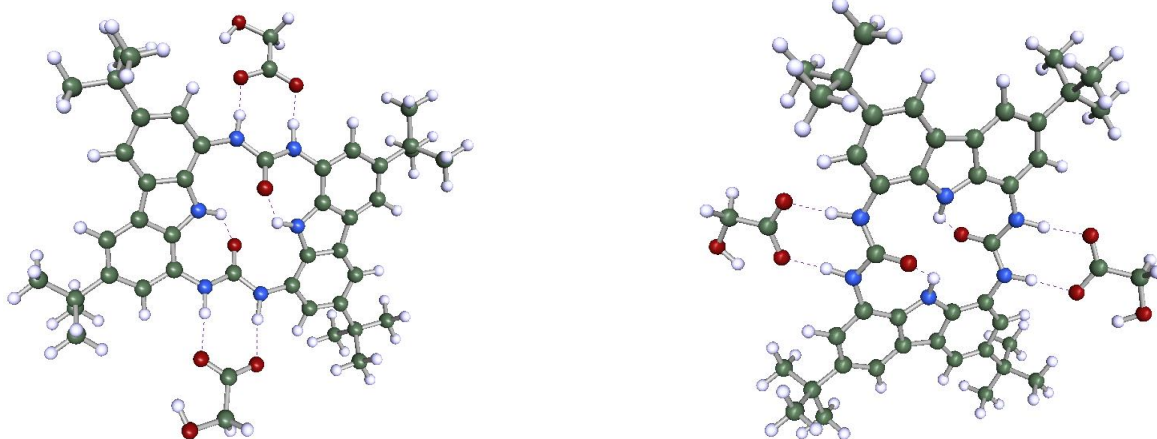
$G = -1611476.38251$ kcal/mol

Lowest energy conformation of receptor **MC001** with benzoate anion



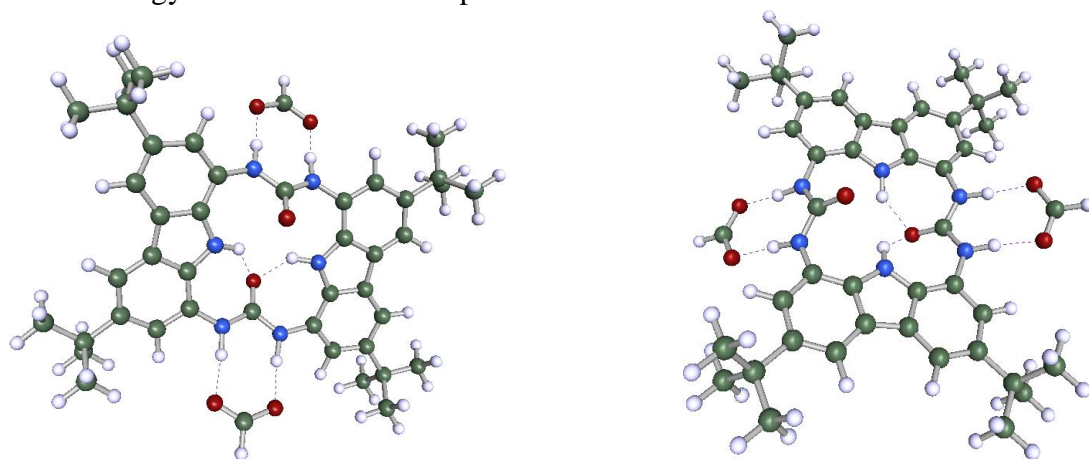
$G = -1852203.69392$ kcal/mol

Lowest energy conformation of receptor **MC001** with lactate anion



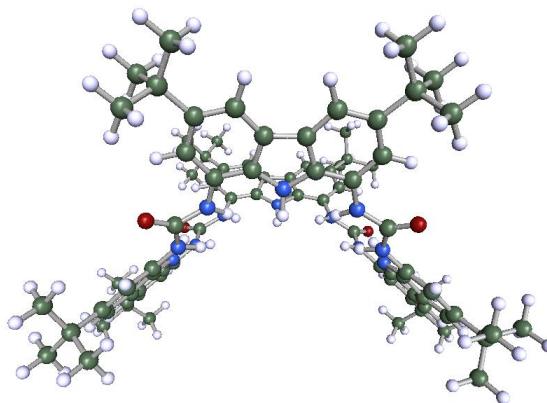
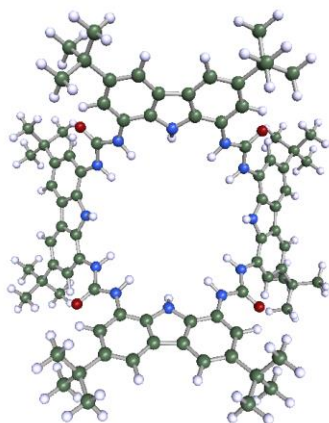
$G = -1705929.89393$ kcal/mol

Lowest energy conformation of receptor **MC001** with formate anion



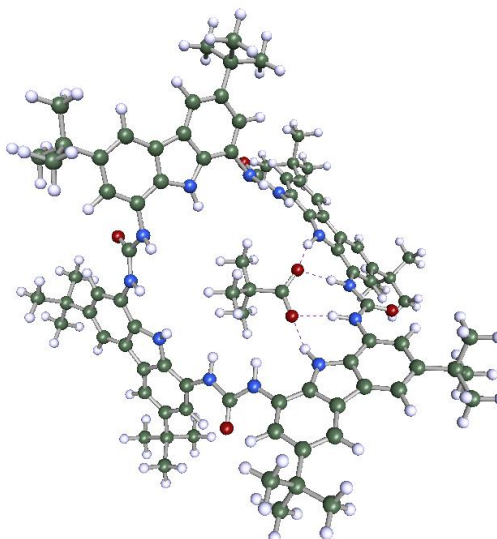
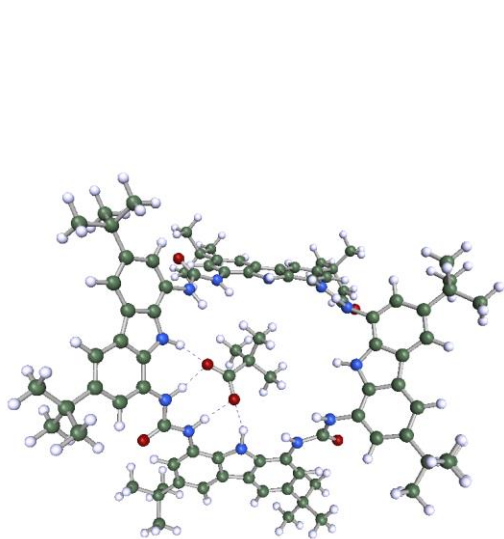
$G = -1562111.52768$ kcal/mol

Lowest energy conformation of receptor **MC002**



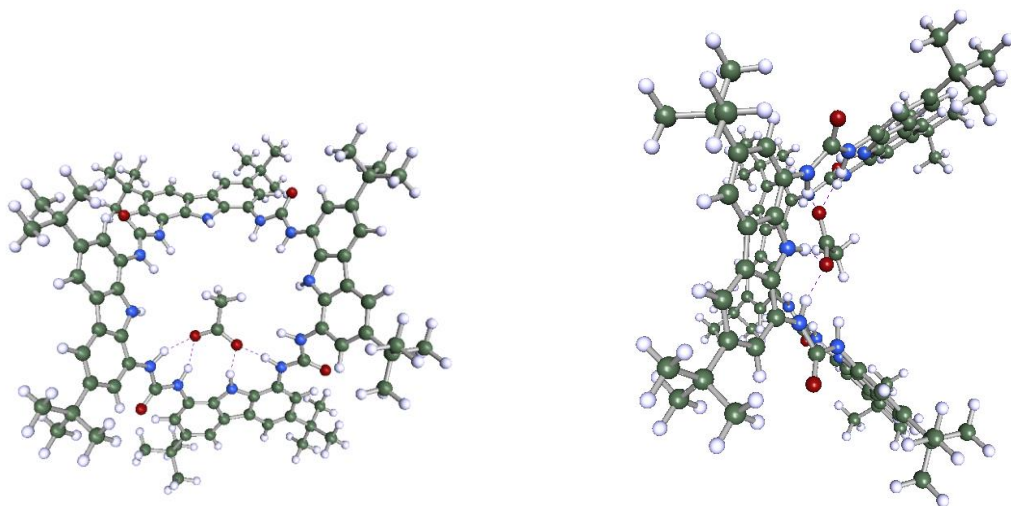
$G = -2648766.27994$ kcal/mol

Lowest energy conformation of receptor **MC002** with pivalate anion



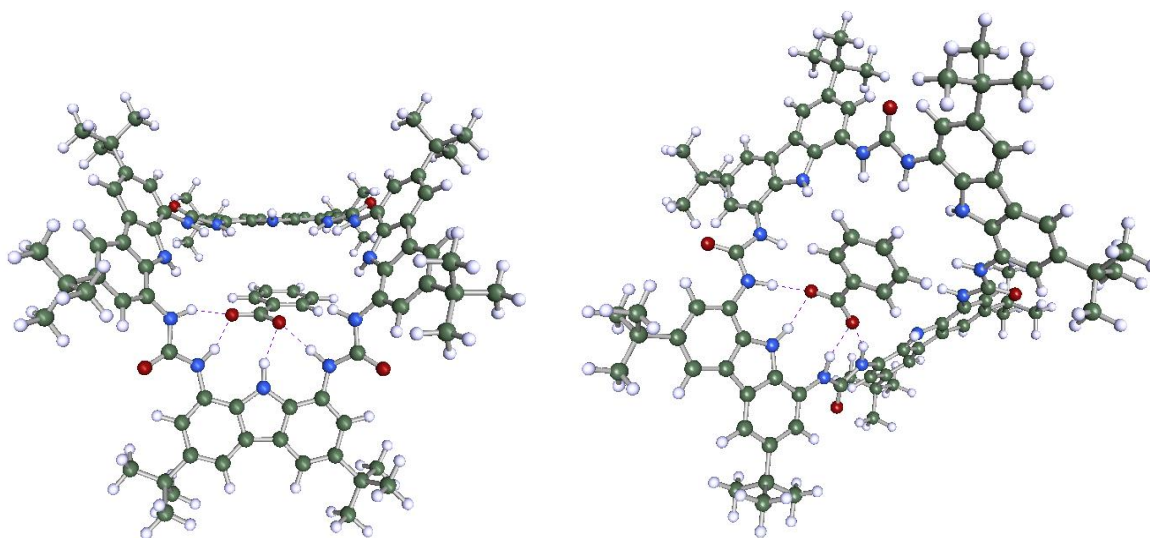
$G = -2866337.50211$ kcal/mol

Lowest energy conformation of receptor **MC002** with acetate anion



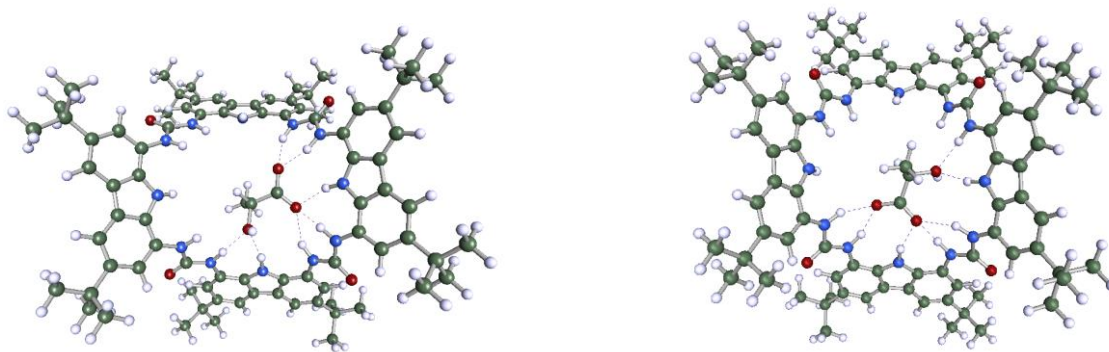
$G = -2792310.16571$ kcal/mol

Lowest energy conformation of receptor **MC002** with benzoate anion



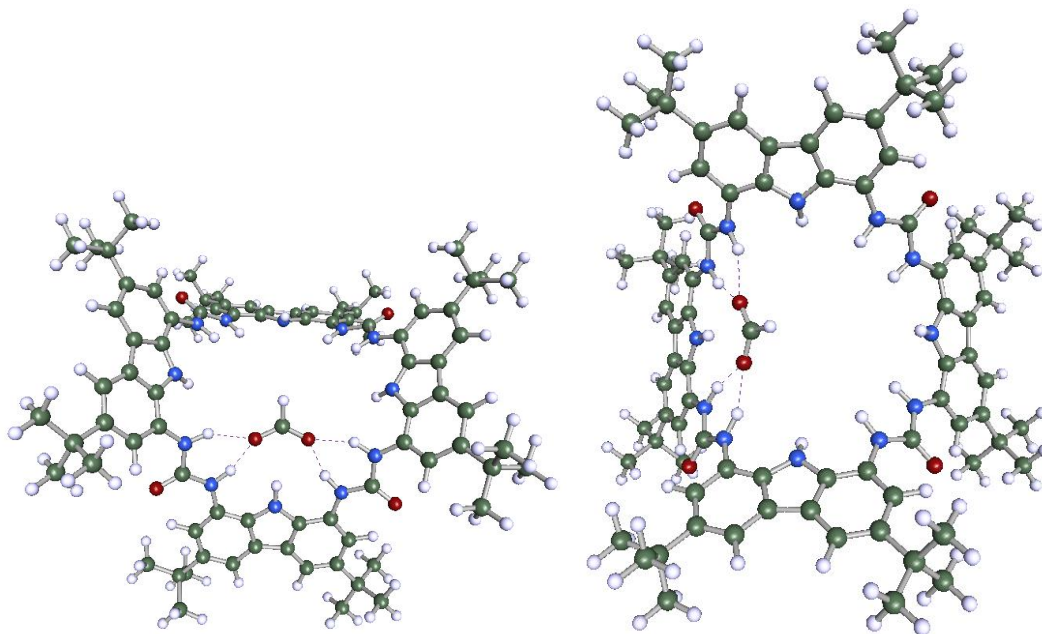
$G = -2912671.14577$ kcal/mol

Lowest energy conformation of receptor **MC002** with lactate anion



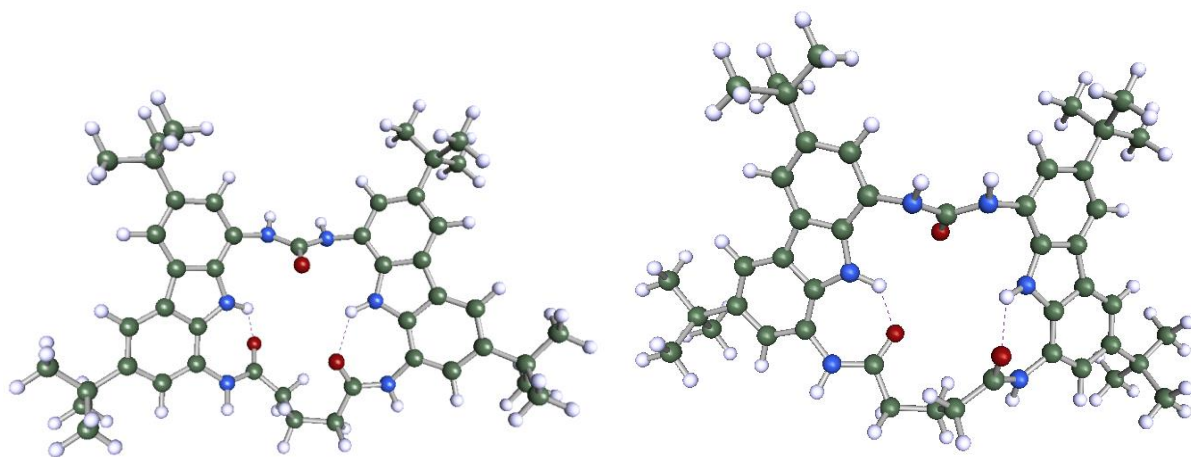
$G = -2864220.05839$ kcal/mol

Lowest energy conformation of receptor **MC002** with formate anion



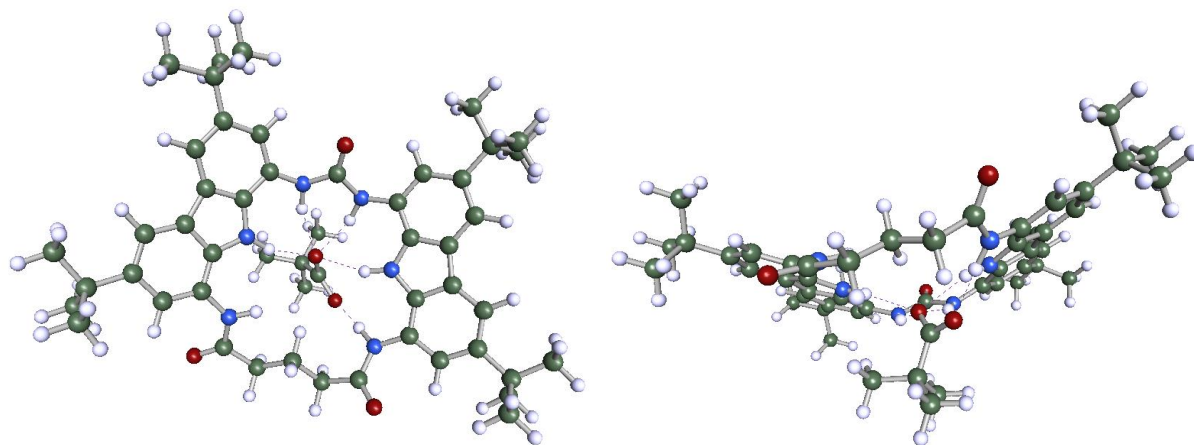
$G = -2767634.54569$ kcal/mol

Lowest energy conformation of receptor **MC003**



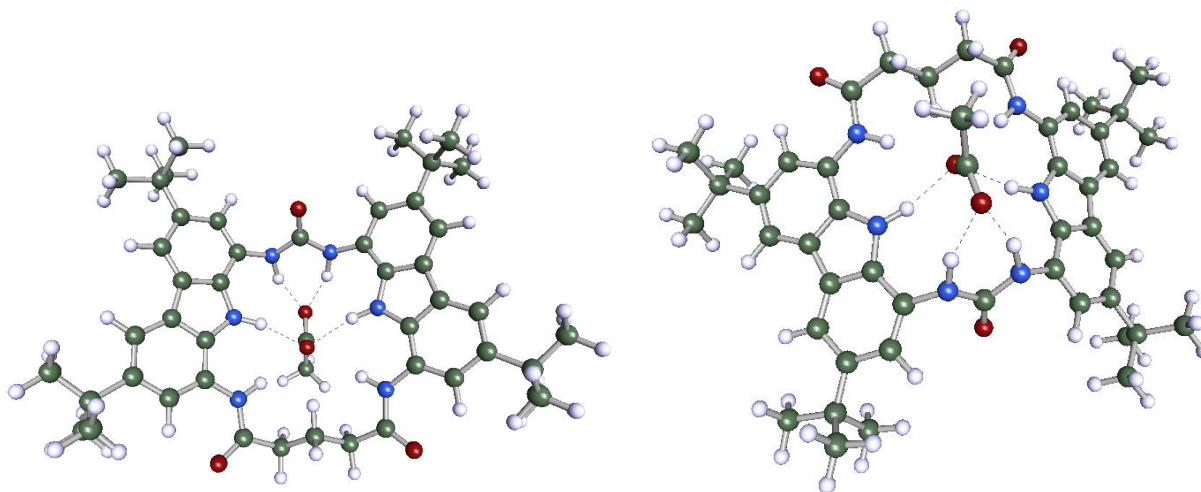
$G = -1469566.91734$ kcal/mol

Lowest energy conformation of receptor **MC003** with pivalate anion



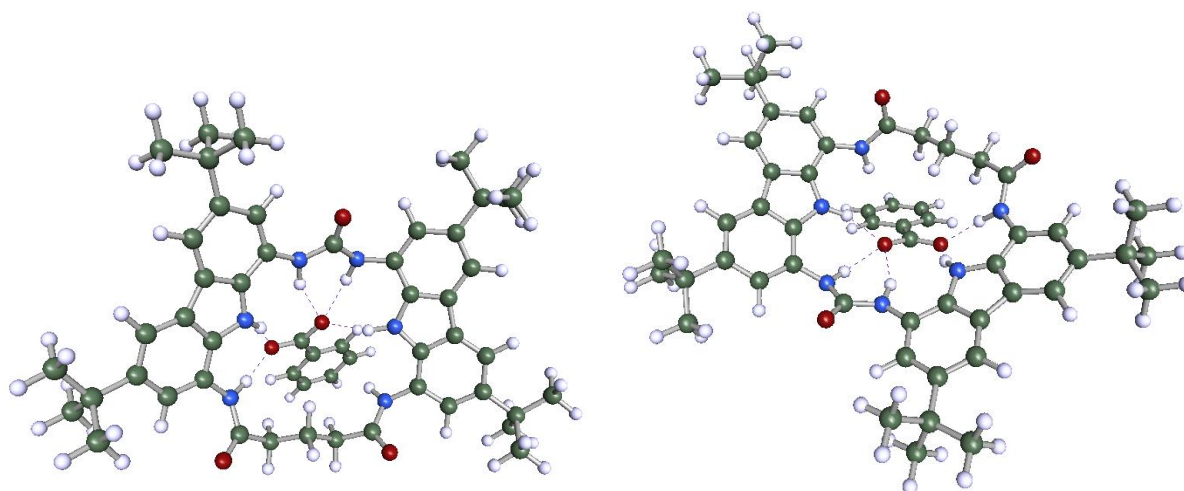
$G = -1687151.33703$ kcal/mol

Lowest energy conformation of receptor **MC003** with acetate anion



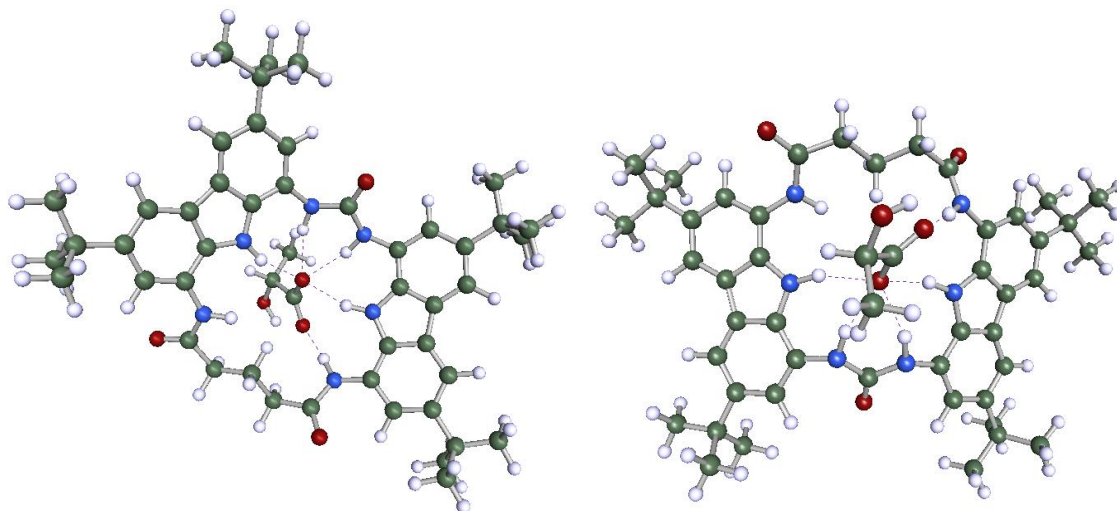
$G = -1613118.82814 \text{ kcal/mol}$

Lowest energy conformation of receptor **MC003** with benzoate anion



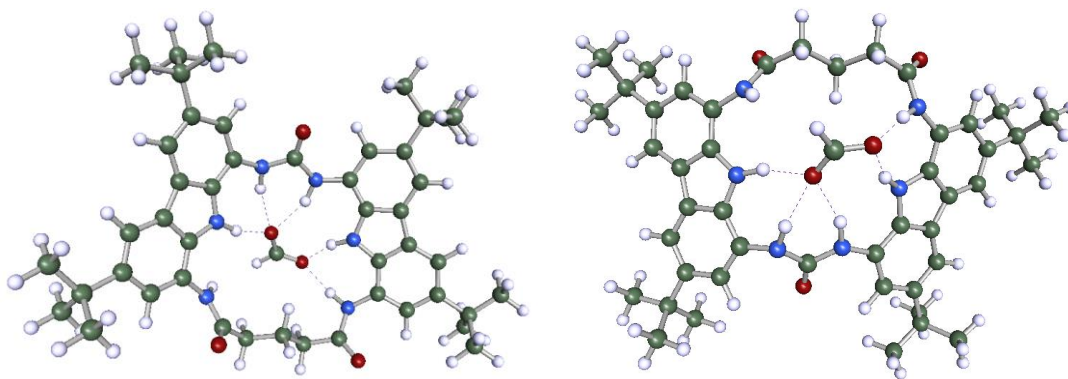
$G = -1733483.80492 \text{ kcal/mol}$

Lowest energy conformation of receptor **MC003** with lactate anion



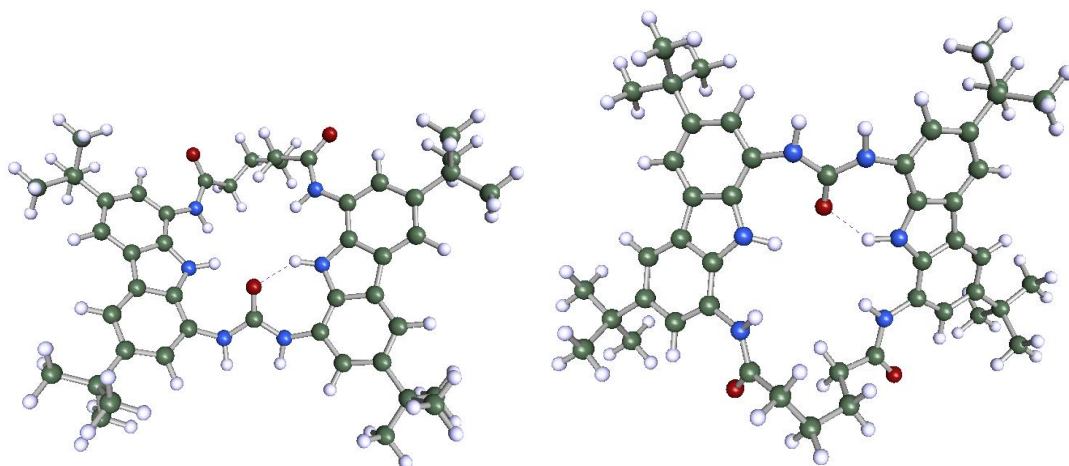
$G = -1685024.93247$ kcal/mol

Lowest energy conformation of receptor **MC003** with formate anion



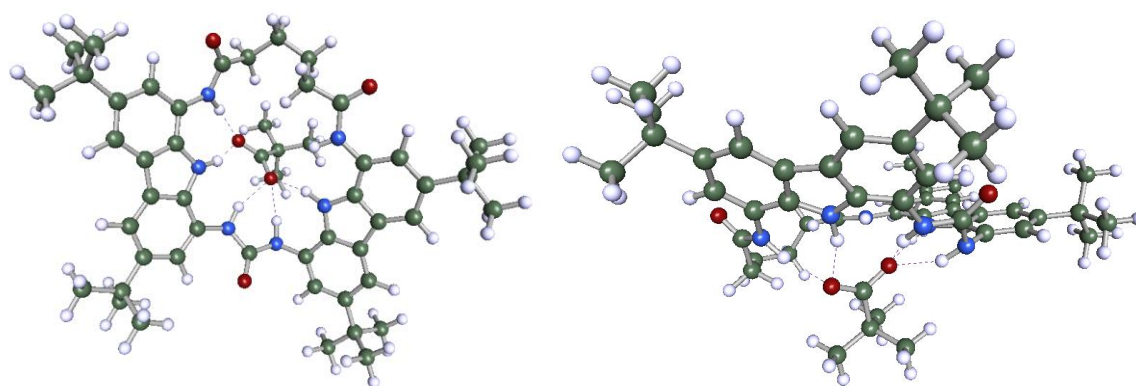
$G = -1588438.70245$ kcal/mol

Lowest energy conformation of receptor **MC004**



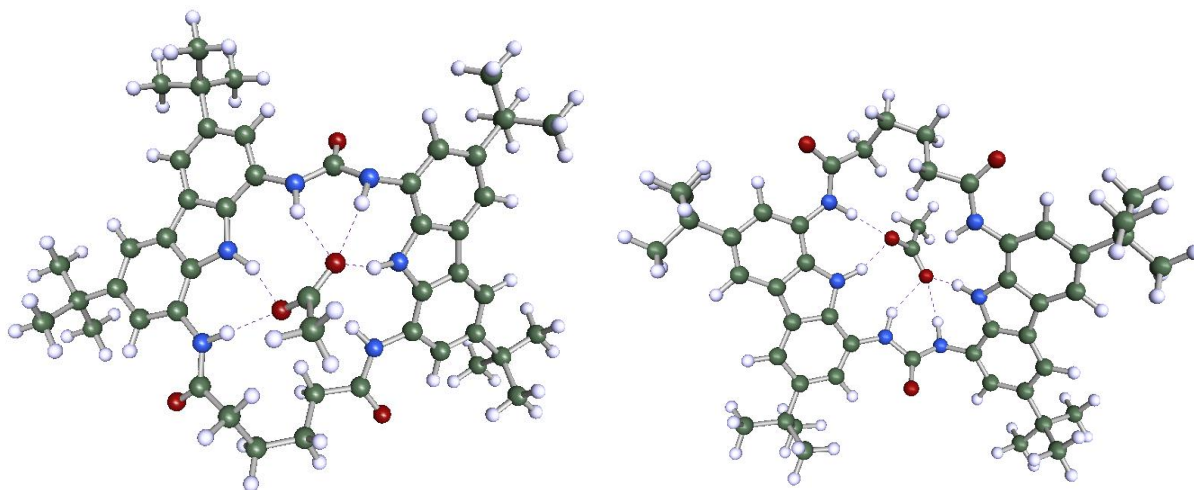
$G = -1494248.55193$ kcal/mol

Lowest energy conformation of receptor **MC004** with pivalate anion



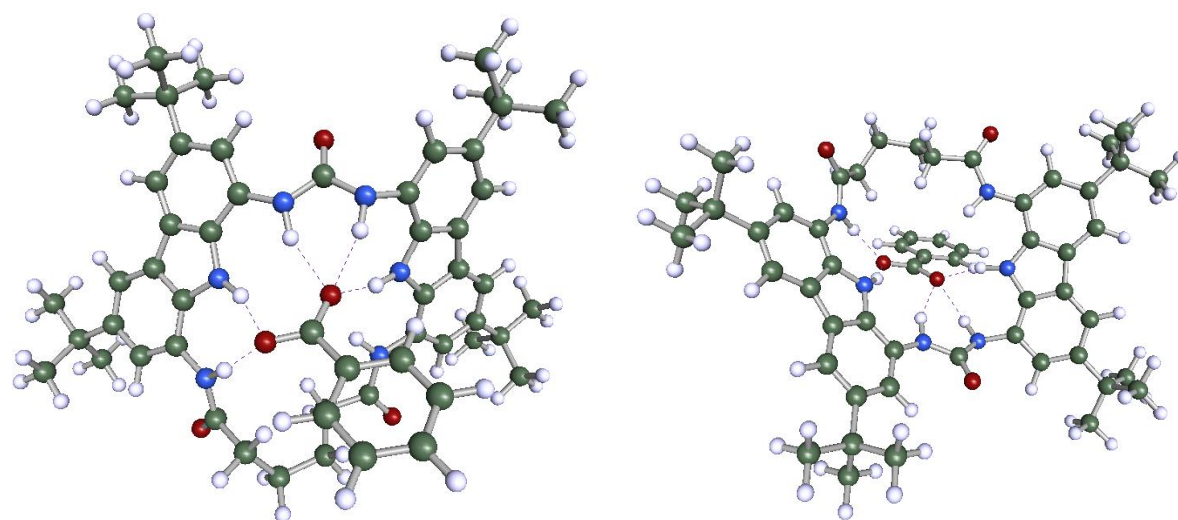
$G = -1711829.70872$ kcal/mol

Lowest energy conformation of receptor **MC004** with acetate anion



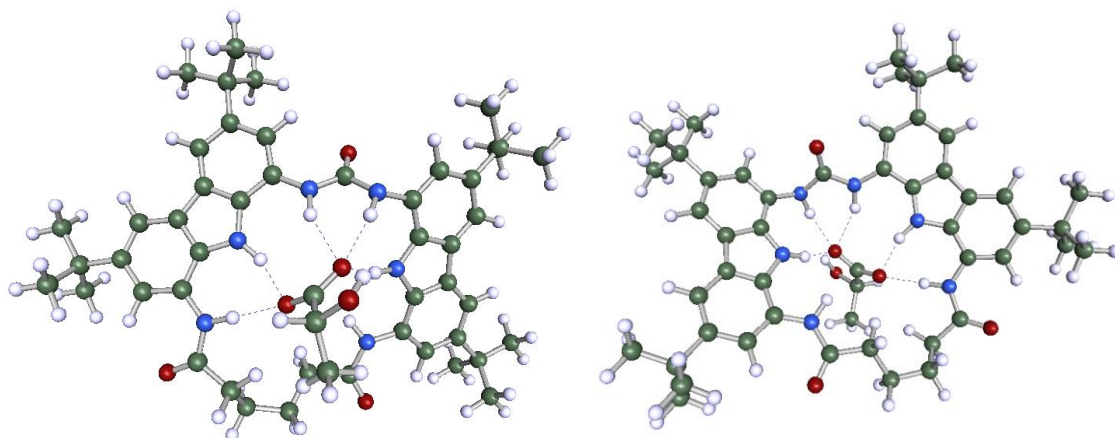
$G = -1637796.77666 \text{ kcal/mol}$

Lowest energy conformation of receptor **MC004** with benzoate anion



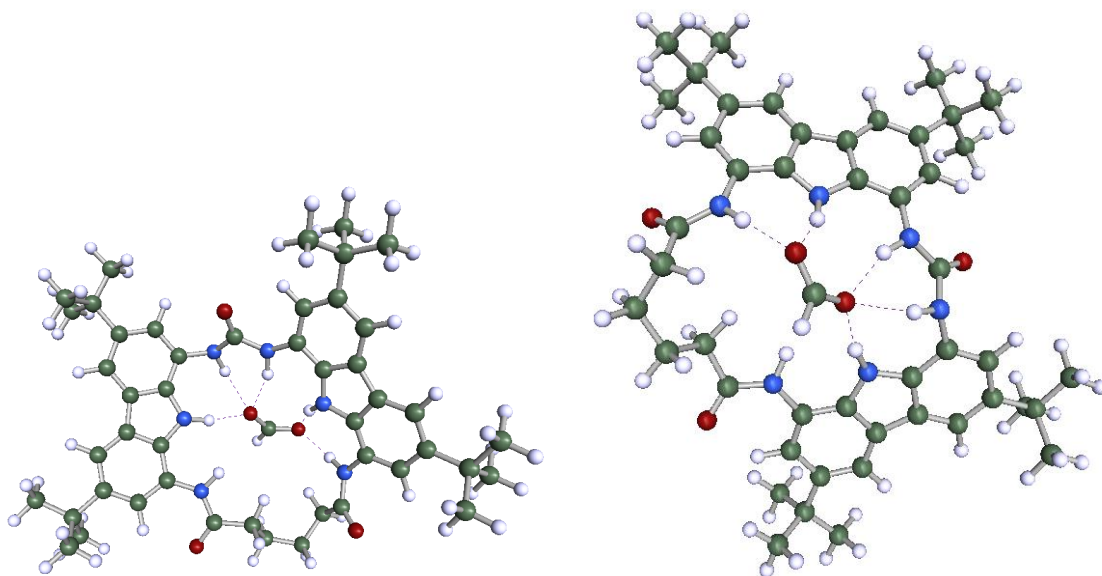
$G = -1758160.92459 \text{ kcal/mol}$

Lowest energy conformation of receptor **MC004** with lactate anion



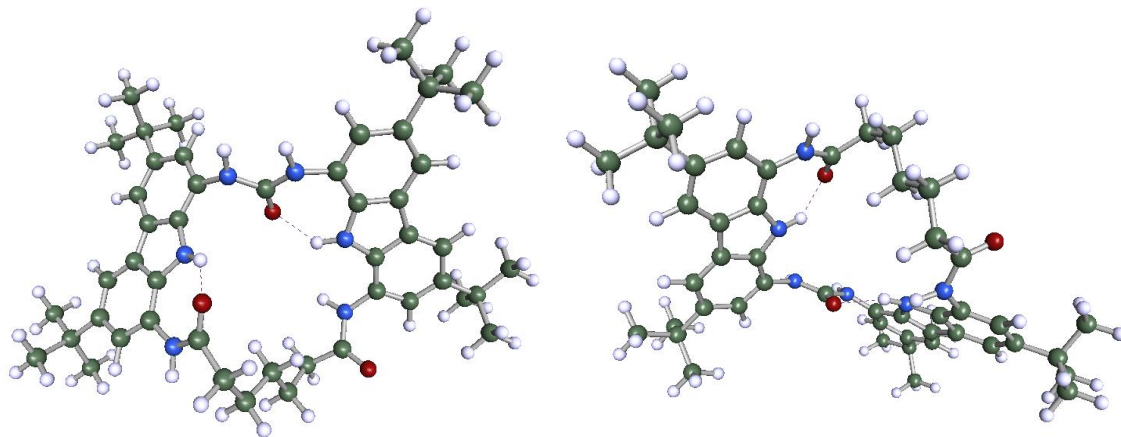
$G = -1709700.31149$ kcal/mol

Lowest energy conformation of receptor **MC004** with formate anion



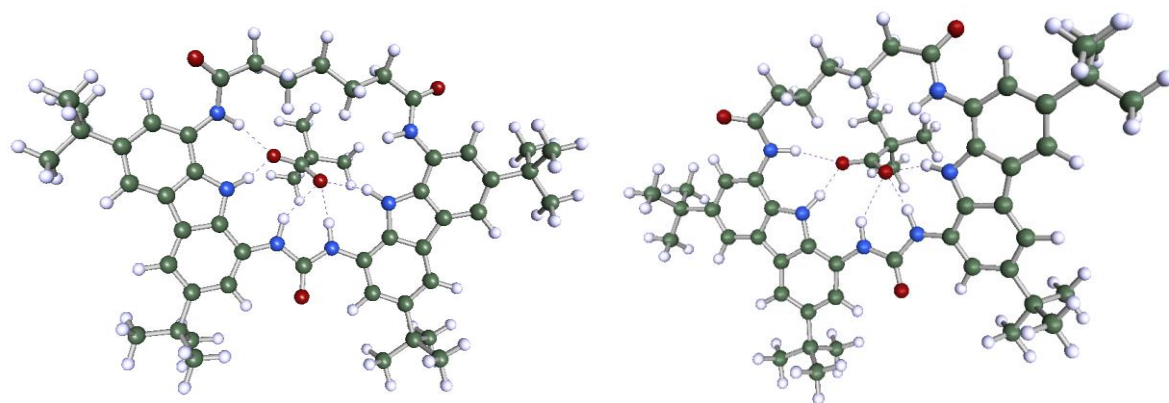
$G = -1613115.44806$ kcal/mol

Lowest energy conformation of receptor **MC005**



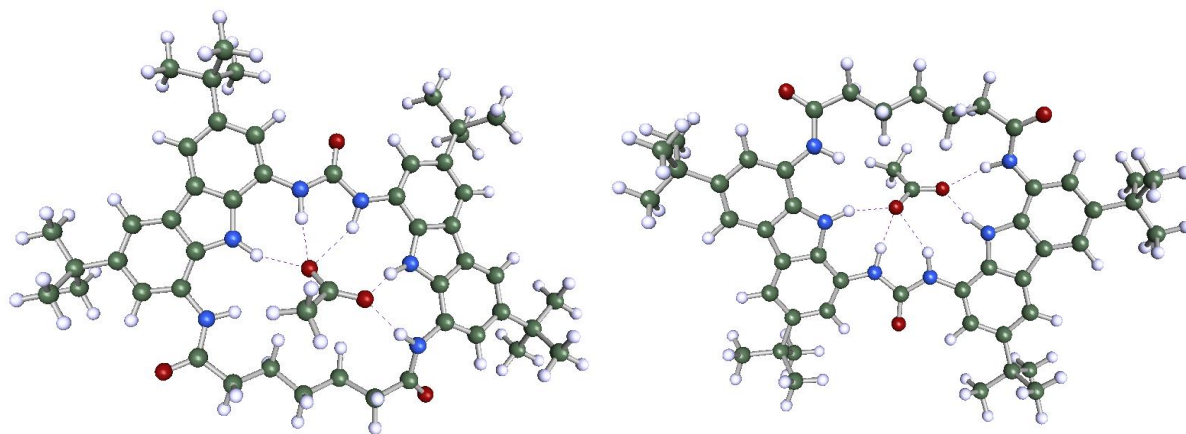
$G = -1518926.98220$ kcal/mol

Lowest energy conformation of receptor **MC005** with pivalate anion



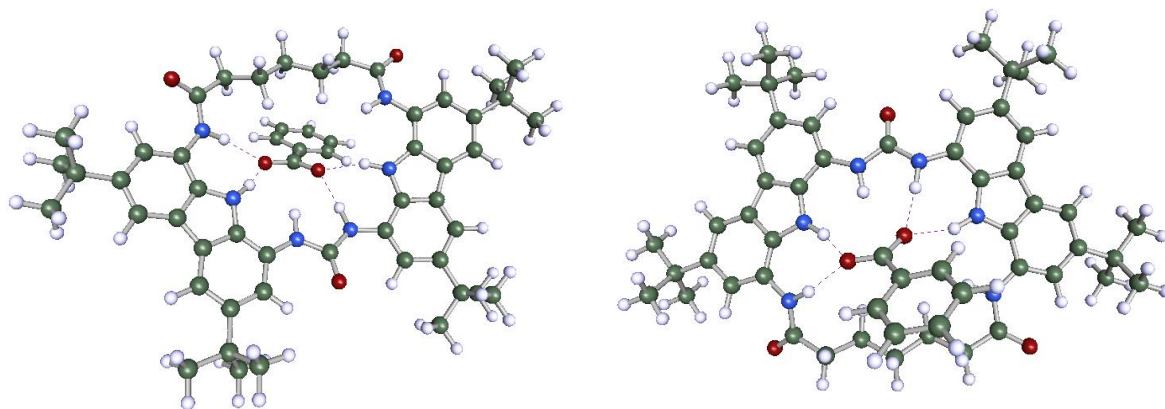
$G = -1736508.43110$ kcal/mol

Lowest energy conformation of receptor **MC005** with acetate anion



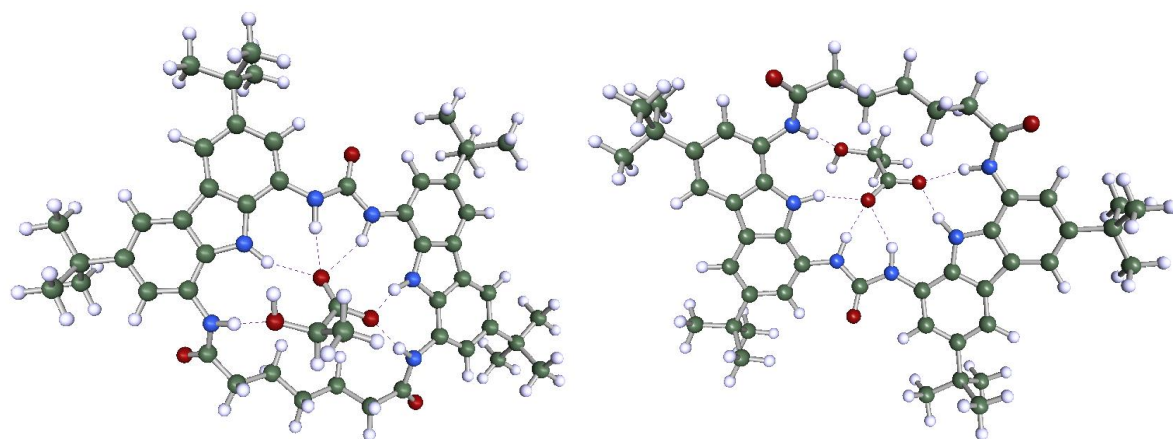
$G = -1662477.72110$ kcal/mol

Lowest energy conformation of receptor **MC005** with benzoate anion



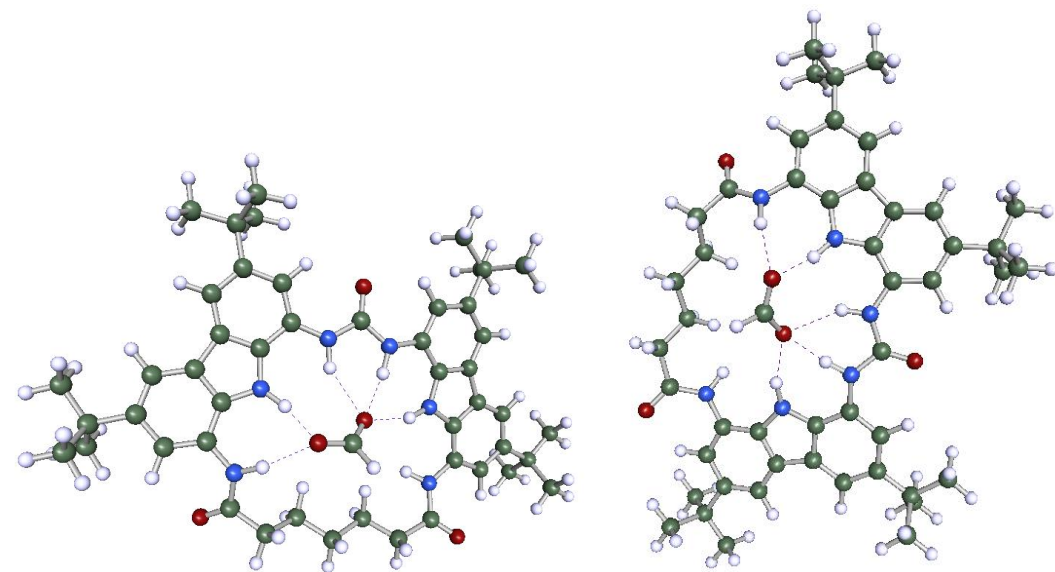
$G = -1782840.25461$ kcal/mol

Lowest energy conformation of receptor **MC005** with lactate anion



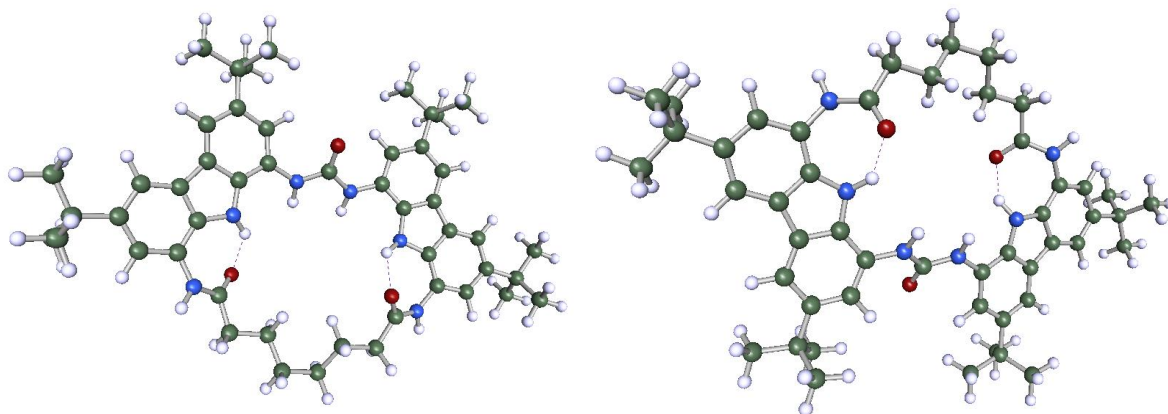
$G = -1734386.24267$ kcal/mol

Lowest energy conformation of receptor **MC005** with formate anion



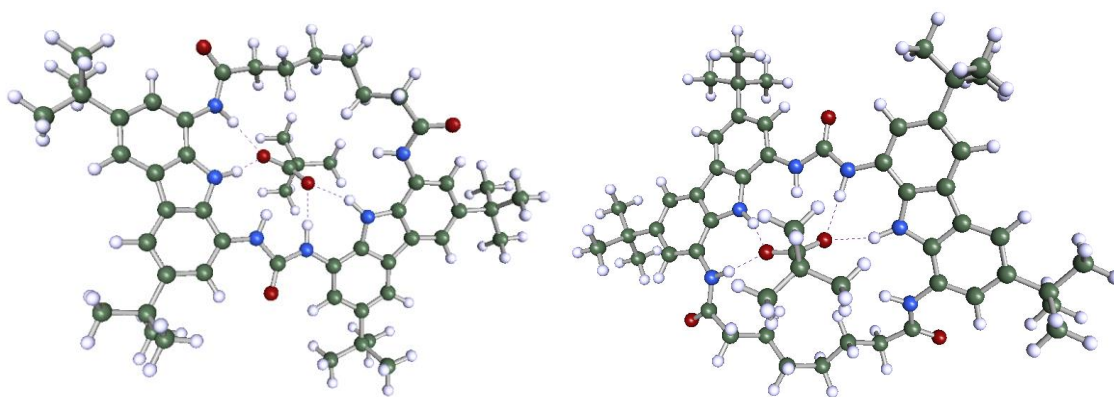
$G = -1637795.20174$ kcal/mol

Lowest energy conformation of receptor **MC006**



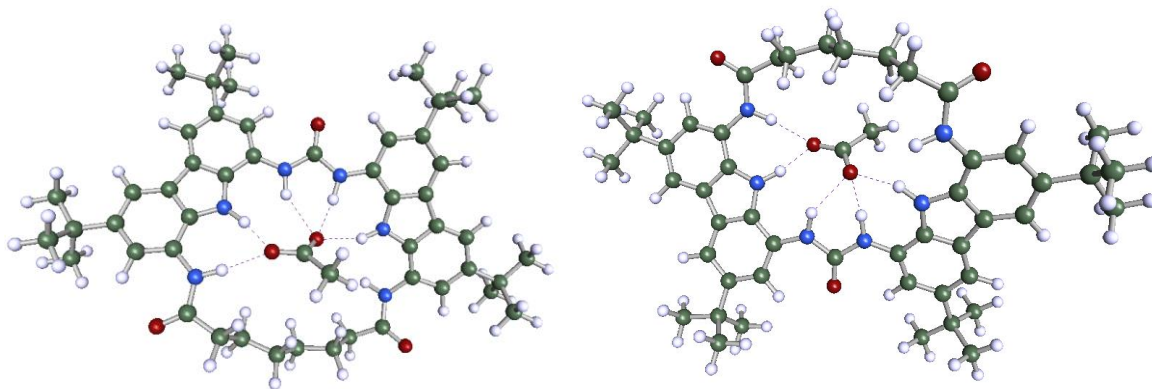
$G = -1543604.89540$ kcal/mol

Lowest energy conformation of receptor **MC006** with pivalate anion



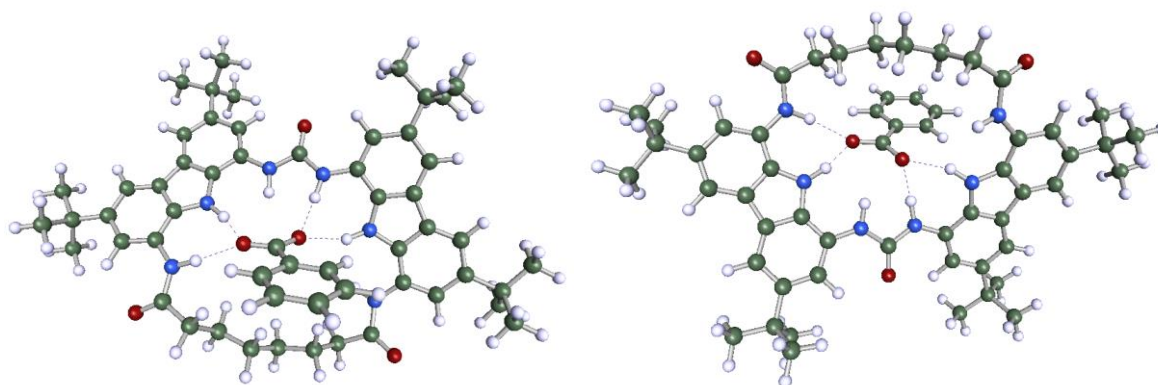
$G = -1761185.90669$ kcal/mol

Lowest energy conformation of receptor **MC006** with acetate anion



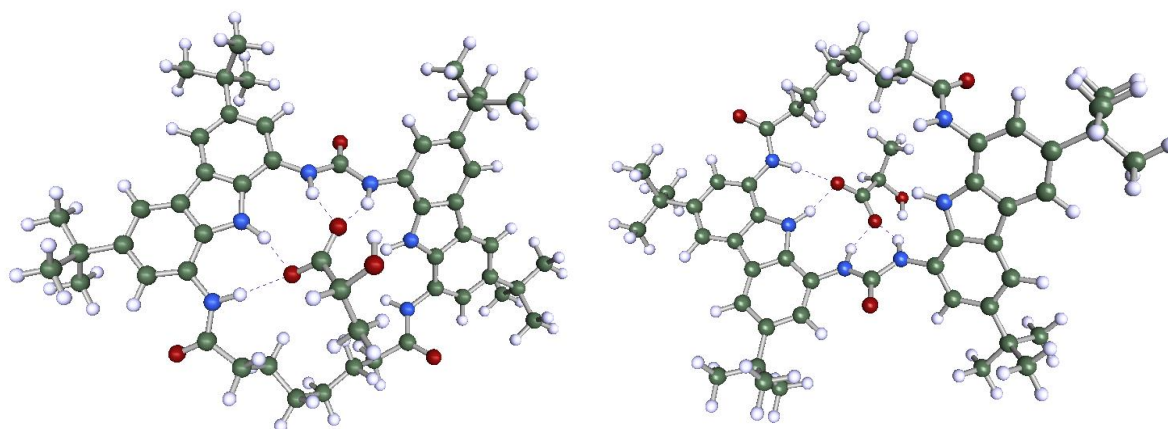
$G = -1687156.27049$ kcal/mol

Lowest energy conformation of receptor **MC006** with benzoate anion



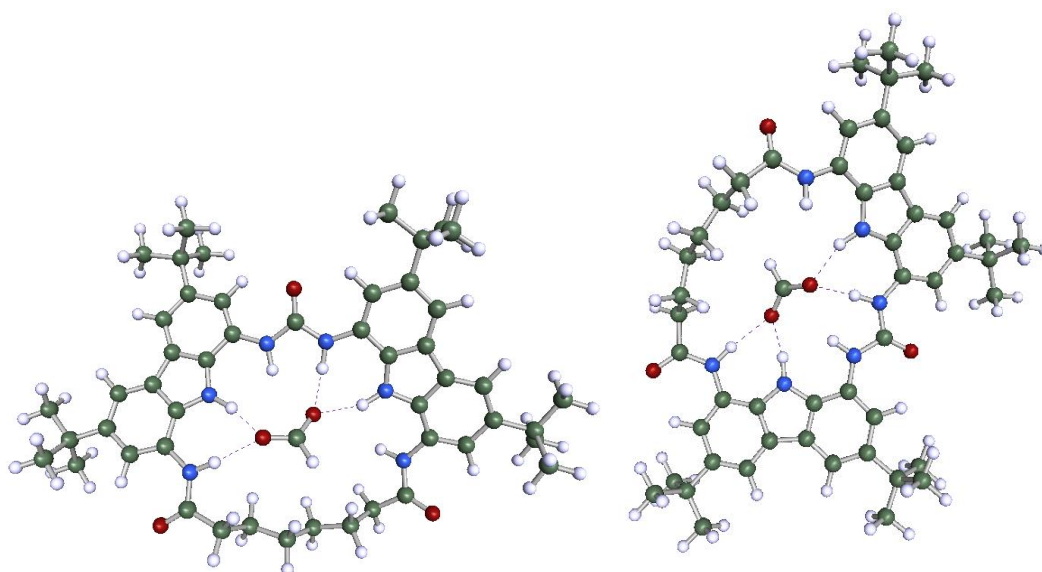
$G = -1807517.68972$ kcal/mol

Lowest energy conformation of receptor **MC006** with lactate anion



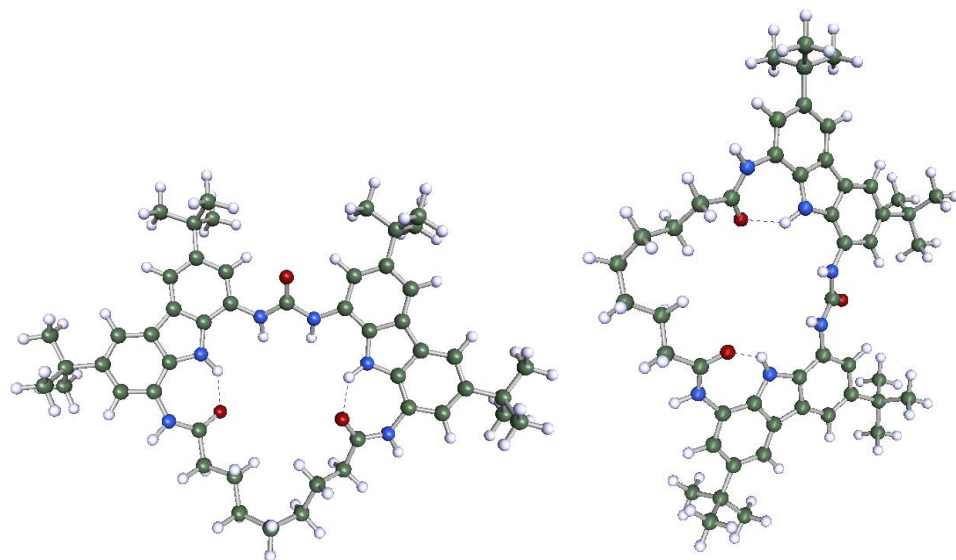
$G = -1759060.05069$ kcal/mol

Lowest energy conformation of receptor **MC006** with formate anion



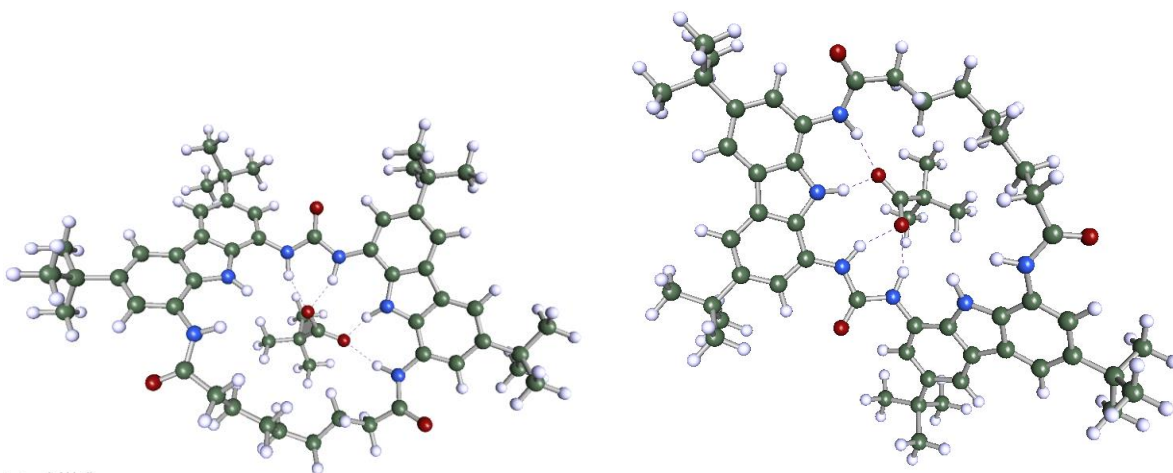
$G = -1662474.70525$ kcal/mol

Lowest energy conformation of receptor **MC007**



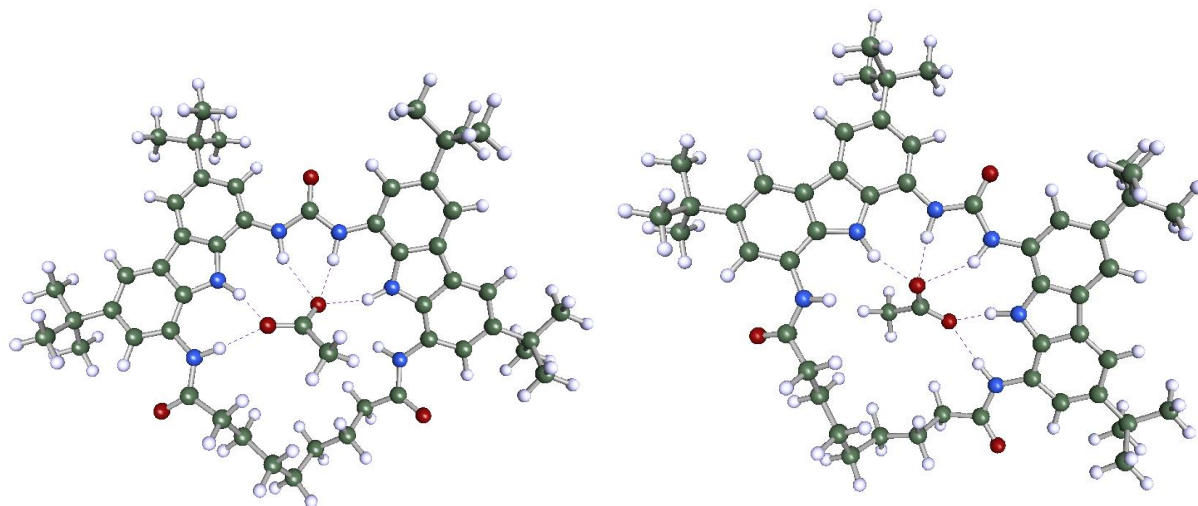
$G = -1568283.77596$ kcal/mol

Lowest energy conformation of receptor **MC007** with pivalate anion



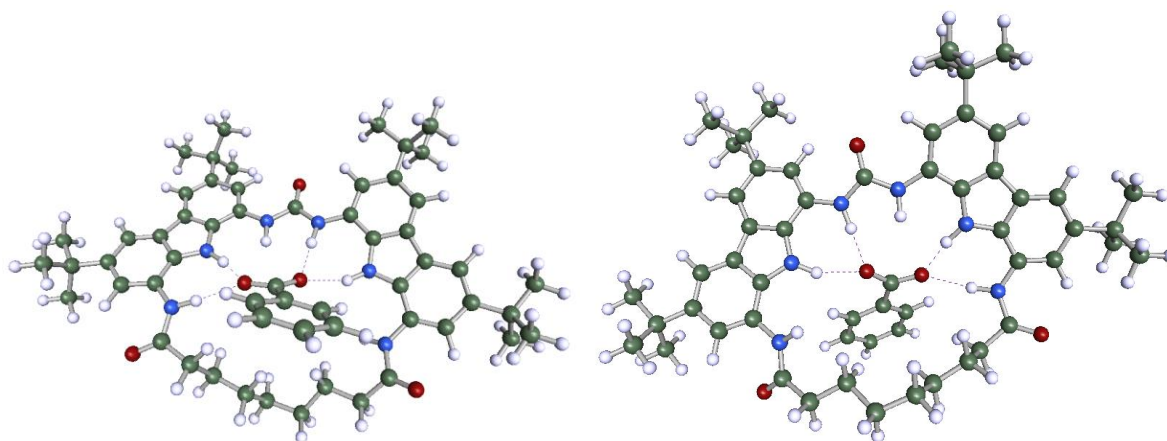
$G = -1785862.89606$ kcal/mol

Lowest energy conformation of receptor **MC007** with acetate anion



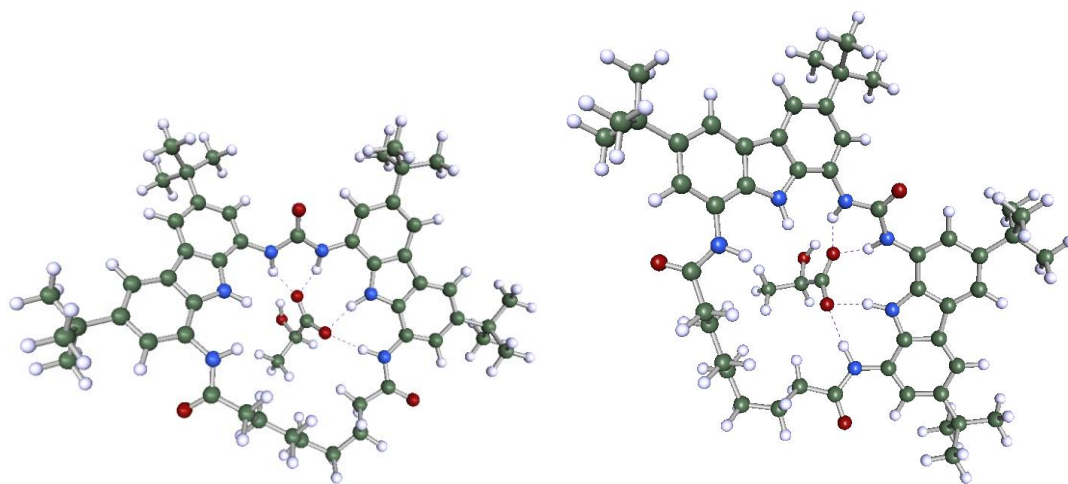
$G = -1711837.51767$ kcal/mol

Lowest energy conformation of receptor **MC007** with benzoate anion



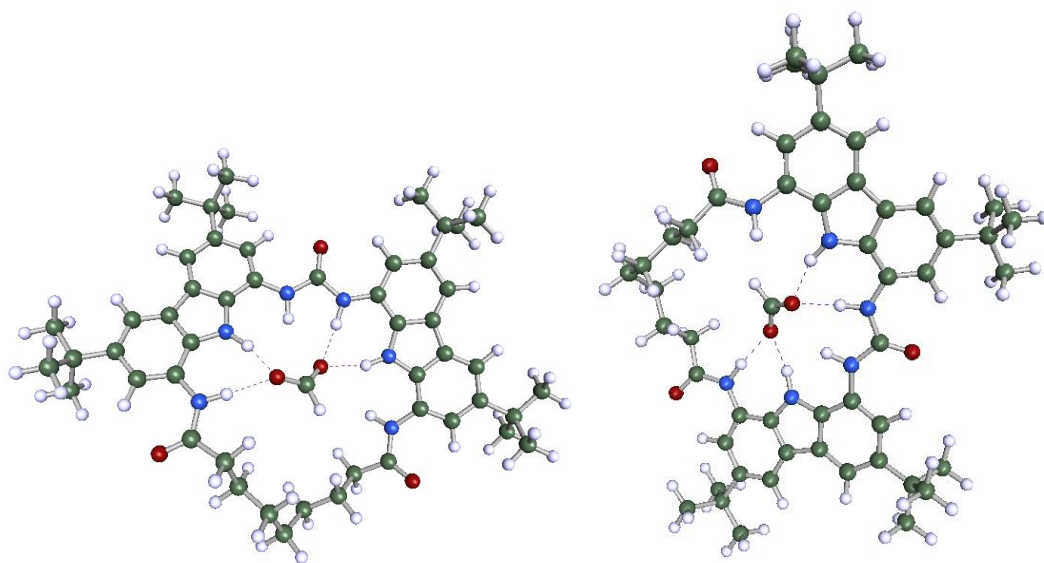
$G = -1832195.75582$ kcal/mol

Lowest energy conformation of receptor **MC007** with lactate anion



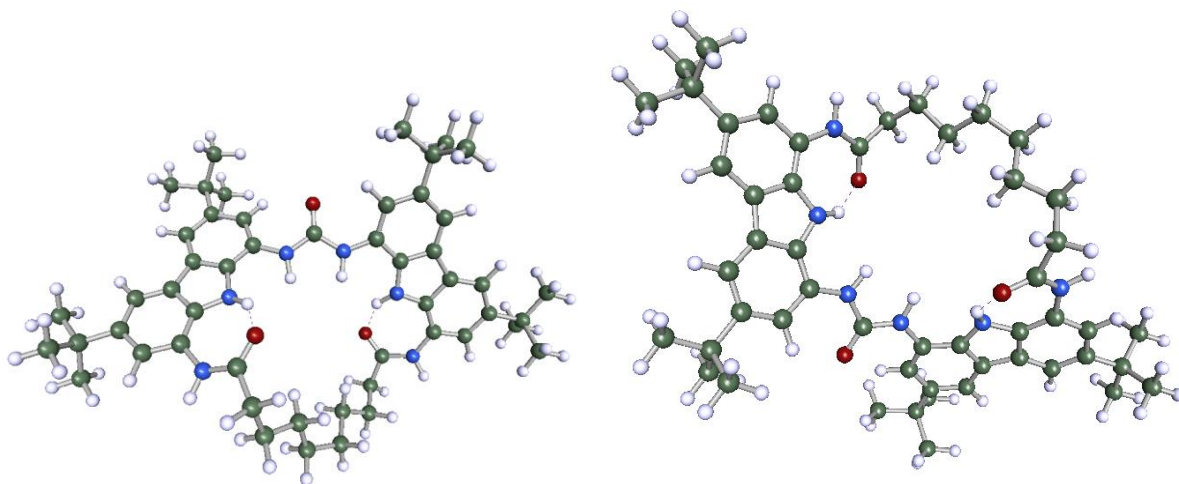
$G = -1783739.48423$ kcal/mol

Lowest energy conformation of receptor **MC007** with formate anion



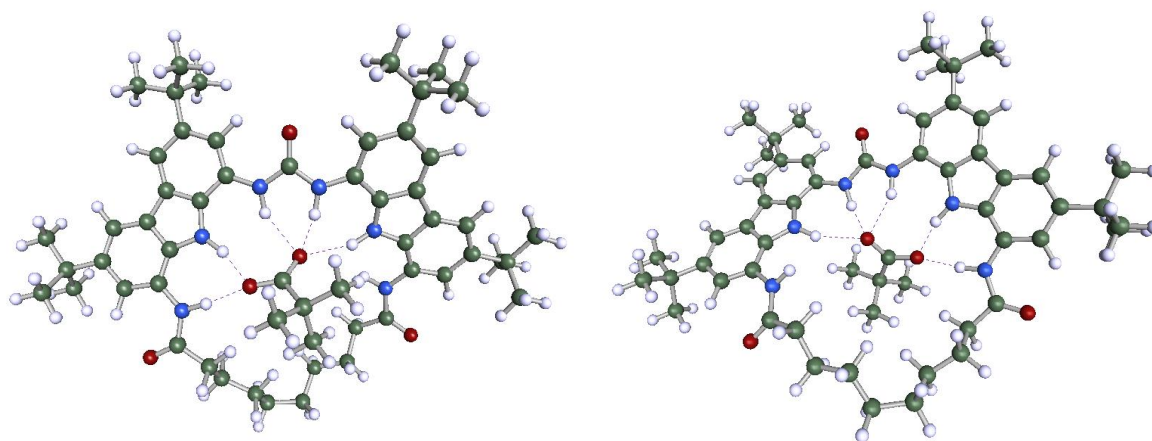
$G = -1687155.70120$ kcal/mol

Lowest energy conformation of receptor **MC008**



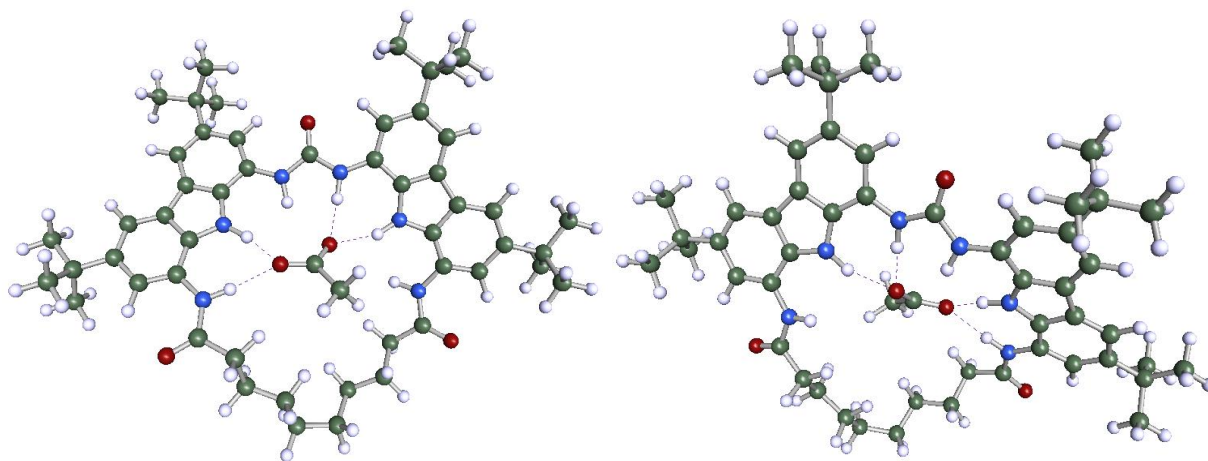
$G = -1592960.38942$ kcal/mol

Lowest energy conformation of receptor **MC008** with pivalate anion



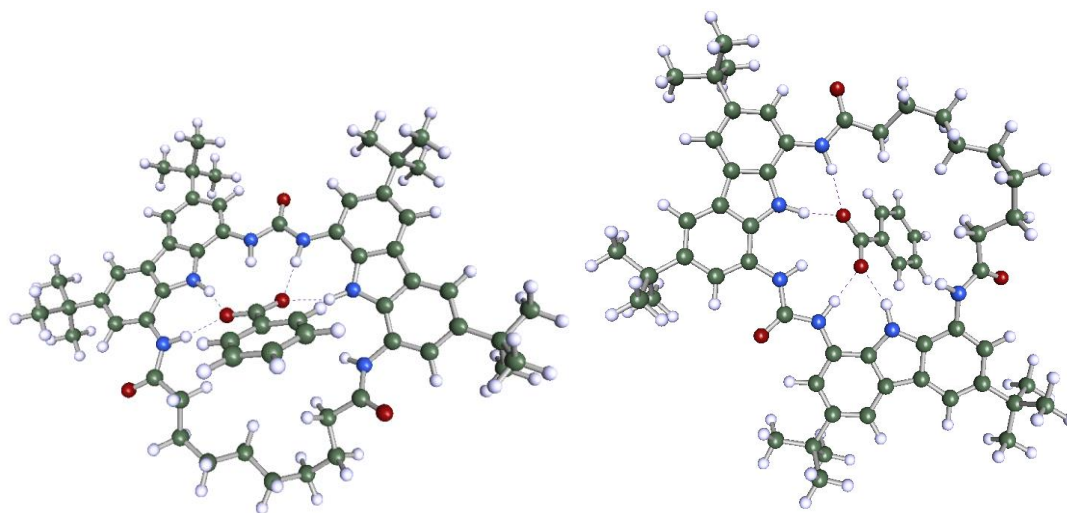
$G = -1810542.30752$ kcal/mol

Lowest energy conformation of receptor **MC008** with acetate anion



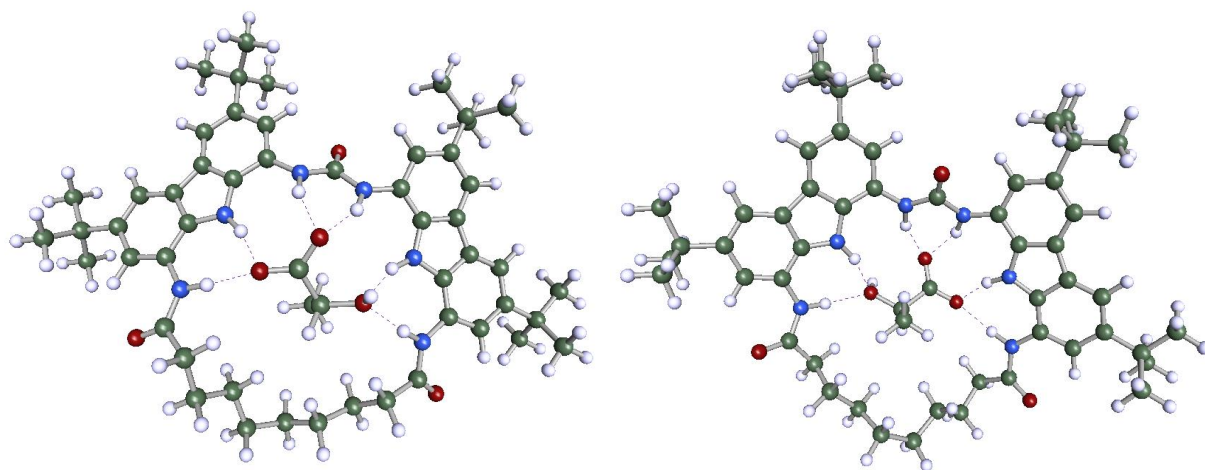
$G = -1736513.26323$ kcal/mol

Lowest energy conformation of receptor **MC008** with benzoate anion



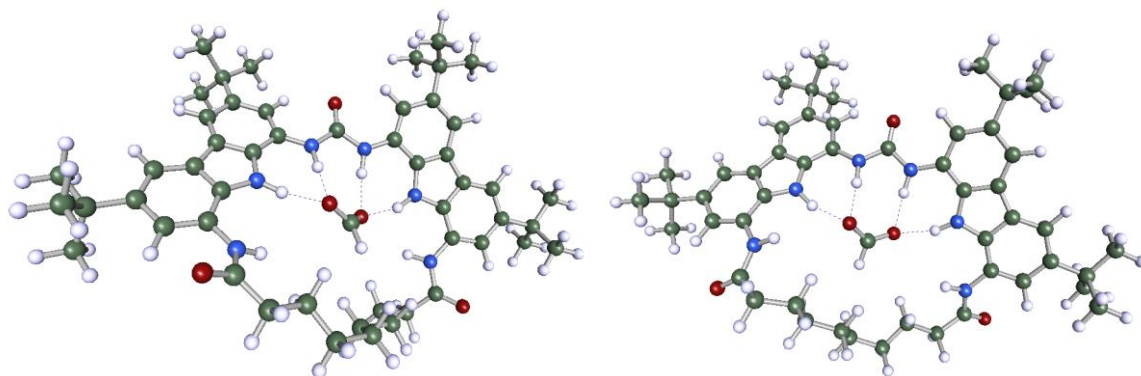
$G = -1856873.88068$ kcal/mol

Lowest energy conformation of receptor **MC008** with lactate anion



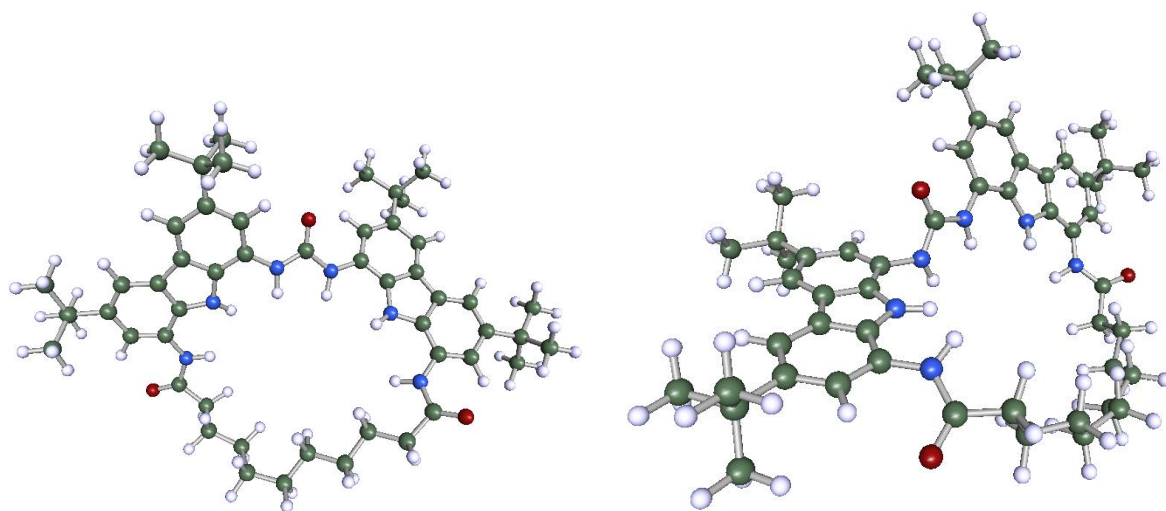
$G = -1808417.42913$ kcal/mol

Lowest energy conformation of receptor **MC008** with formate anion



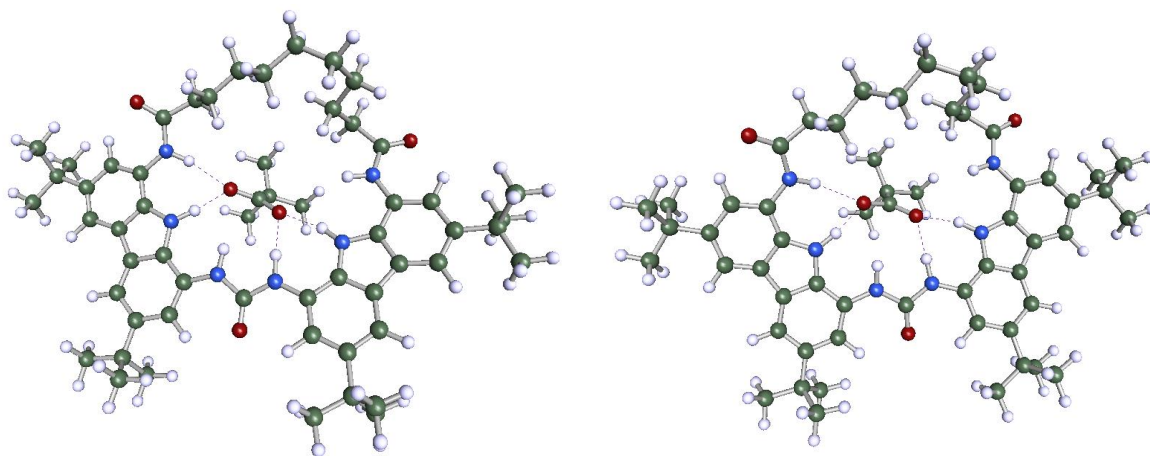
$G = -1711828.05606$ kcal/mol

Lowest energy conformation of receptor **MC009**



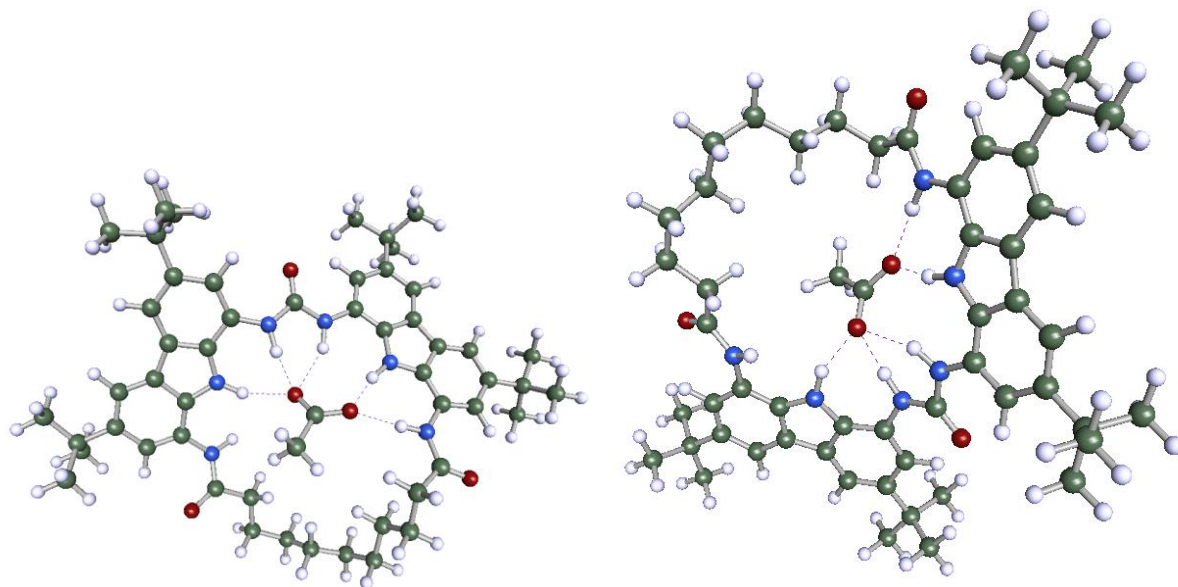
$G = -1617636.36547$ kcal/mol

Lowest energy conformation of receptor **MC009** with pivalate anion



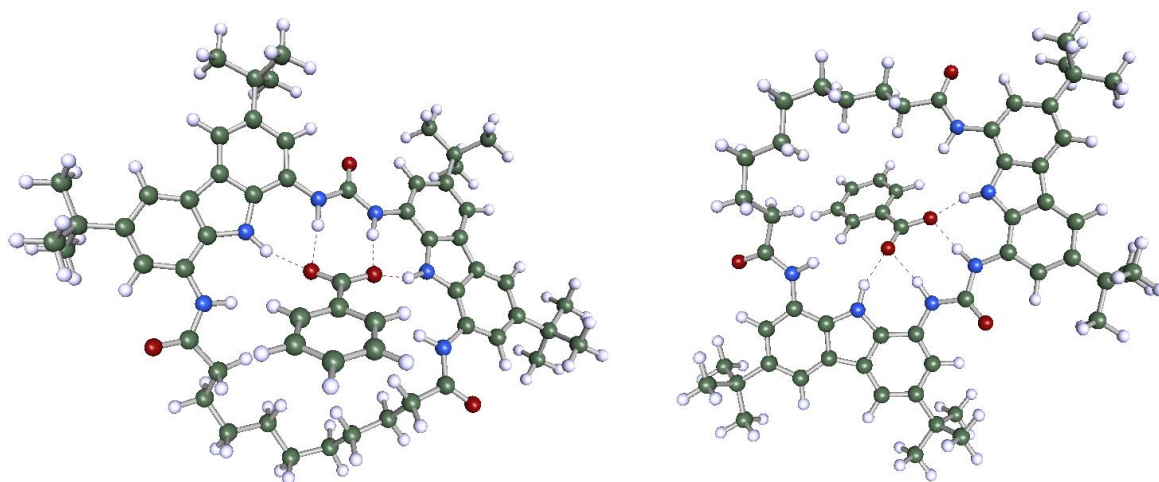
$G = -1835214.67834$ kcal/mol

Lowest energy conformation of receptor **MC009** with acetate anion



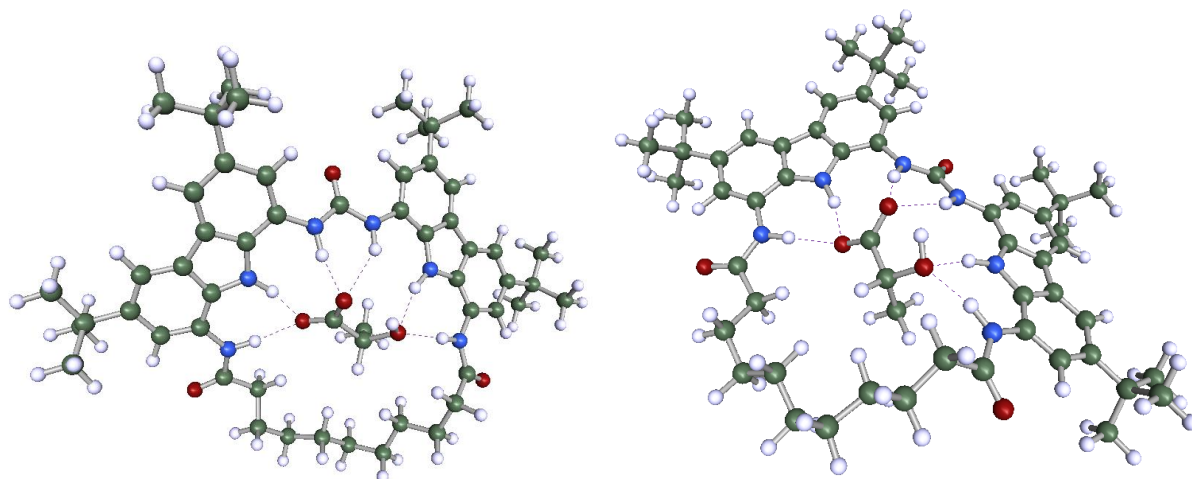
$G = -1761192.93428$ kcal/mol

Lowest energy conformation of receptor **MC009** with benzoate anion



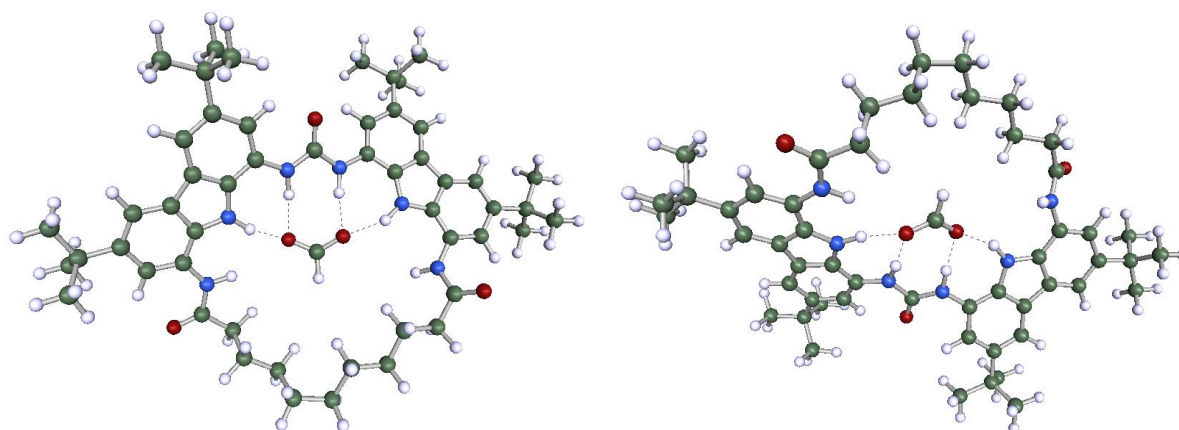
$G = -1881553.39445$ kcal/mol

Lowest energy conformation of receptor **MC009** with lactate anion



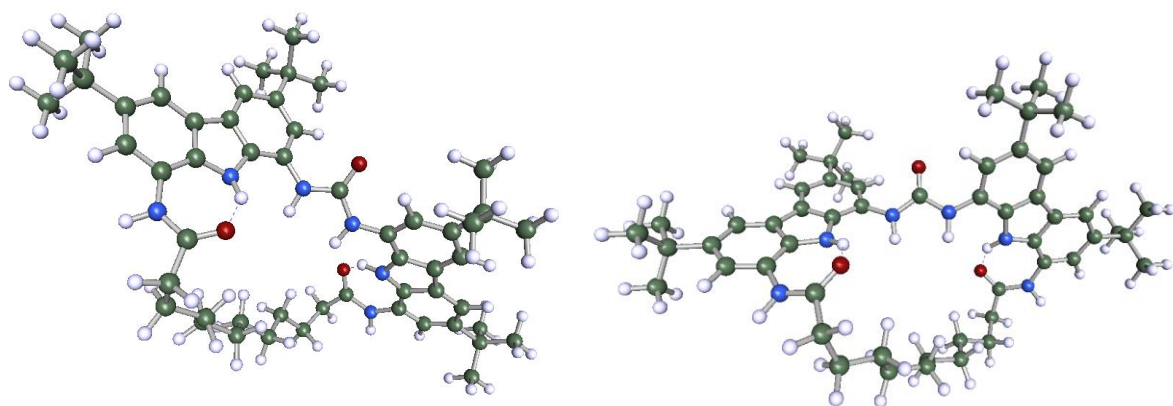
$G = -1833097.81583$ kcal/mol

Lowest energy conformation of receptor **MC009** with formate anion



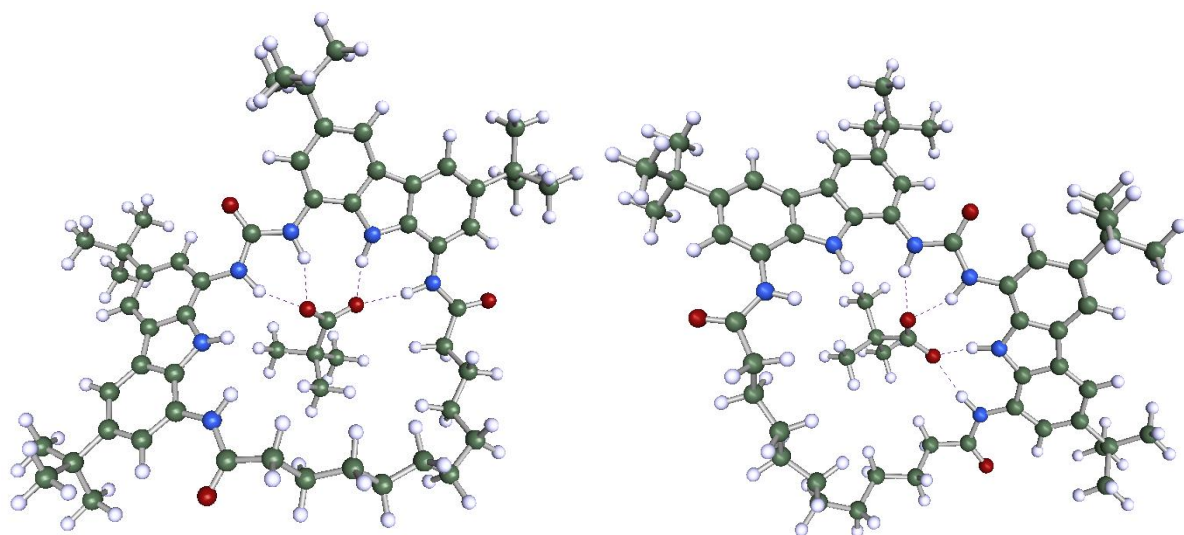
$G = -1736506.44416$ kcal/mol

Lowest energy conformation of receptor **MC010**



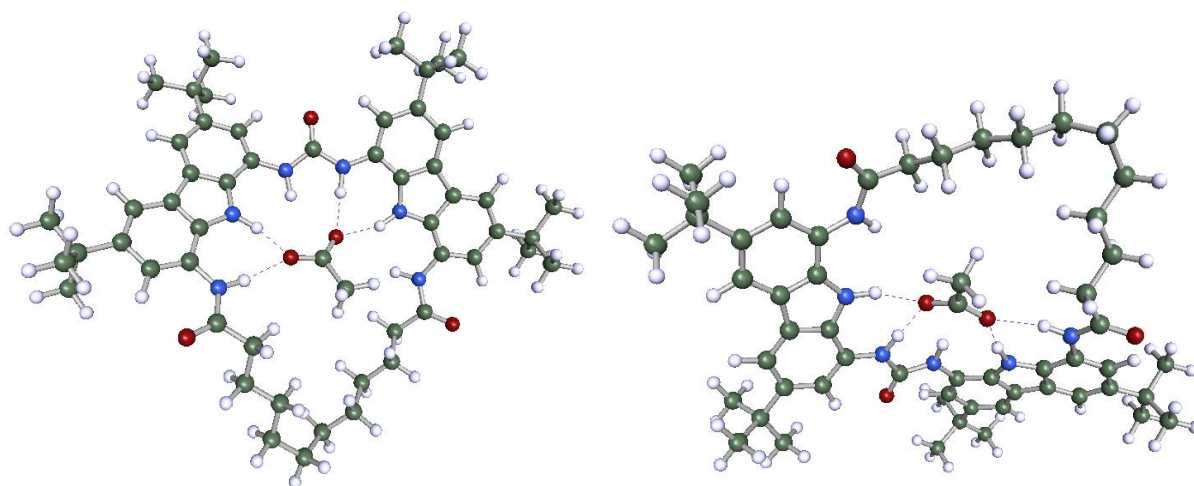
$G = -1642313.24091$ kcal/mol

Lowest energy conformation of receptor **MC010** with pivalate anion



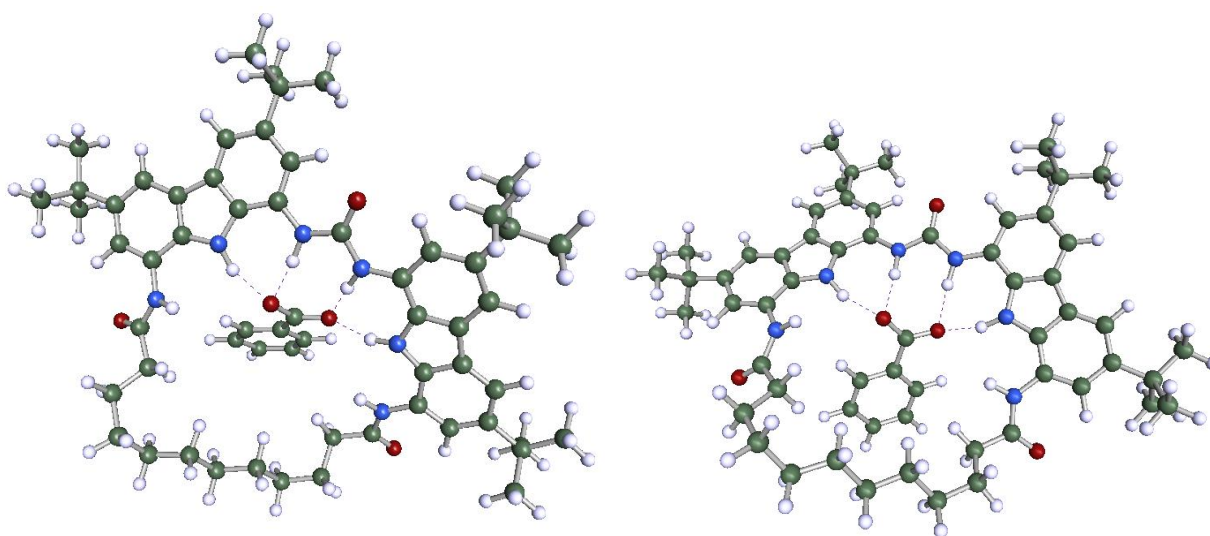
$G = -1859899.36917$ kcal/mol

Lowest energy conformation of receptor **MC010** with acetate anion



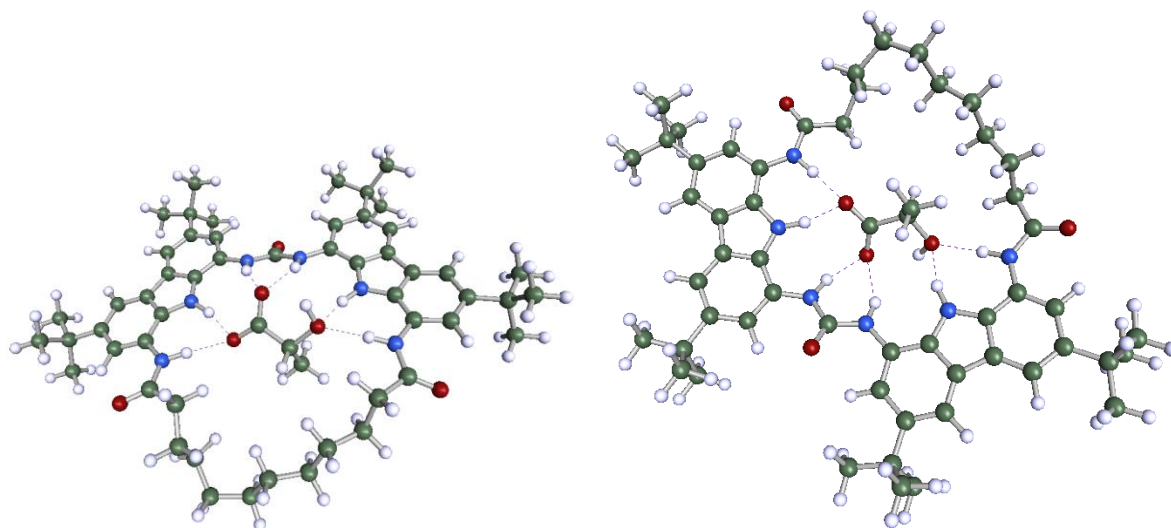
$G = -1785869.40606$ kcal/mol

Lowest energy conformation of receptor **MC010** with benzoate anion



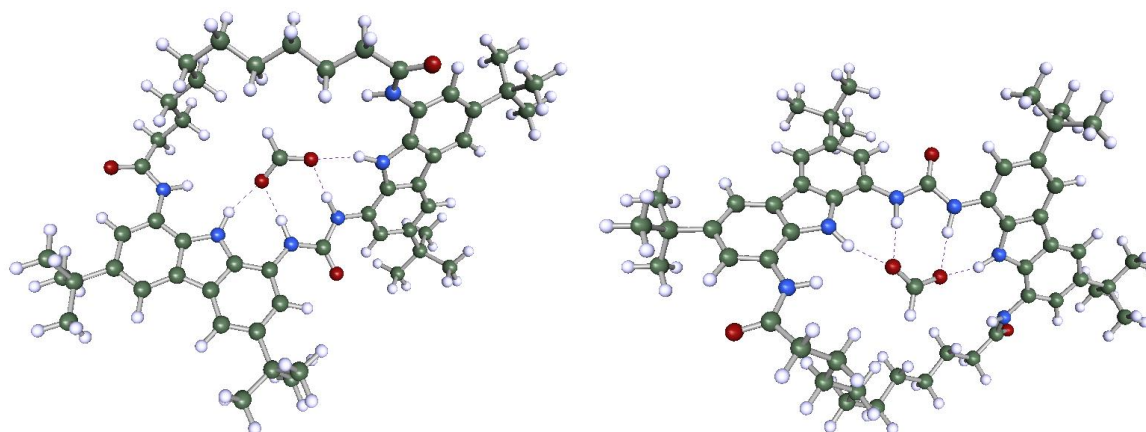
$G = -1906229.24005$ kcal/mol

Lowest energy conformation of receptor **MC010** with lactate anion



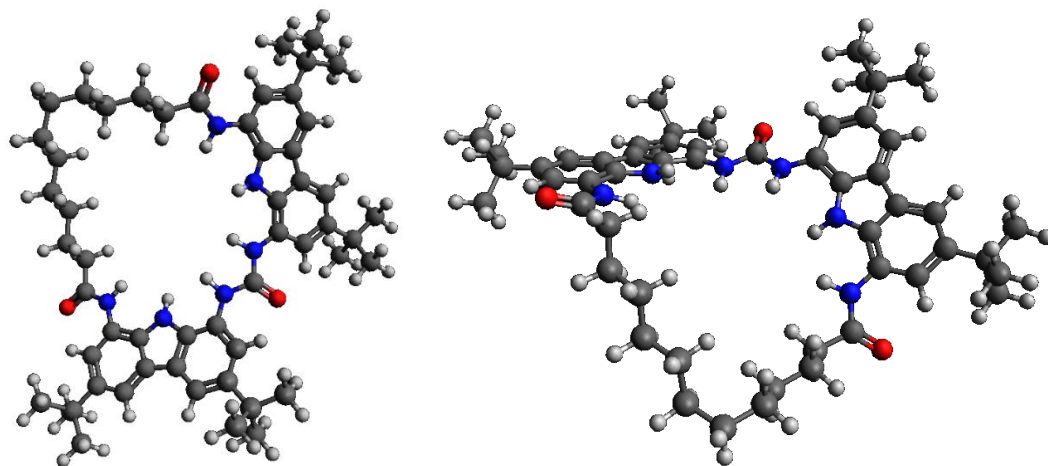
$G = -1857774.74552$ kcal/mol

Lowest energy conformation of receptor **MC010** with formate anion



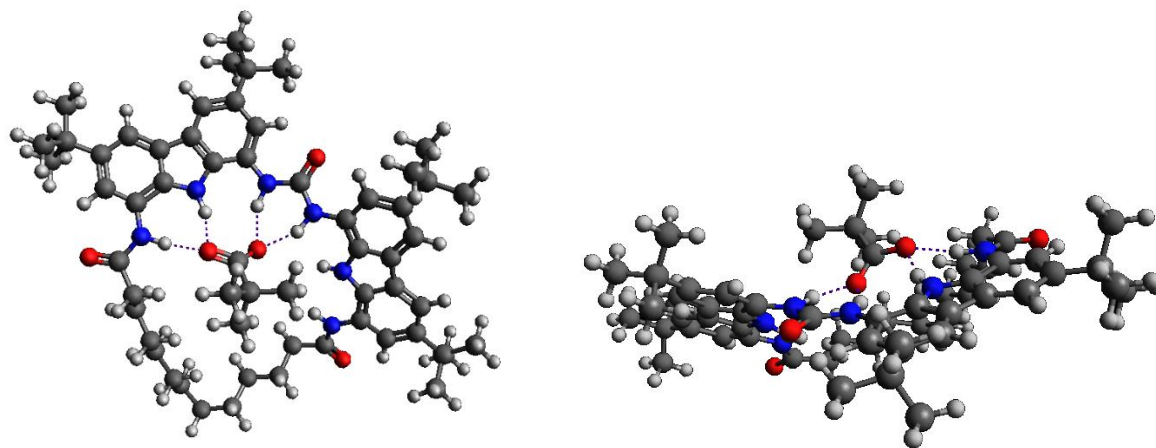
$G = -1761183.57068$ kcal/mol

Lowest energy conformation of receptor **MC011**



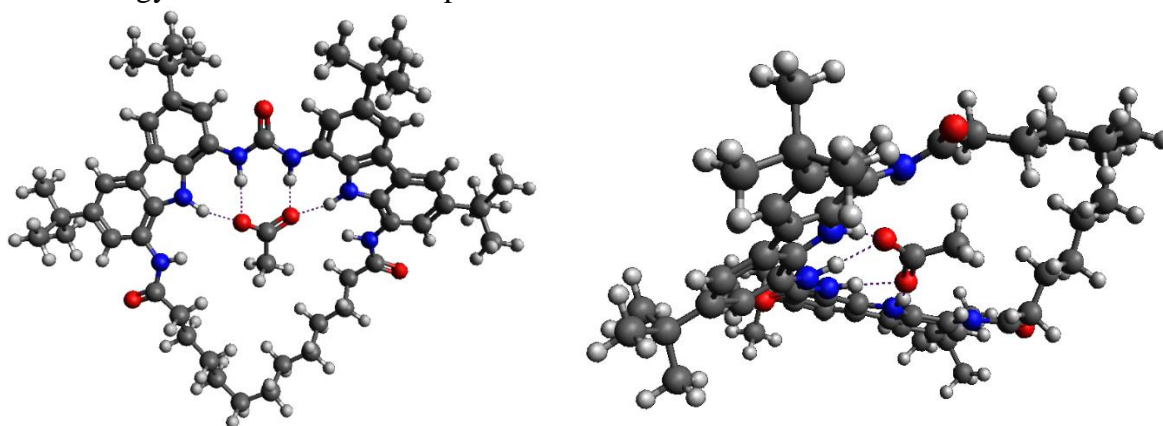
$G = -1666995.75494$ kcal/mol

Lowest energy conformation of receptor **MC011** with pivalate anion



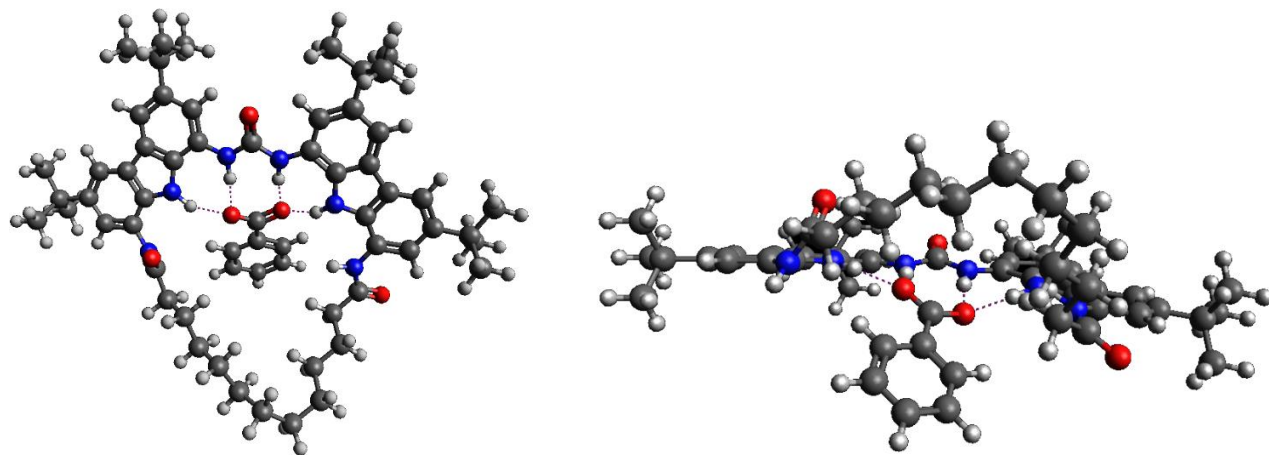
$G = -11884578.42968$ kcal/mol

Lowest energy conformation of receptor **MC011** with acetate anion



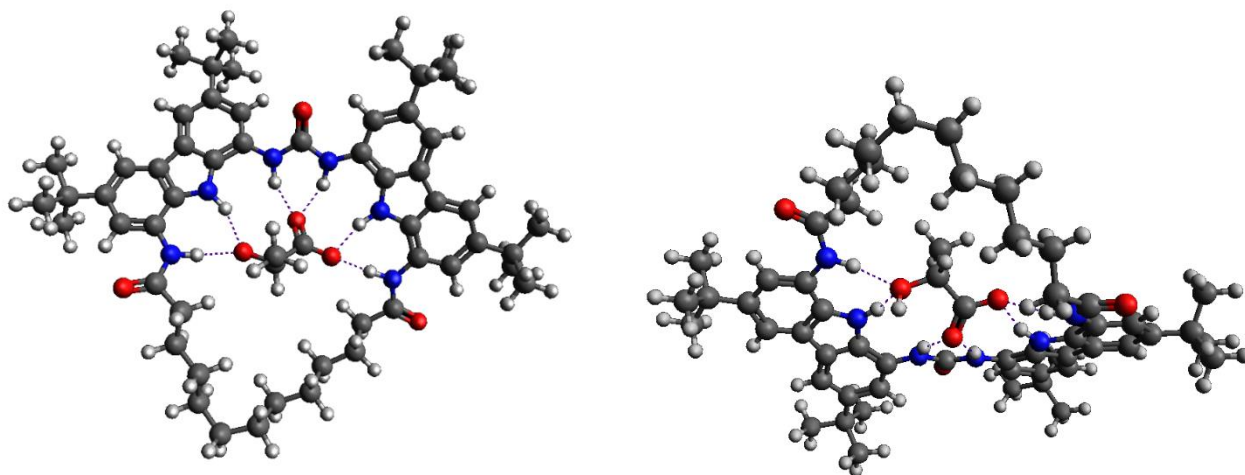
$G = -1810547.72110$ kcal/mol

Lowest energy conformation of receptor **MC011** with benzoate anion



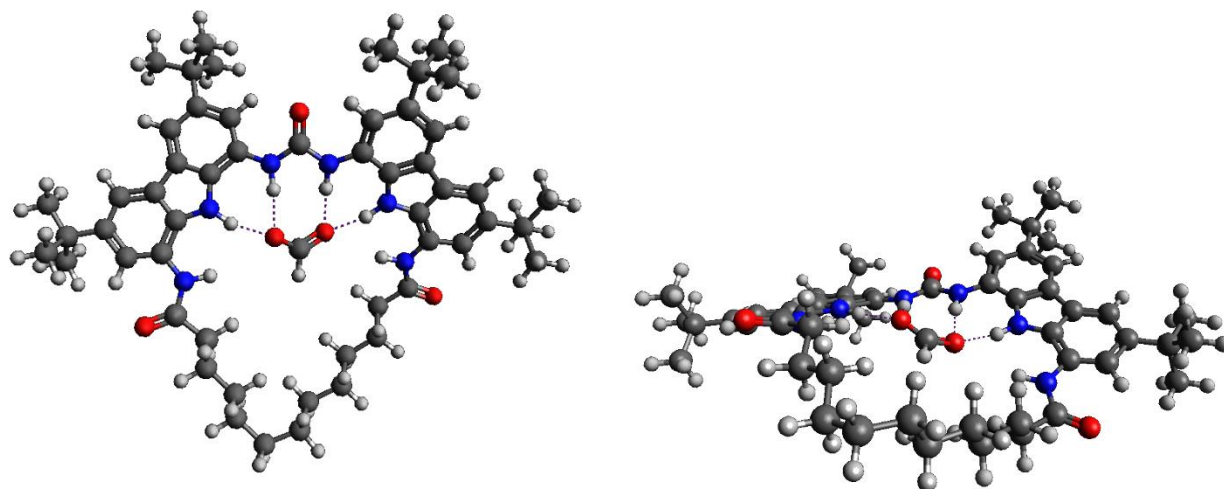
$G = -1930905.78073$ kcal/mol

Lowest energy conformation of receptor **MC011** with lactate anion



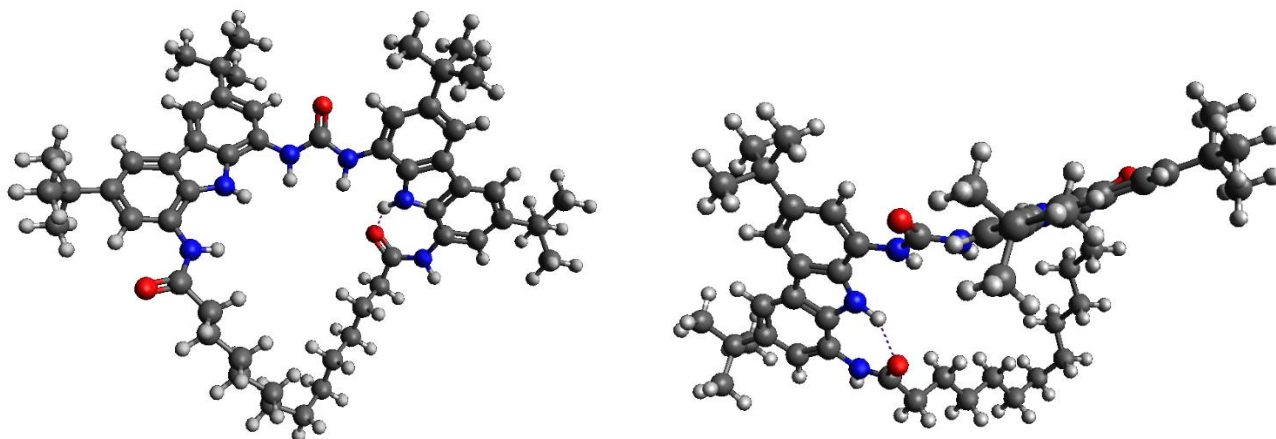
$G = -1882455.59817$ kcal/mol

Lowest energy conformation of receptor **MC011** with formate anion



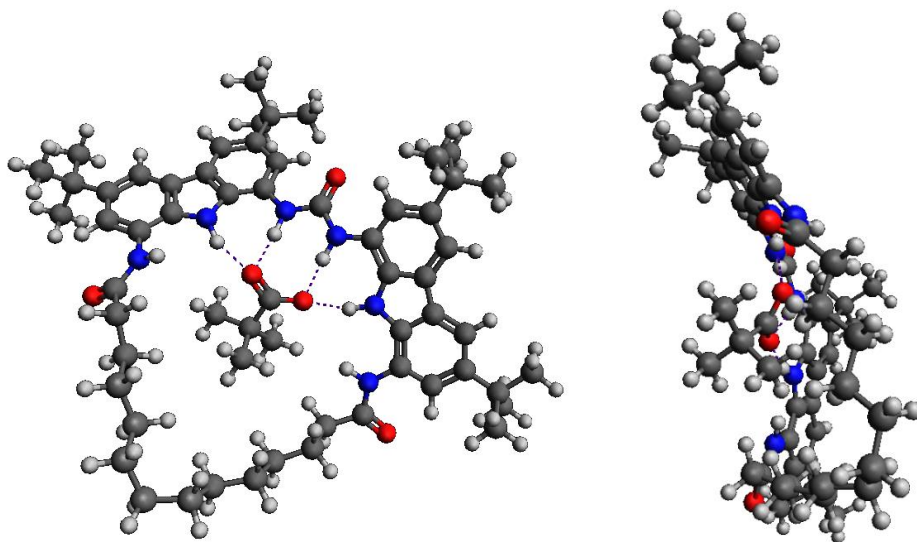
$G = -1785865.82166$ kcal/mol

Lowest energy conformation of receptor **MC012**



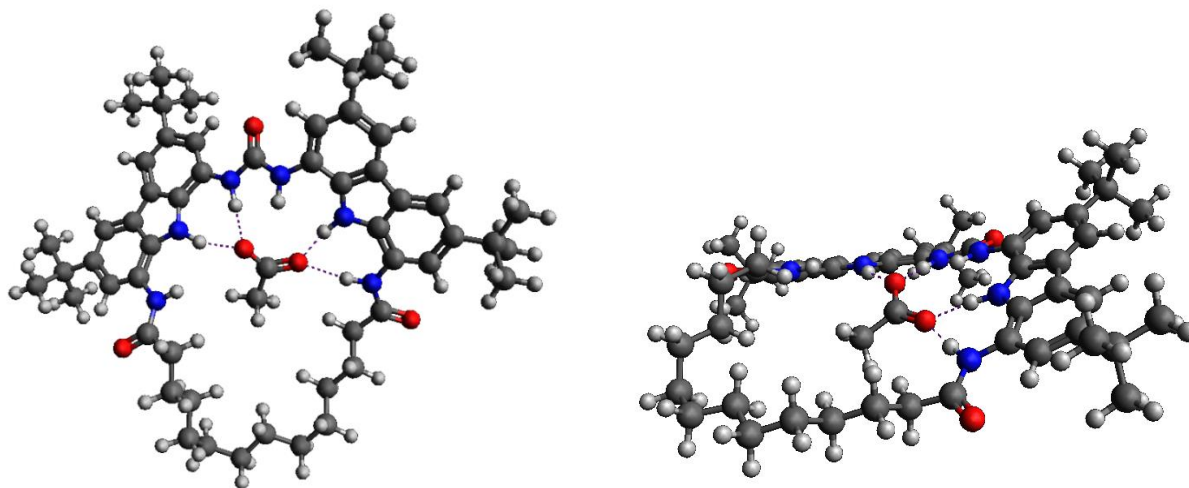
$G = -1691674.08605$ kcal/mol

Lowest energy conformation of receptor **MC012** with pivalate anion



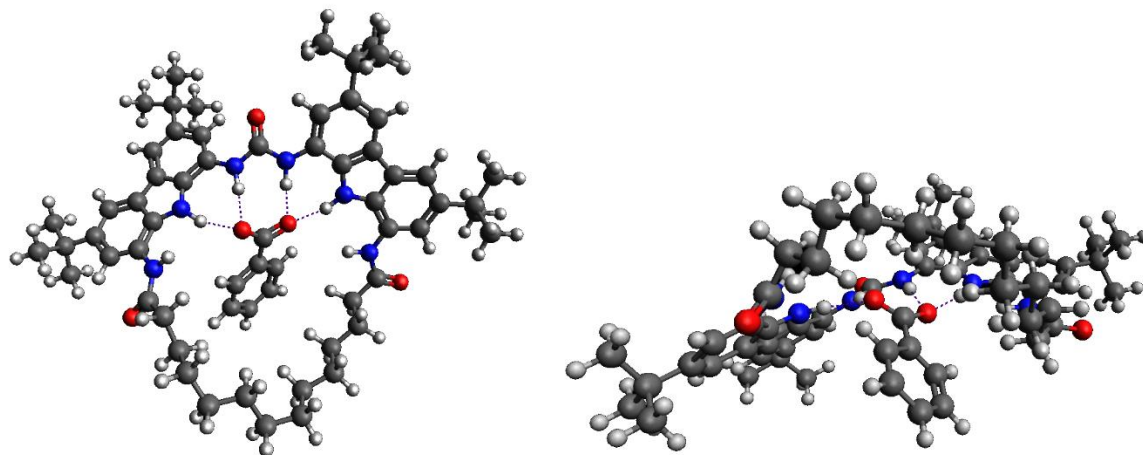
$G = -1909252.26264$ kcal/mol

Lowest energy conformation of receptor **MC012** with acetate anion



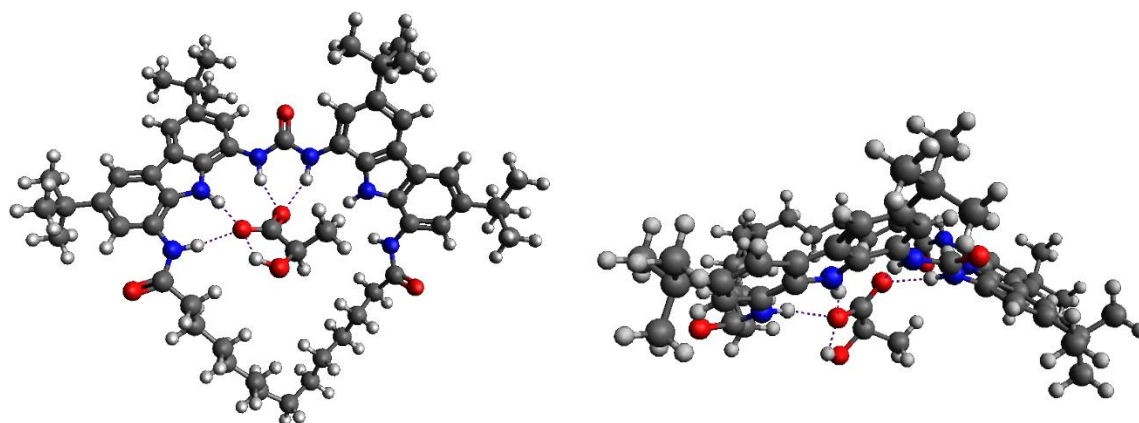
$G = -1835225.73860$ kcal/mol

Lowest energy conformation of receptor **MC012** with benzoate anion



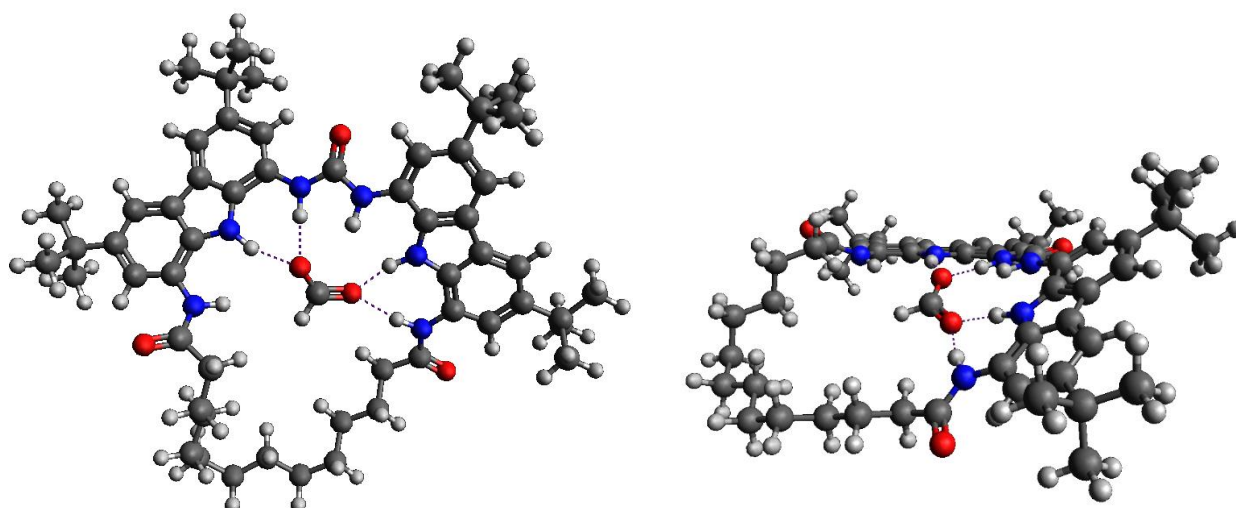
$G = -1955585.03038$ kcal/mol

Lowest energy conformation of receptor **MC012** with lactate anion



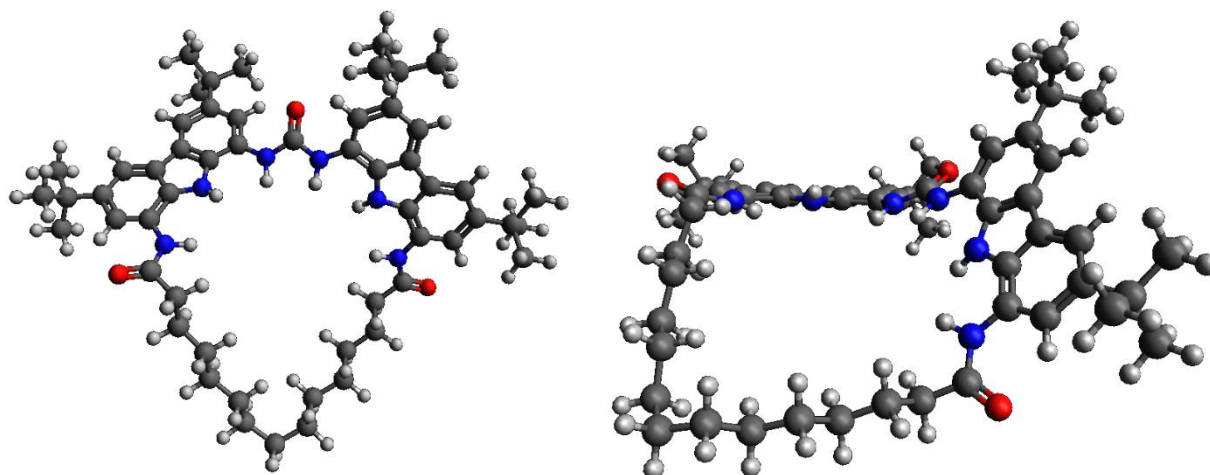
$G = -1907129.15110$ kcal/mol

Lowest energy conformation of receptor **MC012** with formate anion



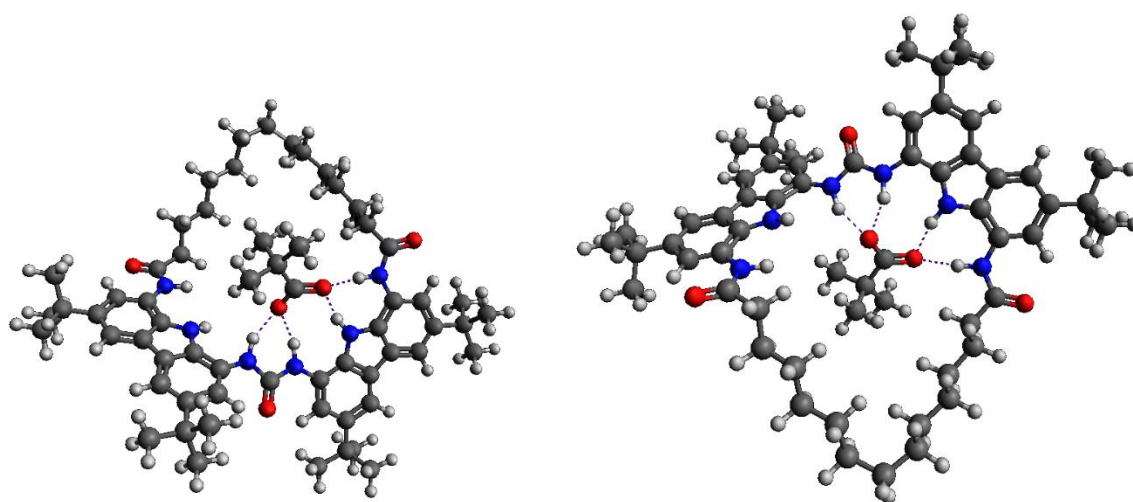
$G = -1810543.68556$ kcal/mol

Lowest energy conformation of receptor **MC013**



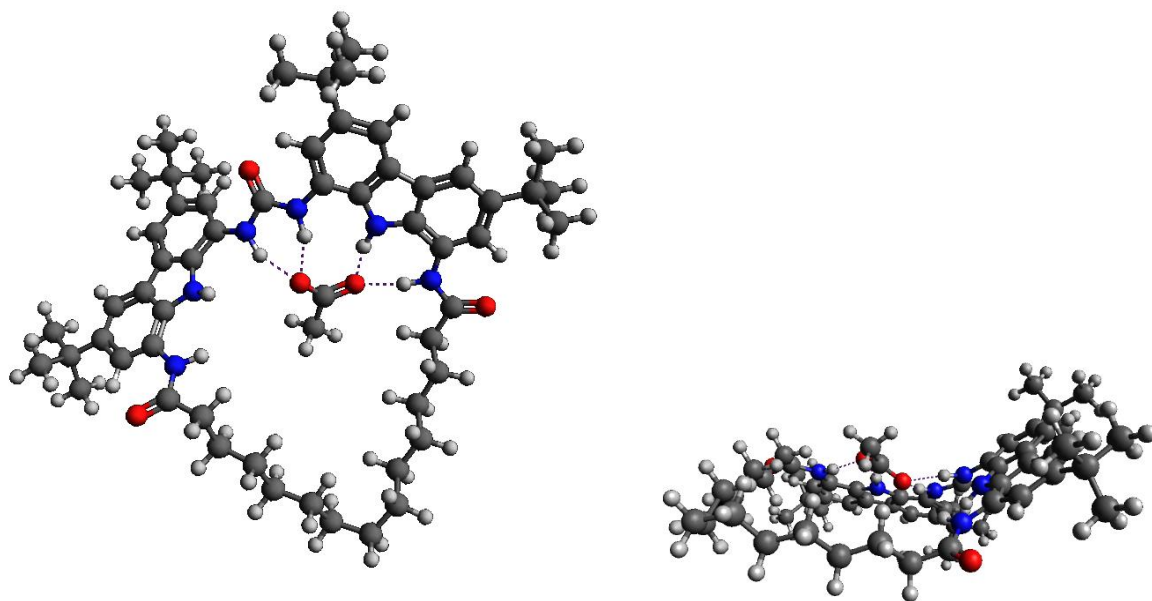
$G = -1716352.25505$ kcal/mol

Lowest energy conformation of receptor **MC013** with pivalate anion



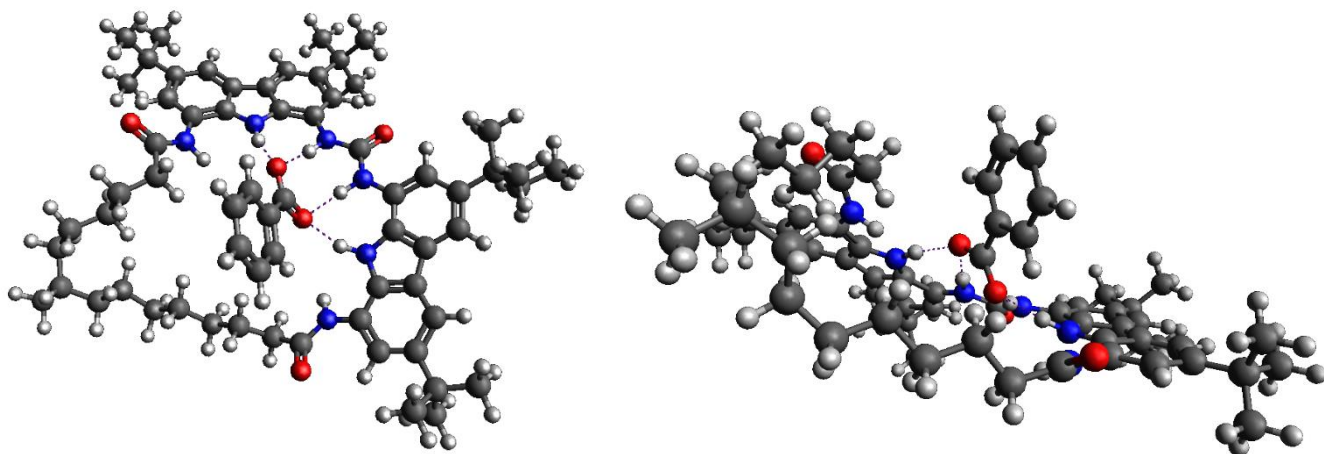
$G = -1933934.70969$ kcal/mol

Lowest energy conformation of receptor **MC013** with acetate anion



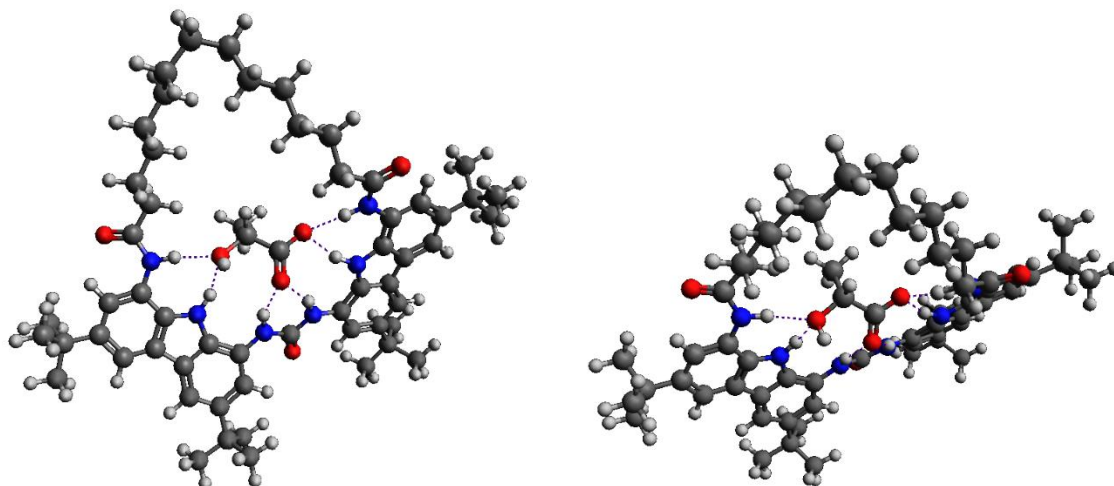
$G = -1859903.77757$ kcal/mol

Lowest energy conformation of receptor **MC013** with benzoate anion



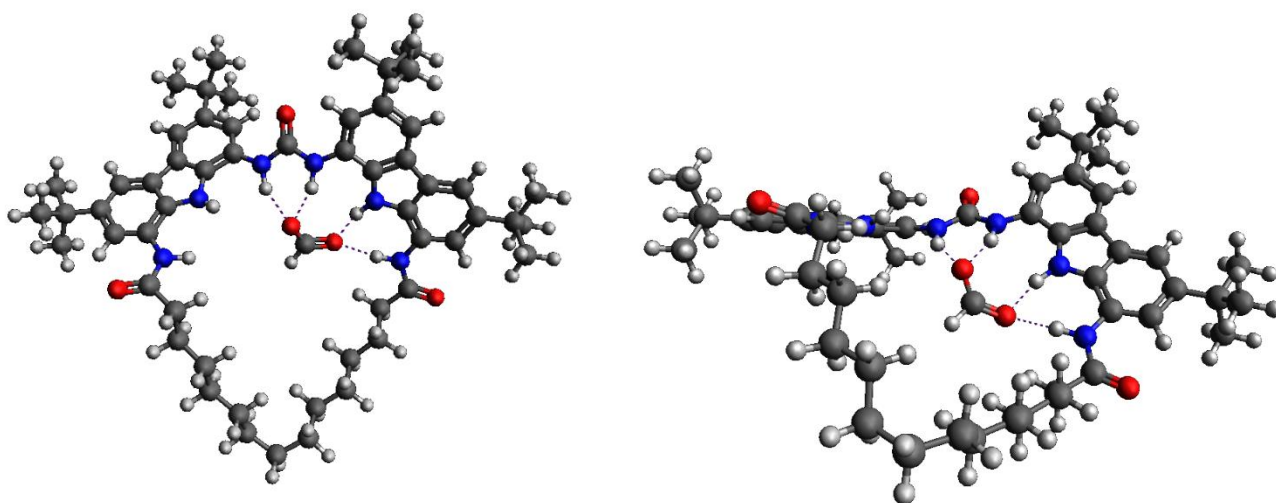
$G = -1980264.91917$ kcal/mol

Lowest energy conformation of receptor **MC013** with lactate anion



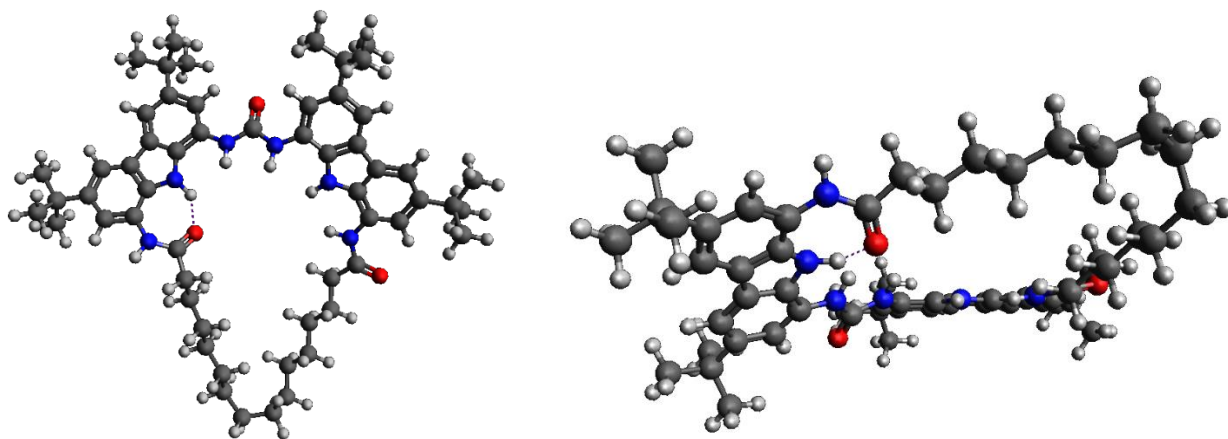
$G = -1931811.69804$ kcal/mol

Lowest energy conformation of receptor **MC013** with formate anion



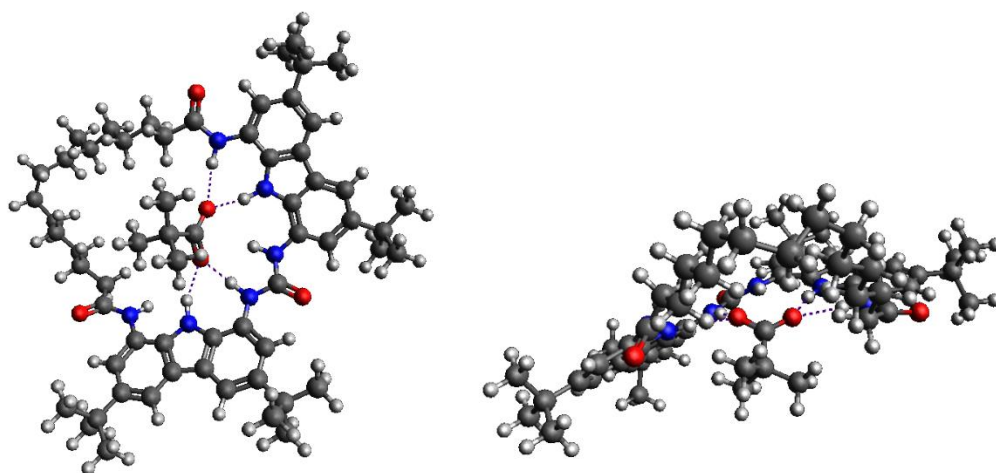
$G = -1835222.20163$ kcal/mol

Lowest energy conformation of receptor **MC014**



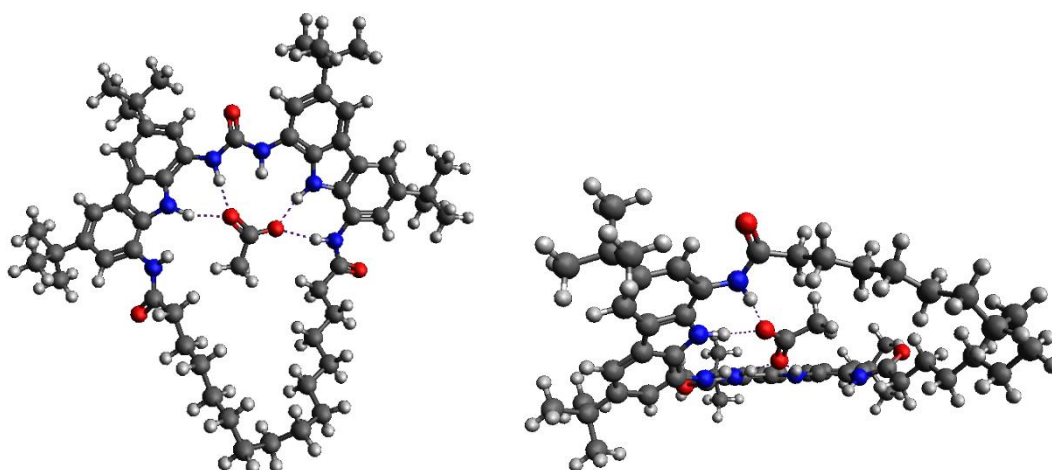
$G = -1741028.91954$ kcal/mol

Lowest energy conformation of receptor **MC014** with pivalate anion



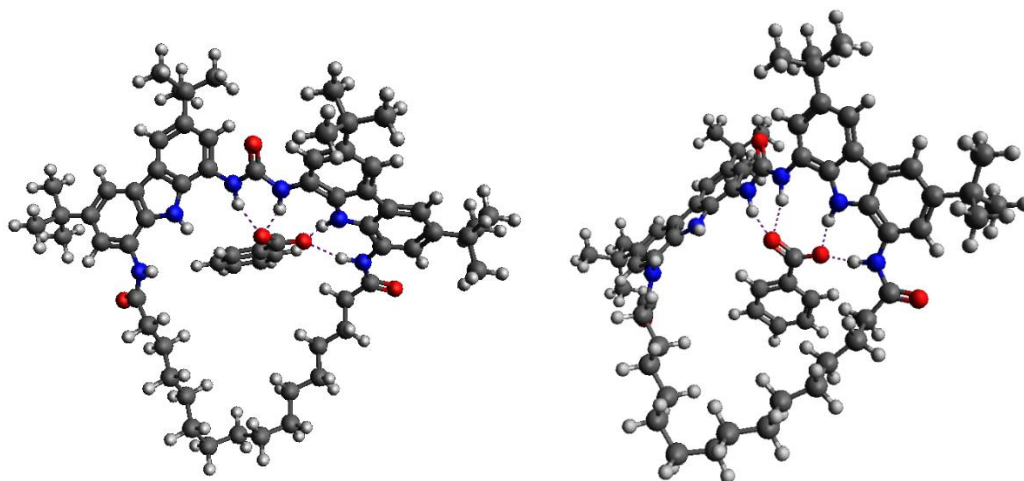
$G = -1958610.17256$ kcal/mol

Lowest energy conformation of receptor **MC014** with acetate anion



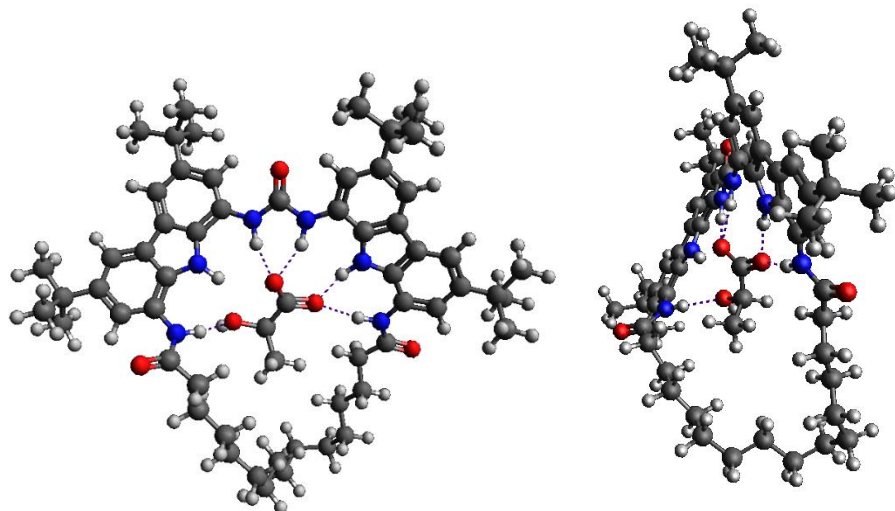
$G = -1884581.51422$ kcal/mol

Lowest energy conformation of receptor **MC014** with benzoate anion



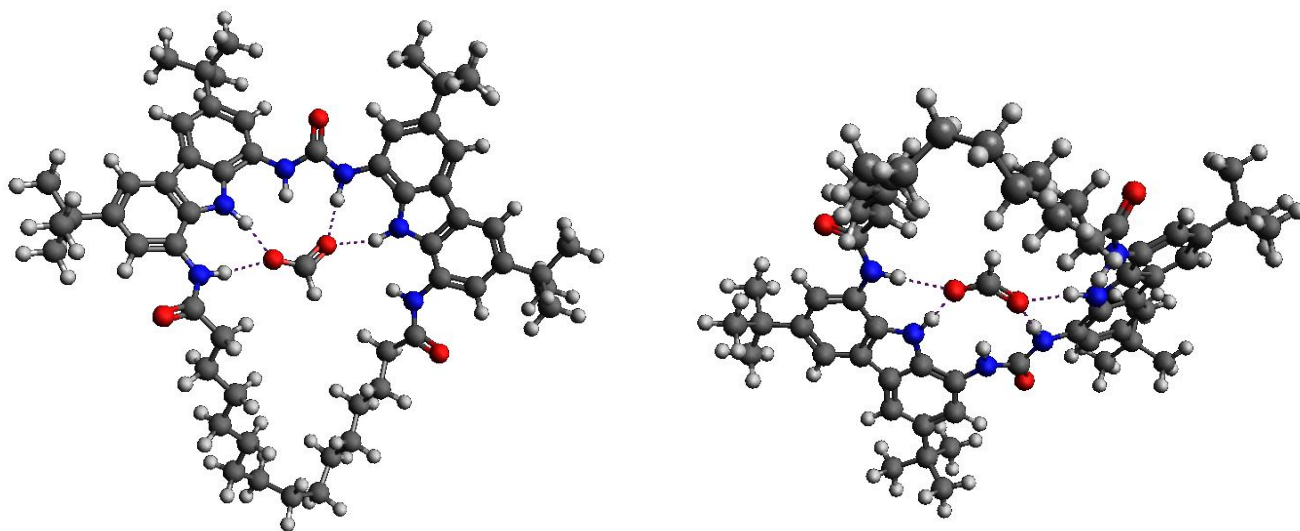
$G = -2004943.68243$ kcal/mol

Lowest energy conformation of receptor **MC014** with lactate anion



$G = -1956485.72757$ kcal/mol

Lowest energy conformation of receptor **MC014** with formate anion



$G = -1859899.50598$ kcal/mol